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WEED CONTROL

NOTES

TURNING POINT SURVEY OF WEED CONTROL AND PRODUCTION PRACTICES IN SUGARBEET IN MINNESOTA AND EASTERN NORTH DAKOTA IN 2016

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The second annual weed control and production practices live polling questionnaire was conducted using Turning Point Technology at the 2017 winter Sugarbeet Grower Seminars. Responses are based on production practices from the 2016 growing season. The survey focuses on responses from growers in attendance at the Fargo, Grafton, Wahpeton, ND, and Willmar, MN, Growers Seminars. Respondents from each seminar indicated the county in which the majority of their sugarbeet were produced (Tables 1, 2, 3, 4). Survey results represents approximately 158,272 acres reported by 235 respondents (Table 5) compared to 183,350 acres represented in 2016. The average sugarbeet acreage per respondent grown in 2016 was calculated from Table 5 at 673 acres, compared to 674 acres in 2015.

Survey participants were asked a series of questions regarding their production practices used in sugarbeet in 2016. Thirty-nine percent of respondents indicated wheat was the crop preceding sugarbeet (Table 6), 39% indicated corn, and 9% indicated soybean. Preceding crop varied dramatically by location with 82% of Fargo growers indicating wheat preceded sugarbeet and 74% of Willmar growers indicated corn as their preceding crop. Seventy-nine percent of growers attending the winter meetings used a nurse or cover crop in 2016 (Table 7), which increased from 72% in 2015. Cover crop species also varied widely by location with oat being used by 58% of growers at the Willmar meeting and no cover crop being used by the majority (38%) of growers at the Grafton meeting.

Growers indicated *Cercospora* Leaf Spot (CLS) was their most serious production problem in sugarbeet in 2016 (Table 8) with 57% of all respondents naming CLS compared to *Rhizoctonia* being named most serious by 35% of all participants in 2015. *Cercospora* was devastating to sugarbeet quality in 2016. Weather was the most serious problem for 23% of growers, mainly those in the northern valley, and weeds were named as most serious by 7% of responses.

Waterhemp was named as the most serious weed problem in sugarbeet in 2016 by 59% of respondents (Table 9) compared to 45% in 2015. Ten percent of respondents indicated common lambsquarters, 9% kochia, and 8% said common ragweed were their most serious weed problem. The increased presence of glyphosate-resistant waterhemp and common ragweed are likely the reason for these weeds being named as the worst weeds. Troublesome weeds varied by location with greater than 80% of Willmar and Wahpeton respondents indicating waterhemp was most problematic while kochia was the worst weed for respondents of the Grafton meeting with 38% of responses.

Respondents to the survey indicated making 0 to 5 glyphosate applications in their 2016 sugarbeet crop (Table 10) with a calculated average of 2.28 applications per acre. The calculated average in 2015 was 2.23 applications per acre.

Glyphosate was most commonly applied with a chloroacetamide herbicide postemergence (lay-by) in 2016 with 36% of responses indicating this herbicide combination was used (Table 11). Fifty-five percent and 42% of Wahpeton and Willmar respondents, respectfully, applied glyphosate with Outlook, S-metolachlor, or Warrant but only 26% and 0% of Fargo and Grafton respondents, respectfully, used this combination. Use of chloroacetamides with glyphosate track to areas where glyphosate-resistant waterhemp is common. Glyphosate alone was the second most common herbicide used in sugarbeet in 2016 with 31% of responses, followed by glyphosate plus a broadleaf herbicide for 21% of the responses. Satisfaction to weed control from glyphosate applied alone is shown in Table 12

and ranged from 15% of responses indicating excellent control to 6% of responses indicating poor weed control. The majority of responses, 42%, indicated glyphosate was still providing good weed control in sugarbeet in 2016.

Preplant incorporated (PPI) or preemergence (PRE) herbicides were applied by 48% of survey respondents in 2016 (Table 13). Less than 10% of Grafton survey participants applied a PPI or PRE herbicide, while 75% of Wahpeton survey participants did apply a PPI or PRE herbicide in sugarbeet in 2016. Once again, a likely reason for this variation is the increased presence of glyphosate-resistant waterhemp in the southern sugarbeet growing areas of the Red River Valley compared to the north end of the Valley. The most commonly used soil herbicide was S-metolachlor with 22% of all responses followed by ethofumesate with 13% of responses. Of the growers who indicated using a soil-applied herbicide, 77% indicated excellent to good weed control from that herbicide (calculated from Table 14).

The application of soil-residual herbicides applied 'lay-by' was implemented by 71% of those responding about their 2016 sugarbeet crop (Table 15). Outlook was the most commonly applied lay-by herbicide with 33% of responses. The majority of growers responding at the Willmar meeting indicated using Outlook (56% of responses), while S-metolachlor was more commonly applied by growers of the Fargo (40% of responses) and Wahpeton (46% of responses) meetings. Satisfaction of weed control from lay-by applications ranged from excellent to poor (Table 16). Of respondents indicating they applied a lay-by herbicide, 78% indicated excellent or good weed control (calculated from Table 16).

Fifty-four percent of survey responses indicated using some form of mechanical weed control or hand labor in 2016 (Table 17). Of the responses given, 32% indicated at least some hand-weeding, 18% used row-cultivation, and 1% indicated using a rotary hoe for weed control in sugarbeet. Nineteen percent reported row-crop cultivation on less than ten percent of their acres (Table 18). One cultivation pass was reported by 94% of respondents who reported cultivating (calculated from Table 19). Respondents who cultivated generally reported good to fair weed control from the cultivation (Table 20).

Hand-weeding the 2016 sugarbeet crop was reported by 47% of respondents (Table 21). Most respondents who hand-weeded indicated less than 10% of their acres were hand-weeded. Less respondents indicated hand-weeding at the Grafton meeting, while more than half the participants of the Fargo and Wahpeton meetings reported some hand weeding. The cost of hand-weeding on a per acre basis ranged from less than \$10 to greater than \$40 per acre (Table 22). For growers who reported hand-weeding, 61% reported 'excellent' or 'good' hand-weeding control (Table 23).

Table 1. 2017 Fargo Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Barnes	3	9
Cass	7	21
Clay	11	32
Norman ¹	8	24
Richland	1	3
Trail	3	9
Wilkin ²	1	3
Total	34	100

¹Includes Mahnommen County

²Includes Otter Tail County

Table 2. 2017 Grafton Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Grand Forks	1	2
Kittson	4	7
Marshall	5	9
Pembina	19	35
Polk	1	2
Walsh	23	43
Other	1	2
Total	54	100

Table 3. 2017 Wahpeton Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Cass	2	4
Clay	3	7
Grant	5	11
Otter Tail	1	2
Richland	7	16
Stevens	1	2
Traverse	5	11
Wilkin	21	47
Total	45	100

Table 4. 2017 Willmar Grower Seminar - Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Chippewa	36	33
Kandiyohi	17	16
Pope	0	0
Redwood	5	5
Renville	31	28
Stearns	3	3
Stevens	1	1
Swift	9	8
Other	7	6
Total	109	100

Table 5. Total sugarbeet acreage operated by respondents in 2016.

Location	Responses	Acres of sugarbeet									
		<99	100-199	200-299	300-399	400-599	600-799	800-999	1000-1499	1500-1999	2000+
-----% of responses-----											
Grafton	54	6	15	11	9	17	9	11	9	2	9
Fargo	33	3	0	15	18	18	6	9	12	6	12
Wahpeton	42	2	7	2	10	33	17	12	10	5	2
Willmar	107	7	15	15	6	22	10	3	14	2	7
Total	235	6	11	12	9	22	11	7	12	3	7

Table 6. Crop grown in 2015 that preceded sugarbeet in 2016.

Location	Responses	Previous Crop								
		Barley	Canola	Corn	Dry Bean	Potato	Soybean	Wheat	Fallow	Other
		-----% of responses-----								
Grafton	53	2	0	0	9	11	2	74	0	2
Fargo	33	6	0	3	0	0	6	82	0	3
Wahpeton	41	0	2	24	0	0	12	61	0	0
Willmar	108	1	0	74	1	0	12	0	0	12
Total	235	2	<1	39	3	3	9	39	0	6

Table 7. Nurse or cover crop used in sugarbeet in 2016.

Location	Responses	Barley	Oat	Rye	Wheat	Other ¹	None
Grafton	52	21	14	0	27	0	38
Fargo	33	42	3	0	12	0	42
Wahpeton	42	45	2	0	40	0	12
Willmar	106	0	58	1	30	2	10
Total	233	19	30	<1	29	1	21

¹Includes Mustard and 'Other'**Table 8. Most serious production problem in sugarbeet in 2016.**

Location	No. of Responses	CLS ¹	Rhizomania	Aph ²	Rhizoctonia	Fusarium	Weeds	Root		Emergence/Stand
								Maggot	Weather	
		-----% of responses-----								
Grafton	56	4	0	14	9	0	0	2	71	0
Fargo	36	44	0	6	8	0	19	0	11	11
Wahpeton	43	84	2	0	5	0	9	0	0	0
Willmar	106	79	2	0	2	0	6	0	10	1
Total	241	57	1	4	5	0	7	<1	23	2

¹Cercospora Leaf Spot²Aphanomyces**Table 9. Most serious weed problem in sugarbeet in 2016.**

Location	Responses	biww ¹	colq	cora	Foxtail				Smartweed	RR	
					spp.	kochia	gira	rrpw		Canola	wahe
		-----% of responses-----									
Grafton	53	6	27	10	0	38	0	12	0	6	2
Fargo	35	0	6	23	0	3	6	6	3	8	46
Wahpeton	43	0	2	7	0	0	2	5	0	2	81
Willmar	104	0	6	2	0	0	4	3	0	2	84
Total	234	1	10	8	0	9	3	6	<1	4	59

¹biww=biennial wormwood, colq=common lambsquarters, cora=common ragweed, gira=giant ragweed, rrpw=redroot pigweed,

wahe=waterhemp

Table 10. Average number of glyphosate applications per acre in sugarbeet during 2016 season.

Location	Responses	0	1	2	3	4	5
Grafton	51	4	22	57	16	0	2
Fargo	35	0	14	63	23	0	0
Wahpeton	46	0	4	39	50	7	0
Willmar	106	1	11	42	40	5	1
Total	238	1	13	48	34	3	1

Table 11. Herbicides used in a weed control systems approach in sugarbeet in 2016.

Location	Responses	Glyphosate Application Tank-Mixes					
		Gly Alone	Gly+Lay-by	Gly+Broadleaf	Gly+Grass	Other	None Used
		-----% of responses-----					
Grafton	51	80	0	16	0	0	4
Fargo	43	30	26	37	7	0	0
Wahpeton	58	17	55	22	5	0	0
Willmar	187	21	42	19	14	3	1
Total	339	31	36	21	10	1	1

Table 12. Satisfaction in weed control from glyphosate applied in sugarbeet in 2016.

Location	Responses	Satisfaction of Weed Control from Glyphosate					
		Excellent	Good	Fair	Poor	Unsure	Not Used Alone
		-----% of responses-----					
Grafton	49	47	49	2	0	2	0
Fargo	34	6	65	21	3	0	6
Wahpeton	46	2	35	41	4	0	17
Willmar	104	9	35	29	10	2	16
Total	233	15	42	24	6	1	12

Table 13. Preplant incorporated and preemergence herbicides used in sugarbeet in 2016.

Location	Responses	PPI or PRE Herbicides Applied					None
		S-metolachlor	ethofumesate	Ro-Neet SB	+ethofumesate	Other	
		-----% of responses-----					
Grafton	50	0	0	0	2	4	94
Fargo	35	37	0	0	3	3	57
Wahpeton	44	43	11	2	16	2	25
Willmar	108	19	24	0	6	9	42
Total	237	22	13	<1	7	6	52

Table 14. Satisfaction in weed control from preplant incorporated and preemergence herbicides in 2016.

Location	Responses	PPI or PRE Weed Control Satisfaction					
		Excellent	Good	Fair	Poor	Unsure	None Used
		-----% of responses-----					
Grafton	54	13	2	0	2	0	83
Fargo	34	21	21	12	3	0	44
Wahpeton	42	12	50	14	2	0	21
Willmar	105	17	30	10	3	1	39
Total	235	16	25	9	2	<1	47

Table 15. Soil-residual herbicides applied early postemergence (lay-by) in sugarbeet in 2016.

Location	Responses	Lay-by Herbicides Applied					None
		S-metolachlor	Ethofumesate	Outlook	Warrant	Other	
		-----% of responses-----					
Grafton	53	0	0	0	0	2	98
Fargo	35	40	3	9	0	6	43
Wahpeton	48	46	17	19	6	0	13
Willmar	148	8	7	56	20	1	7
Total	284	17	7	33	12	2	29

Table 16. Satisfaction of weed control from soil-residual herbicides applied early postemergence (lay-by) in sugarbeet in 2016.

Location	Responses	Lay-by Weed Control Satisfaction					None Used
		Excellent	Good	Fair	Poor	Unsure	
		-----% of responses-----					
Grafton	52	0	0	0	15	0	85
Fargo	36	14	33	6	3	0	44
Wahpeton	42	10	60	19	2	0	10
Willmar	108	32	48	10	1	1	7
Total	238	18	37	9	5	<1	30

Table 17. Mechanical weed control methods used in sugarbeet in 2016.

Location	Responses	Rotary Hoe	Row-Cultivation	Hand-Weeded	Other	None
		-----% of responses-----				
Grafton	51	2	2	12	4	80
Fargo	37	0	8	46	0	46
Wahpeton	48	2	4	23	4	67
Willmar	130	1	32	40	3	25
Total	266	1	18	32	3	46

Table 18. Percent of sugarbeet acres row-crop cultivated in 2016.

Location	Responses	% Acres Row-Cultivated				
		0	< 10	10-50	51-100	>100
-----% of responses-----						
Grafton	51	59	29	8	2	2
Fargo	35	74	23	0	3	0
Wahpeton	46	70	22	9	0	0
Willmar	103	48	12	8	7	26
Total	235	58	19	7	4	12

Table 19. Number of row-crop cultivation passes in sugarbeet in 2016.

Location	Responses	1	2	3	4	No Row-Cultivation
		-----% of responses-----				
Grafton	53	32	0	0	2	66
Fargo	34	24	0	0	0	76
Wahpeton	44	16	5	0	0	80
Willmar	105	38	4	0	0	58
Total	236	31	3	0	<1	67

Table 20. Satisfaction of weed control from row-crop cultivation in sugarbeet in 2016.

Location	Responses	Excellent	Good	Fair	Poor	Unsure	No Row-Cultivation
		-----% of responses-----					
Grafton	48	6	0	4	15	8	67
Fargo	35	0	9	17	0	6	69
Wahpeton	44	2	0	20	0	2	75
Willmar	105	3	16	18	2	4	57
Total	232	3	9	16	4	5	64

Table 21. Percent of sugarbeet acres hand-weeded in 2016.

Location	Responses	% Acres Hand-Weeded				
		0	< 10	10-50	51-100	>100
		-----% of responses-----				
Grafton	51	71	12	0	0	18
Fargo	36	42	50	8	0	0
Wahpeton	45	67	27	2	2	2
Willmar	103	43	30	19	4	4
Total	235	53	29	10	2	6

Table 22. Cost per acre for hand-weeding for hand weeding sugarbeet in 2016.

Location	Responses	Cost of Hand-Weeding per Acre					No Hand-Weeding
		<\$9.99	\$10-\$19.99	\$20-\$29.99	\$30-\$39.99	\$40+	
		-----% of responses-----					
Grafton	51	12	0	0	2	14	73
Fargo	35	46	6	0	0	6	43
Wahpeton	43	12	9	5	0	5	70
Willmar	105	17	29	6	3	2	44
Total	234	19	15	3	2	6	55

Table 23. Satisfaction of weed control from hand-weeding sugarbeet in 2016.

Location	Responses	Excellent	Good	Fair	Poor	Unsure	No Hand-Weeding
Grafton	50	4	2	6	16	0	72
Fargo	35	31	11	9	9	0	40
Wahpeton	44	9	14	2	2	0	73
Willmar	103	6	31	17	4	1	41
Total	232	10	19	11	7	<1	53

WEED CONTROL FROM ETHOFUMESATE APPLIED POSTEMERGENCE IN SUGARBEET

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SUMMARY

Ethofumesate applied postemergence (POST) twice at rates ranging from 12 to 64 fl oz/A suppressed but did not control lambsquarters and redroot pigweed. Ethofumesate POST is not an effective lambsquarters or pigweed herbicide and cannot be considered a second mode of action for control.

Ethofumesate alone or ethofumesate plus glyphosate improved waterhemp control compared to glyphosate alone. Control might be related to timing of waterhemp germination and emergence compared to lambsquarters or redroot pigweed.

Ethofumesate applied twice at rates ranging from 12 to 64 fl oz/A alone or with glyphosate at 28 fl oz/A caused only minor sugarbeet injury.

INTRODUCTION

Ethofumesate is a time-proven herbicide for grass and small-seeded broadleaf weed control in sugarbeet. Field research from Kansas and Colorado in 1970 indicated 'NC 8438' (ethofumesate) provided greater than 90% green foxtail, foxtail millet, and barnyardgrass control and near 90% redroot pigweed control (Sullivan and Fagala, 1970). Ethofumesate is soil-applied at field use rates up to 7.5 pt/A or applied postemergence up to 12 fl oz/A. Ethofumesate is absorbed by emerging shoots and roots and is translocated to the shoots where it is believed to interfere with lipid biosynthesis (Eshel et al., 1978, Abulnaja et al., 1992). Ethofumesate is sold in the United States using the trade names 'Nortron' by Bayer CropScience, 'Ethotron SC' by UPI, and 'Ethofumesate 4SC' by Willowood USA. Willowood USA is collaborating with the Beet Sugar Development Foundation to develop a new label to expand Ethofumesate 4SC postemergence use rates from 0.8 to 8 pt/A to sugarbeet having greater than two true leaves. Ethofumesate applied in combination with glyphosate may provide an effective second mode of action to complement glyphosate, especially for difficult to control broadleaf weeds in sugarbeet including common lambsquarters, kochia, waterhemp, and common ragweed. However, little is known about postemergence broadleaf weed control from ethofumesate, especially at rates greater than 12 fl oz/A.

Probe experiments were conducted in 2017 to evaluate weed efficacy and sugarbeet safety from single or multiple ethofumesate applications alone or with glyphosate applied postemergence. These probe experiments will serve as a basis for Mrs. Alexa Lystad's MS degree research and will provide recommendations for use of ethofumesate for weed control in sugarbeet grower fields in 2018. The objectives of this research were to determine: a) is ethofumesate safe to sugarbeet; and b) does ethofumesate control weeds?

MATERIALS AND METHODS

Experiments were conducted on indigenous populations of common lambsquarters and redroot pigweed in sugarbeet grower fields near Moorhead and Oslo, Minnesota and Grand Forks, Minto, and Prosper, North Dakota in 2017. Experimental area was prepared with a Kongskilde 's-tine' field cultivator equipped with rolling baskets or with grower cooperatortillage equipment before planting. Experiments were established in fields in 1 or 2 days after grower cooperatortillage field to sugarbeet. Herbicide treatments were applied when sugarbeet was at the 2-leaf and 6-leaf stage with a bicycle wheel sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 40 psi to the center four rows of six row plots 30 feet long. Treatments consisted of two applications of ethofumesate at 6, 12, 18, 24, 32 and 64 fl oz/A either alone or with glyphosate at 28 fl oz/A. All treatments of ethofumesate alone contained Destiny HC at 1.5 pt/A. Treatments of ethofumesate plus Roundup PowerMax (glyphosate) contained Destiny HC at 1.5 pt/A plus N-Pak ammonium sulfate at 2.5% v/v. Destiny HC and N-Pak AMS were provided by Winfield United.

Sugarbeet injury and common lambsquarters and/or redroot pigweed control were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was

randomized complete block with 4 replications. Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

RESULTS

Common lambsquarters control from two postemergence applications of ethofumesate ranged from 0 to 78% across rates and locations (Table 1). Lambsquarters control averaged across ethofumesate rates alone ranged from 27% at Prosper to 49% at Minto. Lambsquarters control generally increased as ethofumesate rate increased from 6 to 64 fl oz/A. However, lambsquarters control was not adequate at any rate within location or at any location for ethofumesate to be considered a stand-alone herbicide for controlling lambsquarters.

Lambsquarters control from two applications of Roundup PowerMax (glyphosate) at 28 fl oz/A was 70% and 90% at Moorhead and Oslo, respectively. Ethofumesate + glyphosate tended to improve lambsquarters control compared to ethofumesate or glyphosate alone.

Table 1. Common lambsquarters control, 27 to 48 DAT, at Moorhead and Oslo, MN and Grand Forks, Minto, and Prosper, ND, 2017

Treatment ¹	Rate fl oz/A	Application timing ²	Moorhead	Oslo	Grand	Minto	Prosper
			MN	MN	Forks ND	ND	ND
Ethofumesate / Ethofumesate	6 / 6	A/ B	20	20	0	25	13
Ethofumesate / Ethofumesate	12 / 12	A/ B	28	35	28	40	15
Ethofumesate / Ethofumesate	18 / 18	A/ B	35	38	30	48	30
Ethofumesate / Ethofumesate	24 / 24	A/ B	35	40	43	60	33
Ethofumesate / Ethofumesate	32 / 32	A/ B	50	40	53	55	35
Ethofumesate / Ethofumesate	64 / 64	A/ B	53	58	78	63	33
PowerMax ³ / PowerMax	28 / 28	A / B	70	90	100	98	95
Etho + PMax / Etho + PMax	6 + 28/6 + 28	A/ B	78	98	100	90	100
Etho + PMax / Etho + PMax	12 + 28/12 + 28	A/ B	78	94	100	98	100
Etho + PMax / Etho + PMax	18 + 28/18 + 28	A/ B	70	100	100	95	100
Etho + PMax / Etho + PMax	24 + 28/24 + 28	A/ B	78	100	100	100	100
Etho + PMax / Etho + PMax	32 + 28/32 + 28	A/ B	78	99	100	100	100
Etho + PMax / Etho + PMax	64 + 28/64 + 28	A/ B	83	99	100	100	100
LSD (0.05)			10	10	10	12	11

¹Treatments of Ethofumesate + Roundup PowerMax were applied with N-Pak AMS at 2.5% v/v and Destiny HC at 1.5 pt/A; Ethofumesate was applied with Destiny HC at 1.5 pt/A

²Application timing A=2 lf sugarbeet; B= 6 lf sugarbeet

³PowerMax or PMax=Roundup PowerMax; Etho=ethofumesate

Redroot pigweed control from ethofumesate was evaluated at Minto and Prosper, ND and Oslo, MN. Pigweed control ranged from 15% to 70% across ethofumesate rates and locations (Table 2). Pigweed control averaged across ethofumesate rates was 34%, 22%, and 41%, at Oslo, Minto, and Prosper, respectively, or similar to lambsquarters control. As with lambsquarters, ethofumesate applied postemergence is not an effective stand-alone herbicide for controlling redroot pigweed.

Waterhemp control from ethofumesate at Moorhead was a different story than redroot pigweed or lambsquarters. Waterhemp control ranged from 95% from two applications of ethofumesate at 12 fl oz/A to 100% control from two applications at 32 fl oz/A. Waterhemp control tended to increase as the ethofumesate rate increased from 6 to 64 fl oz/A. Waterhemp control from ethofumesate was superior to control from glyphosate.

Differences in broadleaf control from ethofumesate might be related to weed species emergence patterns and application timing. We know the number of growing degree days to trigger lambsquarters and redroot pigweed germination and emergence is much less (lambsquarters) to less (redroot pigweed) than waterhemp (Werle, 2014). Also, since we know that ethofumesate does not translocate from treated leaves to new tissue in emerged vegetation (Eshel, 1978), then it is likely that ethofumesate applied postemergence does little to control emerged weeds but is effective on later flushes once activated by precipitation.

Table 2. Redroot pigweed and waterhemp (Moorhead) control, 30 to 41DAT, at Moorhead and Oslo, MN and Minto, and Prosper, ND, 2017

Treatment ¹	Rate	Application timing ²	Waterhemp		Redroot pigweed	
			Moorhead MN	Oslo MN	Minto ND	Prosper ND
	fl oz/A		-----% control-----			
Ethofumesate / Ethofumesate	6 / 6	A / B	83	25	15	23
Ethofumesate / Ethofumesate	12 / 12	A / B	95	35	15	28
Ethofumesate / Ethofumesate	18 / 18	A / B	95	33	18	38
Ethofumesate / Ethofumesate	24 / 24	A / B	98	28	20	40
Ethofumesate / Ethofumesate	32 / 32	A / B	100	33	25	45
Ethofumesate / Ethofumesate	64 / 64	A / B	99	50	40	70
PowerMax ³ / PowerMax	28 / 28		68	93	95	100
Etho + PMax / Etho + PMax	6 + 28/6 + 28	A / B	95	100	90	100
Etho + PMax / Etho + PMax	12 + 28/12 + 28	A / B	98	95	95	100
Etho + PMax / Etho + PMax	18 +28/18 + 28	A / B	100	100	93	100
Etho + PMax / Etho + PMax	24 +28/24 + 28	A / B	100	100	90	100
Etho + PMax / Etho + PMax	32 +28/32 + 28	A / B	100	99	94	100
Etho + PMax / Etho + PMax	64 +28/64 + 28	A / B	100	100	98	100
LSD (0.05)			8	10	8	15

¹Treatments of Ethofumesate + Roundup PowerMax were applied with N-Pak AMS at 2.5% v/v and Destiny HC at 1.5 pt/A; Ethofumesate was applied with Destiny HC at 1.5 pt/A

²Application timing A=2 lf sugarbeet; B= 6 lf sugarbeet

³PowerMax or PMax=Roundup PowerMax; Etho=ethofumesate

Sugarbeet injury from two applications of ethofumesate alone was negligible across locations in these experiments (Table 3). Sugarbeet injury was negligible even when ethofumesate rate increased from 6 to 64 fl oz/A. Sugarbeet injury from ethofumesate plus glyphosate was similar to injury from either ethofumesate or glyphosate alone.

Table 3. Sugarbeet injury, 27 to 48 DAT, at Moorhead and Oslo, MN and Grand Forks, Minto, and Prosper, ND, 2017

Treatment ¹	Rate	Application timing ²	Moorhead	Oslo	Grand	Minto	Prosper
			MN	MN	Forks ND	ND	ND
	fl oz/A		-----% injury-----				
Ethofumesate / Ethofumesate	6 / 6	A / B	8	3	0	0	3
Ethofumesate / Ethofumesate	12 / 12	A / B	0	5	0	0	0
Ethofumesate / Ethofumesate	18 / 18	A / B	3	3	0	0	3
Ethofumesate / Ethofumesate	24 / 24	A / B	3	3	0	0	3
Ethofumesate / Ethofumesate	32 / 32	A / B	3	3	3	5	0
Ethofumesate / Ethofumesate	64 / 64	A / B	3	8	0	0	10
PowerMax / PowerMax	28 / 28	A / B	0	3	0	0	3
Etho + PMax / Etho + PMax	6 + 28/6 + 28	A / B	3	5	0	0	0
Etho + PMax / Etho + PMax	12 + 28/12 + 28	A / B	3	3	0	3	0
Etho + PMax / Etho + PMax	18 +28/18 + 28	A / B	0	3	0	3	3
Etho + PMax / Etho + PMax	24 +28/24 + 28	A / B	7	5	3	0	8
Etho + PMax / Etho + PMax	32 +28/32 + 28	A / B	13	5	0	0	0
Etho + PMax / Etho + PMax	64 +28/64 + 28	A / B	5	10	5	3	8
LSD (0.05)			NS	NS	NS	NS	NS

¹Treatments of Ethofumesate + Roundup PowerMax were applied with N-Pak AMS at 2.5% v/v and Destiny HC at 1.5 pt/A; Ethofumesate was applied with Destiny HC at 1.5 pt/A

²Application timing A=2 lf sugarbeet; B= 6 lf sugarbeet

³PowerMax or PMax=Roundup PowerMax; Etho=ethofumesate

LITERATURE CITED

- Abulnaja, KO, Tighe CR, and Harwood JL (1992) Inhibition of fatty acid elongation provides a basis for the action of the herbicide, ethofumesate on surface wax formation. *Phytochem* 31:1155-1159.
- Eshel, J, Zimdahl RL, and Schweizer EE (1978) Uptake and translocation of ethofumesate in sugarbeet plants. *Pestic. Sci.* 9: 301-304.
- Sullivan EF and Fagala, LT (1970) Herbicide evaluations on sugar beets Research Report, NC Weed Cont Conf. 27:25-27.
- Werle R, Sandell LD, Buhler DD, Hartzler RG, and Lindquist JL (2014) Predicting Emergence of 23 Summer Annual Weed Species. *Weed Sci.* 62:267-279.

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CONTINUED REFINEMENT OF THE WATERHEMP CONTROL STRATEGY IN SUGARBEET

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SUMMARY

Chloroacetamide herbicide application timing tended to have a greater effect on waterhemp control than choice of chloroacetamide herbicide.

Split application of chloroacetamide herbicides improved waterhemp control compared to a single chloroacetamide herbicide application.

Applying Dual Magnum preemergence (PRE) fb a chloroacetamide herbicide lay-by improved waterhemp control compared to chloroacetamide alone.

Lambsquarters control from glyphosate + ethofumesate was not affected by chloroacetamide herbicide applied with glyphosate and ethofumesate (data not presented).

INTRODUCTION

Survey data indicates waterhemp is the primary weed control challenge in sugarbeet fields in Southern Minnesota Beet Sugar Cooperative, in Minn-Dak Farmers' Cooperative, and in fields south of Grand Forks in American Crystal Sugar Cooperative. Waterhemp populations are a mixture of glyphosate susceptible and resistant biotypes. Roundup PowerMax at 28 fl oz/A controlled 78% of the first flush of emerged waterhemp based on waterhemp counts taken immediately prior to and 9 days following application (Peters, 2015). However, control does not improve by increasing the glyphosate rate or with repeat glyphosate applications. Early-season weed escapes cause late-season weed control failures and weed disasters at harvest. There are no effective POST herbicide options for rescue control of resistant biotypes, especially when waterhemp is greater than 4-inches tall.

Ethofumesate or Ro-Neet provide effective early-season waterhemp control but are expensive or do not provide full-season control (Peters, 2016). Use of site of action (SOA) 15 herbicides (chloroacetamides) applied early postemergence (EPOST) provide the most effective and consistent waterhemp control (Peters, 2015; Peters, 2016; Peters, 2017). However, several important statements should be made about chloroacetamide herbicides and waterhemp control. First, sugarbeet must reach the 2-leaf stage before chloroacetamides can be applied. Thus, planting date influences how and when they can be applied. Second, chloroacetamides need to be activated by timely precipitation in order to control waterhemp. Third, waterhemp seems to be emerging earlier in the spring. Are we selecting for earlier germinating biotypes or have we improved awareness and identification? Maybe some of both. Finally, sugarbeet grower surveys indicate approximately 85% satisfaction (excellent or good response) with current waterhemp control strategies. How can we improve satisfaction to 90% or 95%?

Waterhemp control in soybean was improved using repeat application of chloroacetamide herbicides; a practice referred to as 'layering' (Steckel, 2002). Sugarbeet experiments conducted at Herman and Moorhead, MN in 2015 investigate repeat applications of chloroacetamide herbicides in sugarbeet. Dual Magnum (s-metolachlor) at 0.5 pt/A was applied PRE followed by glyphosate + ethofumesate plus either S-metolachlor, Warrant or Outlook at 2-leaf sugarbeet stage. Waterhemp control averaged greater than 90% using the layering strategy compared to S-metolachlor, Warrant, or Outlook applied EPOST (Figure 1).

Outlook often is split-applied at 12 fl oz/A at the 2-leaf sugarbeet stage followed by 12 fl oz/A at the 6-leaf stage. This practice is common when glyphosate plus Outlook is tank-mixed with an insecticide for black cutworm control since there is a concern that applying multiple products formulated as emulsifiable concentrates may injury sugarbeet, especially under cold and wet spring environmental conditions. Split application can also improve waterhemp control consistency (conversation with Jim Radermacher, 2015). Split lay-by application buffers against the possibility of inadequate or untimely precipitation since the first application in May is followed by a second application, 14 to 21 days later, in June.

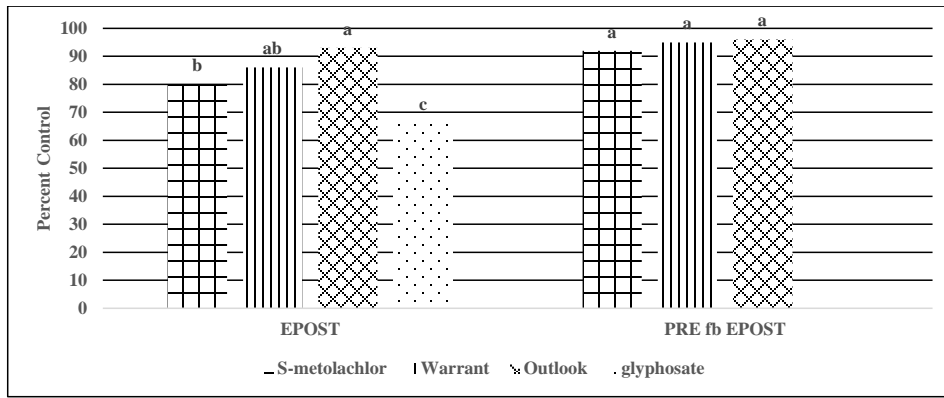


Figure 1. Waterhemp control from soil-residual herbicides applied early postemergence (EPOST) or S-Metolachlor at 0.5 pt/A preemergence (PRE) followed by soil-residual herbicides applied EPOST, averaged across Herman and Moorhead, MN, 2015.

Following successes with Outlook, sugarbeet growers and Agriculturalists have asked if Warrant and S-metolachlor should also be split-applied. The objectives of 2016 and 2017 experiments were to evaluate sugarbeet safety and waterhemp control at multiple locations from: a) Dual Magnum PRE-followed by S-metolachlor, Warrant, or Outlook EPOST in single or multiple applications and; b) S-metolachlor, Warrant, or Outlook EPOST in single or multiple applications. This report summarizes experiments conducted at Roseland, MN in 2016 and Lake Lillian, MN, and Galchutt, ND in 2017.

MATERIALS AND METHODS

Experiments were conducted on natural populations of waterhemp near Moorhead, MN in 2016 and Lake Lillian, MN and Galchutt, ND in 2017. Experimental area was prepared using a field cultivator prior to planting. Hilleshog ‘HM4302RR’ sugarbeet treated with Tachigaren, at 45 grams product, Cruiser Maxx (contains Cruiser 5FS at 60 gram active ingredient (g a.i.), Apron XL at 15 g a.i., and Maxim 4FS at 2.5 g a.i.) and Vibrance at 2g a.i. per 100,000 seeds was seeded 1.25 inches deep in 22 inch rows at 60,825 seeds per acre on May 12, 2016 at Moorhead. Crystal ‘M380’ sugarbeet treated with Tachigaren and Kabina at 45 g product and 14 g a.i. per 100,000 seeds, respectfully, was seeded 0.5 inches deep in 22 inch rows at 62,100 seeds per acre on May 8, 2017 at Lake Lillian, MN. ‘HM4022RR’ sugarbeet treated with Tachigaren, at 45 grams product, Cruiser Maxx (contains Cruiser 5FS at 60 gram active ingredient (g a.i.), Apron XL at 15 g a.i., and Maxim 4FS at 2.5 g a.i.) and Vibrance at 2g a.i. per 100,000 seeds was seeded 1.25 inches deep in 22 inch rows at 60,825 seeds per acre on May 9, 2017 at Galchutt.

Table 1. Application information for sugarbeet trial near Moorhead, MN in 2016.

Application code	A	B	C
Date	May 16	June 6	June 20
Time of Day	9:00 AM	2:00 PM	2:30 PM
Air Temperature (F)	51	67	73
Relative Humidity (%)	56	56	37
Wind Velocity (mph)	7	12	10
Wind Direction	N	NW	NW
Soil Temp. (F at 6")	48	62	70
Soil Moisture	Poor	Good	Good
Cloud Cover (%)	80	90	10
Sugarbeet stage (avg)	PRE	4-6 lf	10 lf
Waterhemp	-	0.5 inch	1-3 inch

Herbicide treatments were applied at Moorhead on May 16, June 6, and June 20, 2016; May 11, June 1, and June 16, 2017 at Lake Lillian, and May 9, June 1, and June 20, 2017 at Galchutt. All treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 40 psi to the center four rows of six row plots 30 feet in length in fields with moderate to heavy infestations of glyphosate-resistant waterhemp. Ammonium sulfate (AMS) in all treatments was 'N-Pak' AMS, a liquid formulation from Winfield United. 'Destiny HC' high surfactant methylated oil concentrate (HSMOC) was also used and is a product from Winfield United.

Table 2. Application information for sugarbeet trial near Lake Lillian, MN in 2017.

Application code	A	B	C
Date	May 11	June 1	June 16
Time of Day	9:00 AM	9:00 AM	9:00 AM
Air Temperature (F)	58	70	79
Relative Humidity (%)	27	27	42
Wind Velocity (mph)	12	3	5-10
Wind Direction	NNW	SSW	SSE
Soil Temp. (F at 6")	68	70	-
Soil Moisture	Good	Good	Good
Cloud Cover (%)	-	-	Partly Cloudy
Sugarbeet stage (avg)	PRE	2-4 lf	6-8 lf
Waterhemp	-	0.5 inch	1-3 inch

Sugarbeet injury was evaluated June 24 and July 22, 2016 at Moorhead, June 6, June 26 and July 6, 2017 at Lake Lillian, and June 16, 2017 at Galchutt. Waterhemp control was evaluated June 24, June 28, July 22, and August 24, 2016 at Moorhead, June 15, June 26 and July 6, 2017 at Lake Lillian and June 16, July 5, and July 24, 2017 at Galchutt. Common lambsquarters and redroot pigweed control also was evaluated at each location, but data are not included in this report since glyphosate provided complete or near complete control of both species. All evaluations were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was randomized complete block with 4 replications. Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

Table 3. Application information for sugarbeet trial near Galchutt, ND in 2017.

Application code	A	B	C
Date	May 9	June 1	June 20
Time of Day	12:00 PM	9:00 AM	12:00 PM
Air Temperature (F)	64	70	68
Relative Humidity (%)	37	32	47
Wind Velocity (mph)	10	3	6
Wind Direction	NW	S	NW
Soil Temp. (F at 6")	54	59	64
Soil Moisture	Good	Good	Good
Cloud Cover (%)	50	10	10
Sugarbeet stage (avg)	PRE	2-lf	8-10 lf
Waterhemp	-	1 inch	2 inch

RESULTS

Waterhemp control was influenced by herbicide and application timing at Moorhead in 2016 and Lake Lillian and Galchutt in 2017 (Figure 2, Figure 3, Figure 4). In general, application timing had greater influence on waterhemp control than chloroacetamide herbicide.

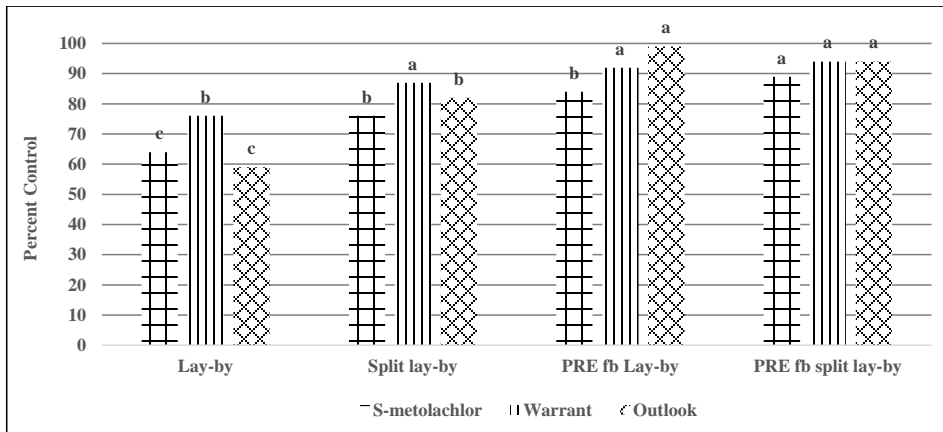


Figure 2. Waterhemp control from single lay-by or split lay-by herbicide applications and S-metolachlor preemergence (PRE) followed by lay-by or split lay-by herbicide applications, Moorhead, MN in 2016, average of July 22 and August 24 evaluation.

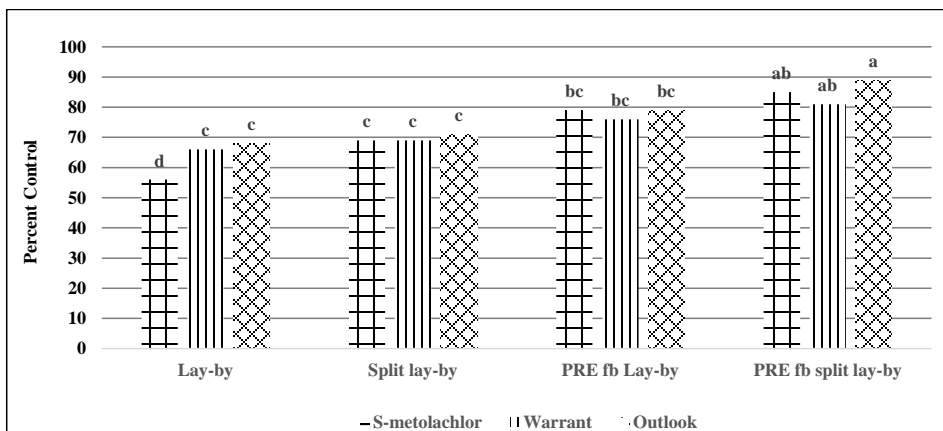


Figure 3. Waterhemp control from single lay-by or split lay-by herbicide applications and S-metolachlor preemergence (PRE) followed by lay-by or split lay-by herbicide applications, Lake Lillian, MN, 2017, July 6 evaluation.

There are several factors to consider when selecting a chloroacetamide herbicide for waterhemp control aside from relationships with a company or company representatives. Warrant costs less per acre on a rate basis than Outlook or S-metolachlor. Outlook is more water soluble than either S-metolachlor or Warrant and requires less precipitation for activation. Once activated, Warrant has longer residual than Outlook or S-metolachlor. Outlook and Warrant have a broader weed control spectrum than S-metolachlor. However, sugarbeet can be planted directly into S-metolachlor residues in the event of replant whereas three to four weeks' time is required before residue levels of Outlook and Warrant will allow sugarbeet replanting. Finally, S-metolachlor and Warrant are safer on sugarbeet than Outlook although injury generally is negligible with all chloroacetamide herbicides. Most of the factors to consider when selecting a chloroacetamide herbicide are based more around risk of sugarbeet injury than level of waterhemp control.

Waterhemp control from chloroacetamide herbicides was evaluated across locations in 2014 to 2017. Precipitation followed within 7-days of chloroacetamide activation in 2014 and 2015. However, timely precipitation did not occur in 2016 or 2017. 2016 was a dry spring, creating erratic germination and emergence patterns in experiments and in grower fields. Early postemergence chloroacetamide application was delayed five days to account for erratic emergence at the Moorhead location. Likewise, precipitation was spotty and possibly up to 24 days between the precipitation event that activated PRE herbicides and precipitation events to activate lay-by herbicides in 2017 at Lake Lillian. These climate phenomena partially explain waterhemp control observations in fields in 2016 and 2017 (Figure 2 and Figure 3).

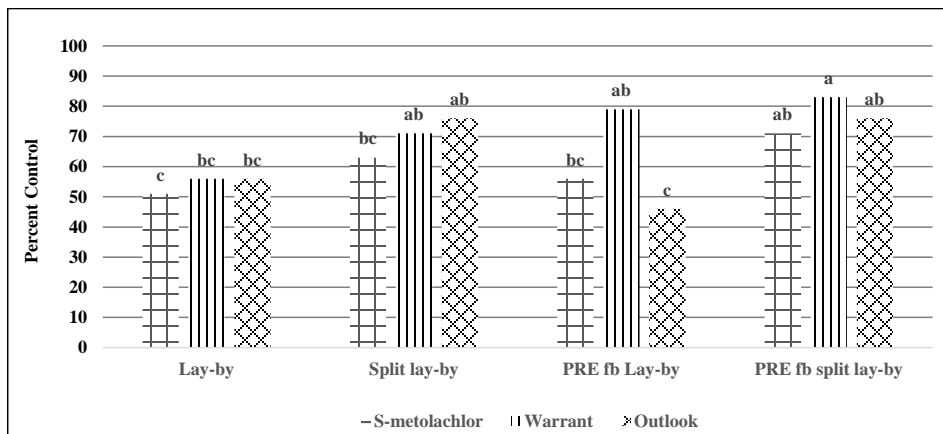


Figure 4. Waterhemp control from single lay-by or split lay-by herbicide applications and S-metolachlor preemergence (PRE) followed by lay-by or split lay-by herbicide applications, Galchutt, ND, 2017, July 25 evaluation.

The Galchutt, ND location received timely precipitation for activation of herbicides in 2017 (Figure 4). However, there was significant sugarbeet stand loss caused by rhizoctonia root rot, possibly caused by above average precipitation in June and July. Stand loss created an open canopy suitable for waterhemp germination and emergence well into July. Under these conditions, split application of chloroacetamide herbicides (EPOST fb POST) or PRE followed by split applications of chloroacetamide herbicides tended to provide better waterhemp control than single lay-by application of chloroacetamide herbicide alone or following PRE S-metolachlor.

At each of the three locations, 12 different treatment combinations of herbicide (S-metolachlor, Warrant, and Outlook) and timing (lay-by, split lay-by, PRE fb lay-by, and PRE fb split lay-by) were tested for a total of 36 observable treatments. In an effort to compare these treatments and determine which method of application resulted in the greatest and most constant control across locations, the following steps were taken. At each evaluation from each location, waterhemp control data was ranked in numerical order from greatest control to least control based upon the least significant difference (LSD). Herbicide treatments that were statistically the same as the best treatment at each evaluation timing from each location were grouped into a cluster and labeled 'good'. The remaining treatments were once again ranked and grouped into a second and third cluster based on LSD value and labeled 'fair' and 'poor', respectively. Clusters were titled 'good', 'fair' and 'poor' since treatments in the good cluster generally corresponded to 80% or greater waterhemp control, the fair cluster corresponded to 80 to 65% waterhemp control, and the poor cluster corresponded to 65 to 40% waterhemp control. Chloroacetamide herbicides were combined and were grouped by application timing into four classes: lay-by, split lay-by, PRE fb lay-by, and PRE fb split lay-by. The number of observations corresponding to each cluster (good, fair, or poor) were summed and are presented in Figure 5. Data indicates PRE fb lay-by and PRE fb split lay-by application methods provided the most consistent waterhemp control across locations and years.

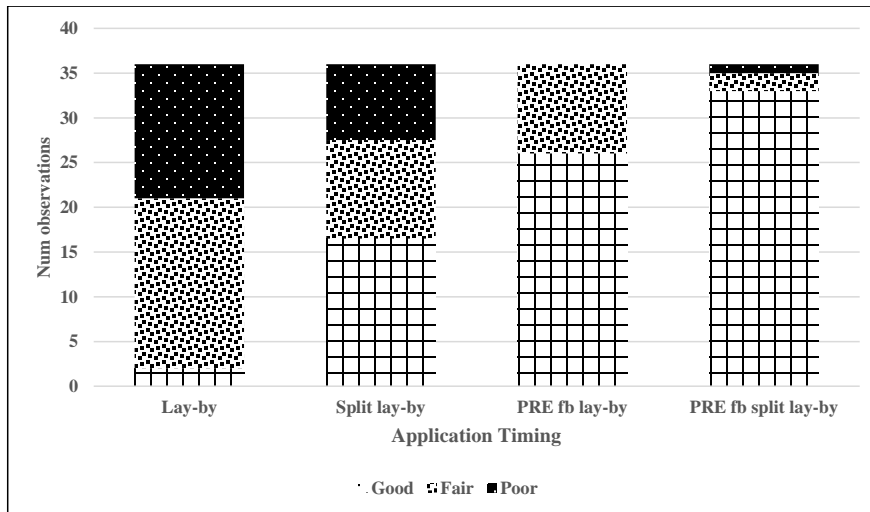


Figure 5. Number of good, fair, and poor estimates of waterhemp control across herbicides and application timing, summed across evaluations, locations, and years

CONCLUSIONS

Sugarbeet planting date is likely the most important factor to consider for herbicide selection and application timing for waterhemp control (Table 4). Split lay-by application of chloroacetamide herbicides is the preferred approach for waterhemp control for early planted sugarbeet. However, PRE followed by a split lay-by application buffers risk against early germinating weeds or uncertainty of when precipitation will occur to activate lay-by herbicides, even in early planted sugarbeet.

Late planted sugarbeet may not reach the sugarbeet 2-If stage by May 15 (date when the growing degree day model typically forecasts waterhemp germination and emergence). Thus, Dual Magnum and/or ethofumesate should be applied PRE followed by split lay-by application of chloroacetamide herbicides. Timing of the lay-by applications will be dependent on sugarbeet planting date, precipitation to activate PRE, and waterhemp pressure in the field.

Continue to scout sugarbeet fields for waterhemp in July and August. Tank-mixes of Betamix or UpBeet with Roundup plus ethofumesate or cultivation are recommended for POST waterhemp control. Apply in combination with HSMOC adjuvant at 1.5 pt/A and AMS at 8.5 to 17 lb/100 gallon water carrier.

Table 4. Recommendation for waterhemp control in sugarbeet, by planting date.

Planting Date	Recommendation
Plant Sugarbeet in April	Split lay-by application (early postemergence / postemergence) of chloroacetamide herbicides applied at 2-If sugarbeet fb 4 to 6-If sugarbeet
	Dual Magnum and/or ethofumesate PRE followed by a split lay-by application at 2 to 4-If stage fb 4 to 6-If stage
	Single lay-by application when sugarbeet is at the 2-If stage or greater
Plant Sugarbeet in May	Dual Magnum and/or ethofumesate PRE followed by a split lay-by
Either	Continue to scout fields for late germinating waterhemp in late June and July
Either	Be prepared to rescue with Betamix + ethofumesate, UpBeet + ethofumesate or Betamix + UpBeet (be aware of resistant biotypes)

LITERATURE CITED

- Peters, TJ, Carlson AL (2015) Controlling Waterhemp in Fields Planted to Sugarbeet. Sugarbeet Res. Ext. Rept. 45: 29-35.
- Peters, TJ, Lueck AB, Radermacher J (2016) A Strategy for Managing Waterhemp in Sugarbeet. Sugarbeet Res. Ext. Rept. 46: 22-30.
- Peters, TJ, Lueck AB, Groen C (2017) Continued Evaluation of the Strategy for Managing Waterhemp in Sugarbeet. Sugarbeet Res. Ext. Rept. 47: 30-38.
- Steckel, LE, Sprague CL, Hager AG (2002) Common Waterhemp (*Amaranthus rudis*) Control in Corn (*Zea mays*) with Single Preemergence and Sequential Applications of Residual Herbicides. Weed Technology. 16:755-761.

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- Research Specialist Andrew Lueck, graduate students, Alexa Lystad and Nathan Haugrud and summer student employee Bethany Christensen.
- North Dakota State University Experiment Station and University of Minnesota Crookston Research and Outreach Center

COMPARING HERBICIDES FOR BROADLEAF WEED CONTROL IN SUGARBEET

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The objective of this study was to evaluate broadleaf weed control from single applications of individual herbicides currently registered for use in Roundup Ready (RR) sugarbeet.

MATERIALS AND METHODS

An experiment was conducted near Hickson, ND in 2017. Fertilizer was spread April 11 and incorporated the same day with a field cultivator equipped with a spring tooth harrow. The trial site was prepared using a Kongskilde 's-tine' field cultivator with rolling baskets on May 13, 2017. Four-foot-wide strips of bioassay species including canola, amaranth, quinoa, and flax were seeded perpendicular to sugarbeet on May 13. Seedex 'Winchester' sugarbeet, treated with NipsIt Suite, Tachigaren at 45g per unit, and Kabina at 7g per unit, were then seeded in 22-inch rows at 60,560 seeds per acre on May 13 with a John Deere 1700XP 6-row planter. Post emergence (POST) treatments were applied June 9. All herbicide treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 40 psi to the center four rows of six row plots 35 feet in length.

All sugarbeet injury and weed control evaluations were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was randomized complete block with 4 replications for each trial. Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

Table 1. Application Information – Hickson, ND 2017

Date	June 9
Time of Day	12:30 PM
Air Temperature (F)	82
Relative Humidity (%)	42
Wind Velocity (mph)	9
Wind Direction	SE
Soil Temp. (F at 6")	69
Soil Moisture	Fair
Cloud Cover (%)	30
Next Rainfall (amount)	June 11 (0.11")
Sugarbeet Stage	4 leaf
Amaranth (and natural redroot pigweed)	2-6 lf / avg 4 lf
RR canola	2-4 lf/ avg 3 lf (2" tall)
Flax	2-4 inch / avg 3 inch
Quinoa (and natural common lambsquarters)	2-3 inch/ avg 3 inch
Yellow Foxtail	2-3 inch/ avg 3 inch

SUMMARY

UpBeet (triflusalufuron) is the only ALS (group 2) herbicide registered for use in sugarbeet. No sugarbeet injury was observed in this trial from either 0.5 or 1.0 oz/A of UpBeet (Table 2). UpBeet provided the greatest Roundup Ready canola control of all herbicides evaluated. Canola control increased from 73% to 90% at 13 DAT as rate increased from 0.5 to 1.0 oz/A. UpBeet gave 70 to 78% pigweed control and provided some suppression of lambsquarters, flax, and yellow foxtail.

Table 2. Sugarbeet injury and weed control from herbicides at Hickson, ND in 2017.

Herbicide	Rate	Rate Unit	16 Jun		-----22 Jun-----				
			Sgbt	Sgbt	rrpw ³	colq ⁴	cano ⁵	flax	yefx ⁶
			----% inj----		-----% cntl-----				
UpBeet ¹	0.5	oz/A	0	0	70	45	73	45	55
UpBeet ¹	1	oz/A	0	0	78	38	90	65	58
Nortron ¹	12	fl oz/A	5	0	25	25	20	45	0
Nortron ¹	16	fl oz/A	0	0	35	38	25	45	0
Nortron ¹	32	fl oz/A	13	0	50	50	35	48	0
Nortron ¹	64	fl oz/A	3	0	60	58	53	73	0
Stinger ¹	2	fl oz/A	20	- ⁷	3	23	0	0	0
Stinger ¹	4	fl oz/A	20	-	3	20	0	0	0
Roundup PowerMax ²	22	fl oz/A	0	0	99	91	0	100	100
Roundup PowerMax ²	28	fl oz/A	0	0	100	92	0	100	100
Roundup PowerMax ²	32	fl oz/A	3	0	100	95	0	100	100
Betamix ¹	12	fl oz/A	25	0	35	40	20	25	0
Betamix ¹	16	fl oz/A	40	10	48	53	18	30	5
Betamix ¹	24	fl oz/A	45	30	60	65	40	35	0
Spin-Aid ¹	12	fl oz/A	20	-	10	53	13	23	0
Spin-Aid ¹	24	fl oz/A	33	-	13	50	23	18	0
Spin-Aid ¹	36	fl oz/A	45	-	23	68	40	35	0
LSD (0.05)			15	-	14	19	11	18	5

¹Herbicide applied with MSO from Loveland at 2 pt/A + AMS at 8.5 lb/100 gal

²Herbicide applied with Prefer 90 NIS from West Central at 0.25% v/v + AMS at 8.5 lb/100 gal

³rrpw=redroot pigweed + tame amaranth

⁴colq=common lambsquarters + quinoa

⁵cano=Roundup Ready (RR) canola

⁶yefx=yellow foxtail

⁷- = no injury data was recorded due to weed competition. No LSD was calculated due to the missing data.

Nortron (ethofumesate) is the only herbicide found in group 16 and can be applied pre-plant incorporated (PPI), pre-emergence (PRE) or POST in sugarbeet. Current labeling allows for POST application of up to only 12 fl oz/A of Nortron per season. Nortron rates in this trial ranged from 12 to 64 fl oz/A. Very little sugarbeet injury was observed from any rate of Nortron evaluated in this trial at 7 DAT (0 to 13%) and no injury was observed at 13 DAT. At 12 fl oz/A, Nortron provided little control or suppression of any weed species evaluated. Control of all species increased as rate increased, but never above 75%. Nortron did not control yellow foxtail when applied POST at any rate. Though not tested in this trial, data from other trials demonstrates that Nortron improves weed control, including waterhemp or pigweed, when tank-mixed with other herbicides.

Stinger (clopyralid) is the only group 4 (growth regulator) herbicide currently labeled in sugarbeet. Stinger caused 20% sugarbeet leaf curling injury at both 2 and 4 fl oz/A at 7 DAT. This level of injury is generally tolerable early in the season. Stinger provided little to no control of any of the weeds found in this trial. Stinger is an effective herbicide to use in controlling thistle, common ragweed, and giant ragweed, but it has very little if any efficacy against amaranthus species (pigweeds and waterhemp), lambsquarters, or canola. Stinger has no grass activity.

Roundup PowerMax (glyphosate) is a group 9 herbicide and may be applied in Roundup Ready sugarbeet. Roundup is very safe in RR sugarbeet and no notable sugarbeet injury was observed in this trial at any rate tested. Roundup provided the greatest and most consistent control of all species in this trial, with the exception of RR canola. Common lambsquarters was the most difficult weed to control with Roundup, and control varied from 91 to 95% 13 DAT.

Betamix (phenmedipham + desmedipham) is a group 5 (photosynthesis inhibitor) herbicide labeled for use in sugarbeet. Betamix gave moderate sugarbeet injury at all rates tested. Injury ranged from 25 to 45% 7 DAT and 0 to

30% 13 DAT and increased as rate increased. Injury symptoms were leaf burn and some plant height reduction. Betamix provided poor to fair control of pigweed (35 to 60%) and common lambsquarters (40 to 65%), but control improved as rate increased. Weeds were 3 to 4 inches tall at time of application and Betamix is generally considered most effective when applied to cotyledon pigweed or lambsquarters. Betamix provided some suppression of RR canola and flax, but no control of yellow foxtail.

Spin-Aid (phenmedipham) is a group 5 (photosynthesis inhibitor) herbicide labeled for use in sugarbeet. Spin-Aid gave moderate sugarbeet injury at all rates tested. Injury ranged from 20 to 45% 7 DAT and increased as rate increased. Injury symptoms were leaf burn and some plant height reduction. Compared to Betamix (phenmedipham + desmedipham) Spin-Aid (phenmedipham) gave less control of pigweed (10 to 23%) and similar common lambsquarters control (50 to 68%), and control tended to improve as rate increased. Similar to Betamix, Spin-Aid is generally considered most effective when applied to cotyledon pigweed or lambsquarters. Spin-Aid provided some suppression of RR canola and flax, but no control of yellow foxtail.

CONCLUSIONS

Only six herbicide options exist for controlling broadleaf weeds POST in sugarbeet. In this trial, only Roundup PowerMax (glyphosate) gave greater than 90% control of any weeds present. UpBeet at 1 oz/A gave the greatest control of RR canola at 90%. Using UpBeet, Nortron, Stinger, Betamix, or Spin-Aid alone will not provide adequate control of pigweeds, common lambsquarters, or yellow foxtail. Using the appropriate herbicide, however, in conjunction with glyphosate, may improve control of difficult to control weeds, such as waterhemp, lambsquarters, and common ragweed and delay the selection of glyphosate resistant weeds.

EFFICACY OF ‘RESCUE’ HERBICIDES IN SUGARBEET

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The objective of this trial was to evaluate ‘rescue’ control of waterhemp using herbicides in sugarbeet. Rescue applications of herbicides are made after an initial herbicide application fails to provide adequate weed control. This is often the situation when glyphosate resistance is first observed in weeds in a field and the initial application of glyphosate failed to provide adequate weed control.

MATERIALS AND METHODS

An experiment was conducted near Lake Lillian, MN in 2017. The seedbed was prepared using a ‘s-tine’ field cultivator. Crystal ‘M380’ was seeded in 22-inch rows at 60,500 seeds per acre on May 8. Post emergence (POST) treatments were applied June 6 and 20. All herbicide treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 40 psi to the center four rows of six row plots 40 feet in length.

A similar experiment was conducted near Moorhead, MN in 2017. The seedbed was prepared using a Kongskilde ‘s-tine’ field cultivator equipped with rolling baskets on May 10. Hilleshog ‘HM4022RR’ sugarbeet was seeded in 22-inch rows at 60,560 seeds per acre on May 11 with a John Deere 1700XP 6-row planter. POST treatments were applied June 29 and July 7. All herbicide treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 35 psi to the center four rows of six row plots 40 feet in length.

All weed control evaluations were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was randomized complete block with 4 replications for each trial. Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

Table 1. Application information for trials at Lake Lillian and Moorhead, MN in 2017.

	Lake Lillian, MN		Moorhead, MN	
	A	B	A	B
Date	June 6	June 20	June 29	July 7
Time of Day	10:00 AM	9:45 AM	10:30 AM	9:30 AM
Air Temperature (F)	78	70	70	75
Relative Humidity (%)		48	69	57
Wind Velocity (mph)	10	11	0	6
Wind Direction	SE	N	NE	E
Soil Temp. (F at 6")		71	69	70
Soil Moisture	Good	Good	Good	Good
Cloud Cover (%)	0	10	95	0
Next Rainfall (amount)	June 11 (1.0")	June 28 (1.0")	July 4	July 18
Sugarbeet Stage	4 leaf	8 leaf	10-12 leaf	14-16 leaf
Waterhemp	4 inch	6 inch	2.5 inch	5 inch
Common Lambsquarters	4 inch	6 inch	4 inch	6 inch

SUMMARY

Lake Lillian

Waterhemp showed an intermediate level of glyphosate resistance. Roundup PowerMax (glyphosate) at 28 fl oz/A fb Roundup PowerMax at 28 fl oz + Ethofumesate 4 SC (ethofumesate) at 6 fl oz + Destiny HSMOC at 1.5 pt/A + N-

Pak AMS at 2.5 % v/v gave only 63% and 50% waterhemp control at 6 and 16 days after application (DAT) B, respectively (Table 2). At 16 DAT, neither UpBeet (triflusaluron) at 1 oz/A, Ethofumesate 4 SC at 12 fl oz/A, or a combination of both herbicides gave greater than 25% control of waterhemp. The lack of waterhemp control from UpBeet at 1 oz/A suggests the population may also have been resistant to ALS herbicides. No 'rescue' treatment tested gave acceptable control of waterhemp.

Table 2. Waterhemp and common lambsquarters control from rescue herbicides at Lake Lillian, MN in 2017.

Treatment	Rate/A	Appl ¹	June 26	July 6	July 6
			waterhemp	waterhemp	lambsquarters
			-----% control-----		
UpBeet + MSO	1 oz + 1.5 pt	B	3	18	0
Ethofumesate 4SC + MSO	12 fl oz + 1.5 pt	B	8	25	8
UpBeet + Ethofumesate 4SC	1 oz + 12 fl oz	B	3	20	10
+ MSO	+ 1.5 pt				
Roundup PowerMax fb	28 fl oz fb	A			
Roundup PowerMax+	28 fl oz +		63	50	100
Ethofumesate + N-Pak AMS	6 fl oz + 2.5 % v/v	B			
+ Destiny HC	+ 1.5 pt				
LSD (0.05)			11	15	4

¹Appl= Application code listed in Table 1.

Common lambsquarters control was 100% from the treatment containing Roundup PowerMax at 16 DAT (Table 2). UpBeet failed to provide any lambsquarters control. Ethofumesate 4 SC and the combination of UpBeet + Ethofumesate gave 10% or less lambsquarters control.

Moorhead

Sugarbeet injury was generally negligible from herbicides applied. Betamix at 3 pt/A gave 10% to 15% visual injury at 8 and 17 DAT (Table 3) even though sugarbeet were 14 to 16 leaf at application. Injury symptoms were necrotic spots on leaves. All other treatments gave 10% or less injury.

Waterhemp showed an intermediate level of glyphosate resistance. Control from two applications of Roundup PowerMax + Ethofumesate was 78% at 8 days after the second application but only 22% at 17 days after the second application. Treatments containing Betamix provided control ranging from 28% to 40% at 8 DAT but declined to 13% to 36% at 17 DAT. At 17 DAT, those treatments that were a tank-mix of two herbicides tended to give better control than individual herbicides, though no treatment gave greater than 36% control (Betamix + Ethofumesate). No treatment tested provided adequate control of waterhemp.

Common lambsquarters control ranged from 0 to 48% control at 17 DAT from treatments not containing Roundup. Two applications of Roundup PowerMax + Ethofumesate gave 100% common lambsquarters control at 17 DAT.

Table 3. Sugarbeet injury and waterhemp and common lambsquarters control from rescue herbicides at Moorhead, MN in 2017.

Treatment	Rate/A	Appl ¹	-----July 15-----			-----July 24-----		
			sgbt	wah	colq	sgbt	wah	colq
			-----%-----					
Betamix	3 pt	B	10	28	45	15	13	18
UpBeet	1 oz	B	8	10	3	0	8	0
Ethofumesate 4SC	12 fl oz	B	0	18	15	8	25	33
Betamix +	3 pt +	B	8	40	45	8	33	20
UpBeet	1 oz							
Betamix +	3 pt +	B	8	23	30	10	36	30
Ethofumesate 4SC	12 fl oz							
UpBeet +	1 oz +	B	0	10	23	0	30	43
Ethofumesate 4SC	12 fl oz							
Betamix +	3 pt +	B	8	30	38	5	33	48

UpBeet +	1 oz +							
Ethofumesate 4SC	12 fl oz							
Roundup PowerMax+	28 fl oz +	A						
Ethofumesate fb	6 fl oz fb		0	78	100	0	22	100
Roundup PowerMax+	28 fl oz +	B						
Ethofumesate	6 fl oz							
LSD (0.05)			NS	24	24	8	18	12

CONCLUSIONS

Treatments that did not contain Roundup PowerMax failed to provide adequate control of waterhemp, regardless of herbicide combination or location. Two applications of Roundup PowerMax failed to provide adequate waterhemp control at 16 DAT at either location. Making 'rescue' applications of POST herbicides to control waterhemp that survived a previous POST application will likely result in little to no improvement in waterhemp control in sugarbeet.

Common lambsquarters control was near perfect at both locations from two applications of Roundup PowerMax. All 'rescue' treatments tested failed to provide greater than 48% lambsquarters control at 16 DAT. However, nearly all herbicides evaluated provided some control. This suggests that, if used in conjunction with glyphosate, these herbicides may help delay the onset of glyphosate resistance in common lambsquarters.

SCREENING HERBICIDES FOR CROP SAFETY IN SUGARBEET

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The objective of this trial was to screen pre-emergence and post-emergence herbicides alone and in tank-mixes for sugarbeet crop safety.

MATERIALS AND METHODS

An experiment was conducted near Hickson, ND in 2017. Fertilizer was spread May 2 and incorporated the same day with a field cultivator equipped with a spring tooth harrow. Seedex 'Winchester' sugarbeet, treated with NipsIt Suite, Tachigaren at 45g per unit, and Kabina at 7g per unit was seeded in 22-inch rows at 60,560 seeds per acre on May 3 with a John Deere 1700XP 6-row planter. Pre-emergence (PRE) treatments were applied May 3 immediately after planting. Rain events occurred on May 3, May 7, and May 16 with 0.09, 0.02, and 0.63 inches of rain respectively. Post emergence (POST) treatments were applied June 2. All herbicide treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 40 psi to the center four rows of six row plots 35 feet in length. Sugarbeet stand was counted from 10 feet of each of the center two rows on May 26 when sugarbeet were in the cotyledon to 2 leaf stage. Sugarbeet were counted again at harvest. Roundup PowerMax at 32 fl oz/A + Veracity at 3qt/100 gal was applied June 12 and 26 to provide weed control. Escaped weeds were hand pulled throughout the season. Quadris at 16 fl oz/A was broadcast June 24 to control *Rhizoctonia* root rot. Proline at 5.7 fl oz/A + NIS at 0.125% v/v and AgriTin at 8 fl oz/A + Topsin at 12 fl oz/A were applied July 18 and August 2, respectively, to control *Cercospora* Leaf Spot. Sugarbeet in the center two rows by 27 feet long were harvested September 7, 2017. Roots were weighed and about 25 lbs of representative roots were collected from each plot and taken to Minn-Dak Farmers Cooperative Quality Lab in Wahpeton, ND for sugar and purity analysis.

All sugarbeet injury evaluations were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was randomized complete block with 4 replications for each trial. Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

Table 1. Application Information – Hickson, ND 2017

Date	May 3	June 2
Time of Day	3:00 PM	9:30 PM
Air Temperature (F)	63	86
Relative Humidity (%)	53	45
Wind Velocity (mph)	9	9
Wind Direction	NW	S
Soil Temp. (F at 6")	50	60
Soil Moisture	Good	Good
Cloud Cover (%)	100	5
Next Rainfall (amount)	May 3 (0.09")	June 11 (0.11")
Sugarbeet Stage	PRE	cot-4 leaf/ avg 2 leaf

SUMMARY

Sugarbeet stand counts were taken 7 days before POST treatments were applied. No significant differences were observed among PRE treatments as compared to the untreated check (Table 2). Sugarbeet stands were consistent across the trial. Sugarbeet were counted again on September 7 following defoliation but prior to harvesting. Sugarbeet treated with Satellite Hydrocap (pendamethalin), Cobra (lactofen), or Ultra Blazer (acifluorfen) showed or tended to show decreased stand compared to the untreated check. The treatment of Satellite Hydrocap + Devrinol 2-XT (napropamide) had the fewest sugarbeet of all treatments.

Table 2. Sugarbeet stand and injury ratings from herbicides, Hickson, ND 2017.

Treatment	Rate/A	Timing ¹	May 26	Sep 7	Jun 5	Jun 5	Jun 5	Jun 15	Jun 22	Jun 27
			Stand	Stand	Inj ²	Necr	Chlo	Inj	Inj	Inj
			--- #/100' ---		-----%-----					
Untreated			188	204	0	0	0	0	5	10
KFD 152-02	1 pt	PRE	184	204	25	0	25	3	15	8
Devrinol 2-XT	4 pt	PRE	194	202	8	1	1	3	13	13
KFD 152-02 + Devrinol 2-XT	1 pt + 4 pt	PRE	201	194	30	0	33	8	15	13
Satellite Hydrocap	1.58 pt	PRE	189	183	25	3	0	33	38	38
Satellite Hydrocap + Devrinol 2-XT	1.58 pt + 4 pt	PRE	199	175	25	3	0	33	40	38
Cobra + COC	10 fl oz + 1.5 pt	POST	198	183	65	70	0	70	80	68
UltraBlazer + COC	1 pt + 1.5 pt	POST	198	186	70	80	0	73	68	65
LSD (0.05)			NS	21	13	7	8	9	9	12

¹Timing information displayed in Table 1.²Inj=injury, Necr=necrosis, Chlo=chlorosis

Sugarbeet injury from herbicide treatments varied from 0 to 80% (Table 2). Devrinol 2-XT gave non-significant injury at all visual evaluations. KFD 152-02 (clomazone) applied alone or with Devrinol, showed 25% to 33% chlorosis/bleaching injury early in the season with these injury symptoms diminishing as the season progressed. Satellite Hydrocap applied alone or with Devrinol gave similar sugarbeet injury ranging from 25% to 40% and was generally consistent across evaluations. Variable injury responses were noted from plant to plant from the Satellite application where one plant could be healthy and the adjacent plant showed reduced stature. Cobra or Ultra Blazer applied with crop oil concentrate (COC) gave the greatest amount of injury from 65 to 80%. The injury was leaf necrosis. Both Cobra and Ultra Blazer were applied to small sugarbeet (cot – 4 leaf) and hot weather followed application. These factors may have helped increase injury to such high levels. Injury was generally similar between Cobra and Ultra Blazer, but, as time passed, sugarbeet treated with Ultra Blazer tended to show slightly less injury than those treated with Cobra.

Sugarbeet yield parameters varied by herbicide treatment (Table 3). Root yield was similar from the untreated check, KFD 152-02, Devrinol 2-XT, KFD + Devrinol, Satellite Hydrocap, and Satellite + Devrinol. Sugarbeet treated with Cobra or Ultra Blazer showed 6.2 and 6.8 ton/A reductions in root yield compared to the untreated check. No significant differences were detected in percent sugar, however, there was a tendency from KFD 152-02, Satellite Hydrocap, and Satellite + Devrinol to reduce sugar percentage 0.5% to 0.7% from the untreated check. Purity from these three treatments also tended to be less than the untreated check. Extractable sucrose per acre was greatest from the untreated check. Satellite Hydrocap and Satellite + Devrinol reduced sucrose by about 1,000 lbs/A compared to the check. Cobra and Ultra Blazer reduced extractable sucrose by about 2,000 lbs/A compared to the check.

Table 3. Sugarbeet yield and quality from herbicides, Hickson, ND 2017.

Treatment	Rate/A	Timing ¹	Yield	Sugar	Purity	Ext. Sucrose	Ext. Sucrose
			ton/A	%	%	lb/ton	lb/A
Untreated			31.8	16.7	90.8	288	9149
KFD 152-02	1 pt	PRE	31.3	16.0	89.6	270	8422
Devrinol 2-XT	4 pt	PRE	31.1	16.9	90.3	288	8964
KFD 152-02 + Devrinol 2-XT	1 pt + 4 pt	PRE	30.8	16.9	90.8	291	8967
Satellite Hydrocap	1.58 pt	PRE	30.0	16.2	89.5	273	8185
Satellite Hydrocap + Devrinol 2-XT	1.58 pt + 4 pt	PRE	29.5	16.0	89.8	271	7981
Cobra + COC	10 fl oz + 1.5 pt	POST	25.2	16.6	90.5	284	7082
UltraBlazer	1 pt	POST	25.0	16.7	90.9	289	7128

+ COC	+ 1.5 pt					
LSD (0.05)		2.9	NS	NS	NS	979

[†]Timing information displayed in Table 1.

CONCLUSIONS

Devrinol 2-XT appears very safe to sugarbeet when applied PRE at 4 pt/A. KFD 152-02 and Satellite Hydrocap tended to impact sugarbeet quality to a greater extent than root yield. Sugarbeet treated with Cobra or Ultra Blazer were severely injured and failed to make a full recovery in time for harvest. Improved crop safety from these products may be seen with reducing rates or delaying application to larger sugarbeet, but additional research should be conducted to test this hypothesis.

LIBERTY APPLIED WITH ADJUVANTS IN LIBERTYLINK SOYBEAN

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BACKGROUND

Liberty (glufosinate) is a broad spectrum grass and broadleaf control herbicide used in combination with LibertyLink soybean. Liberty is applied postemergence at 0.53 to 0.65 lb ai/a (29 to 36 fl oz/A) between soybean emergence and pre bloom when weeds are up to three inches tall. A repeat Liberty application can be made at up to 0.53 lb ai/a. Liberty is applied with ammonium sulfate (AMS) at 3 lb/a in at least 15 gal/a water using nozzles and pressure to produce a medium sized droplet. Using Liberty in LibertyLink crops offers growers herbicide diversity since it has a unique site of action (SOA 10) and controls glyphosate-resistant weeds including kochia, common ragweed, and waterhemp.

Ammonium sulfate should always be added when using Liberty herbicide. Ammonium sulfate enhances Liberty absorption and movement through the leaf cuticle. Calcium magnesium, sodium, and potassium have been reported to reduce the efficacy of weak acid herbicides like Liberty. Ammonium sulfate counteracts the antagonistic effects of hard water salts. As water in the spray droplet evaporates, sulfate from AMS binds with antagonistic salts which prevents them from binding with Liberty. In addition, ammonium from AMS binds with Liberty resulting in greater uptake into the plant and greater resultant weed control.

There are many products, including liquid-based products, that improve herbicide uptake and deactivate antagonistic hard water salts. Liquid-based products tend to be easier to handle and have given consistent performance in trials when used with glyphosate. ET-4000 is an acidic ammonium sulfate replacement. ET-4000 is a sulfuric acid based product that turns to a sulfate when in the presence of water. The objective of this study was to evaluate common lambsquarters and waterhemp control from liquid-based AMS replacements applied with Liberty.

MATERIALS AND METHODS

An experiment was conducted near Moorhead, MN in 2017. The trial site was prepared for planting using a Kongskilde s-tine field cultivator on May 10, 2017. Peterson Farm 'L07-16N' LibertyLink soybean was planted in 22-inch rows at 160,000 seeds per acre on May 11 with a John Deere 1700XP 6-row planter. Postemergence (POST) treatments were applied June 19. All herbicide treatments were applied with a bicycle sprayer in 17 gpa spray solution through 8002 XR flat fan nozzles pressurized with CO₂ at 35 psi to the center four rows of six row plots 30 feet in length. Soybean injury and common lambsquarters and waterhemp control were evaluated June 29 and July 11, 2017.

Table 1. Application 'A' Information – Moorhead, MN 2017

Date	June 19
Time of Day	9:30 AM
Air Temperature (F)	65
Relative Humidity (%)	54
Wind Velocity (mph)	4
Wind Direction	N
Soil Temp. (F at 6")	62
Soil Moisture	Good
Cloud Cover (%)	80
Next Rainfall (amount)	June 28 (0.3 inches)
Soybean Stage	3-trifoliolate
Common lambsquarters	6-in tall
Waterhemp	2-in tall

All soybean injury and weed control evaluations were a visual estimate of percent fresh weight reduction in the four treated rows compared to the adjacent untreated strip. Experimental design was randomized complete block with 4 replications for each trial. Data were analyzed with the ANOVA procedure of ARM, version 2017.4, software package.

RESULTS

Common lambsquarters tends to germinate in late April and early May in western Minnesota and eastern North Dakota. There was a very dense common lambsquarters population at this location even though the first flush was controlled by tillage prior to planting. Waterhemp generally emerges in mid to late May and continues to emerge following precipitation events throughout the summer. Waterhemp density was low to moderate at this location and was clearly impacted by lambsquarters competition and from fewer than normal precipitation events in June and July at Moorhead in 2017.

There was no visual soybean injury from Liberty across adjuvants (Table 2). Lambsquarters was the best indicator species of weed control in this experiment. Lambsquarters control ranged from 84 to 93% across treatments at 10 DAT and from 60 to 74% across treatments at 22 DAT. Applying Moccasin (a soil residual herbicide) with Liberty + AMS or Liberty+ET-4000 gave less lambsquarters control at 10 DAT compared to Liberty+AMS. Liberty+ET-4000+Moccasin gave similar lambsquarters control at 22 DAT compared to Liberty+AMS.

Common lambsquarters control was similar among treatments containing dry or liquid AMS adjuvants with Liberty including ET-4000. No significant differences in lambsquarters control were observed at 10 or 22 DAT from any Liberty alone+adjuvant treatments.

Moccasin was applied with Liberty to provide residual lambsquarters and waterhemp control. However, greater than 0.5 inches of precipitation is recommended to sufficiently activate Moccasin and this precipitation did not occur until August 2, or 44 days after application. Lambsquarters control from Liberty plus Moccasin, 10 DAT was less than from Liberty+AMS, suggesting the tank-mix with Moccasin may have antagonized broadleaf control.

Liberty alone with dry AMS, liquid AMS, or ET-4000, or Liberty tank-mixed with Moccasin provided perfect or near perfect waterhemp control in this experiment.

Table 2. Soybean injury and weed control from adjuvants with Liberty at Moorhead, MN in 2017.

Treatment	Rate	Appl ¹	-----June 29-----			-----July 11-----		
			soyb ²	colq	wahe	soyb	colq	wahe
	fl oz/A + adjuvant		%inj	%cntl	%cntl	%inj	%cntl	%cntl
Liberty+dry AMS ³	29 + 3 lb/a	A	0	92	100	0	69	98
Liberty + N-Pak AMS	29 + 5% v/v	A	0	89	100	0	70	100
Liberty + ET-4000	29+ 1.5% v/v	A	0	88	100	0	68	100
Liberty + ET-4000	29 + 3% v/v	A	0	91	98	0	74	100
Liberty + Moccasin ⁴ + N-Pak AMS	29 + 21 + 5% v/v	A	0	84	100	0	60	100
Liberty + Moccasin + ET-4000	29 + 21 + 1.5% v/v	A	0	85	95	0	70	98
LSD (0.05)			NS	4	5	NS	9	NS

¹Appl refers to application timing and corresponding information in Table 1.

²soyb=soybean; colq=common lambsquarters; wahe=waterhemp

³Indicates addition of ammonium sulfate (AMS) at 3 lb/A. N-Pak AMS used at 5 % v/v and provided by Winfield. ET-4000 used at 1.5% v/v and provided by MK Ag Service

⁴S-metolachlor by UPI

CONCLUSIONS

Dry AMS with Liberty provided fair to good lambsquarters control and excellent waterhemp control. N-Pak AMS or ET-4000 with Liberty generally provided similar lambsquarters control. ET-4000 at 3% v/v with Liberty tended to improve lambsquarters control compared to ET-4000 at 1.5% v/v with Liberty. Lambsquarters control from Moccasin plus Liberty, regardless of adjuvant type, was less than from Liberty+adjuvant, especially 10 DAT. The addition of Moccasin did not provide residual control. Waterhemp control from Liberty was similar among the adjuvants and tank mixes tested.

SOIL MANAGEMENT PRACTICES

NOTES

EFFECT OF COMMERCIAL FERTILIZERS AND NUTRIENT MANAGEMENT PRODUCTS ON SUGARBEET YIELD AND QUALITY DURING 2017 GROWING SEASON

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Introduction

Trial results of different fertilizer combinations, biologicals and nutrient management aids were evaluated.

Materials and methods

Roundup Ready sugarbeet cultivar with a good disease resistant package was planted on April 29 and May 4, 2017 at Downer and Ada, respectively. Field plots were laid out in a randomized complete block design with four replications. Individual plots measured 11 ft wide and 30 ft long. Sugarbeet was placed 1.25 inches deep with 5 inch row spacing. A 22 inch row spacing was used. The trial was planted in wheat residue and a fairly wet soil seedbed. Roundup herbicide was applied twice for weed control. Recommended NPK fertilizers were applied and N rate was adjusted to residual soil NO₃-N of 4 ft soil depth. The middle two rows were harvested using a mechanical harvester and sub sample sent to Crystal Sugarbeet Quality lab at Grand Forks. Downer and Ada plots were harvested on 19th September and 9th October, respectively.

Table 1. Initial soil properties

Depth	NO ₃ -N (lb/ac)			Olsen-P (ppm)	K (ppm)	Soil OM%	Soil pH
	0-6"	6-24"	24-48"	0-6"	0-6"		
Downer	19	21	30	10.5	97	3.1	8.6
Ada	16	21	16	9	74	3.6	8.1

Table 2. Mean sugar yield and quality parameters in response to different commercial products.

Trial-Agrispon (Biostimulant), Agricultural Sciences Inc. at Downer, MN					
Treatments	Tons/ac	Sugar%	RSA (lb/ac)	Gross (\$/ac)	
1. Recommended NPK	40.9	19.5	15123	2195	
2. 100%N+Agrispon@13.2oz/a @30 and 60 DAP	39.9	19.1	14422	2052	
3.90%N+ Agrispon@13.2oz/a@30 and 60 DAP	40.4	19.2	14768	2120	
4.85%N+ Agrispon@13.2oz/a@@30 and 60 DAP	38.5	19.5	14355	2100	
5.80%N+ Agrispon@13.2oz/a@@30 and 60 DAP	39.2	19.6	14580	2130	
P<0.05	NS	NS	NS	NS	
LSD	3.54	0.86	1395	257	
Conclusion- In-season side-dress twice with Agrispon at 30 and 60 DAP with 80% recommended-N had no significant difference with 100% recommended N without Agrispon application.					
Trial- Anuvia Plant Nutrients. SymTRX20S product (16-1-0-20S) and SymTRX12S were compared with MAP as replacement at Ada, MN					
No P and S check	32.89 ^B	17.9	11105 ^B	1469.76 ^C	
MAP (Full rate-105 lbs product)	38.88 ^A	18.1	13387 ^A	1810.22 ^A	
105 lbs MAP + 83 lbs AMS	35.20 ^{AB}	17.9	11988 ^B	1602.41 ^{BC}	
105 lbs MAP + 100 lbs SymTRX20S	38.48 ^A	18.2	13386 ^A	1830.17 ^A	
42 lbs MAP + 165 lbs SymTRX12S	39.14 ^A	17.8	13271 ^A	1765.04 ^{AB}	
P<0.05	0.04	NS	0.01	0.003	
LSD	4.04	0.54	1260	177.6	
Conclusion- Phosphorus and sulfur had significant positive effect on yield, recoverable sugar and gross return.					
Trial- Pursell Agri-Tech (Coated urea with three rates) at Ada, MN					
N source-Urea	35.80 ^B	17.64 ^B	12025 ^B	1582.69 ^{BC}	
N source-ESN	36.44 ^B	17.98 ^A	12515 ^{AB}	1687.87 ^{AB}	
N source-Coated urea with 2% Zn (44.5-0-0)	38.66 ^A	17.64 ^B	12999 ^A	1713.02 ^A	
N source-Resin coated urea (43.7-0-0)	37.40 ^{AB}	17.30 ^C	12273 ^{AB}	1572.66 ^C	
N source- Coated urea (44.5-0-0)	35.67 ^B	17.86 ^{AB}	12169 ^B	1629.62 ^{ABC}	
N rate-90 lb N/ac	35.85 ^B	17.52 ^B	11946 ^B	1557.88 ^B	
N rate- 120 lb N/ac	36.57 ^{AB}	17.79 ^A	12407 ^{AB}	1651.08 ^A	
N rate- 150 lb N/ac	37.95 ^A	17.75 ^A	12837 ^A	1702.55 ^A	
N Source	0.03	<0.01	0.08	0.05	
N rate	0.03	0.03	0.01	0.01	
N- Source×rate	0.57	0.01	0.78	0.57	
Conclusion- Coated urea with Zn had potential to increase yield but need more experiment to validate the finding. Significant increase in sugar and return was observed with increasing N rate from 90 to 120 lb N/ac but no difference was found between 120 and 150 lb N/ac.					

FALL VS. SPRING NITROGEN APPLICATION ON SUGARBEET PRODUCTION

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Introduction: Sugarbeet growers apply fertilizer N either in fall or spring often dependent on workload, soil compaction concerns, and without knowledge of relative N use efficiency. A risk of leaching, denitrification and erosion loss is prolonged for fall-N, but fall-N can be readily available to seed during germination and produce early vigor. Spring-N application reduces the chance of N loss due to a narrow interval between N application and uptake. Further, it is also important to know the relative response from applications of fertilizer N split between fall and spring. For sugarbeet, soil N-availability plays a significant role in yield and quality. Estimation of soil N supply as influenced by relative proportion of fall and spring fertilizer N application has potential to increase sugarbeet N use efficiency. Main objectives were to (i) determine the sugarbeet yield and quality as influenced by N application rate and timing, (ii) determine the ratio of fall and spring N application to optimize yield and quality, and (iii) compare the N use efficiency of two soil types in response to fertilizer N application and timing.

Materials and Methods: This field experiment was conducted at Crookston and Downer sites. Treatments consisted of the two fertilizer-N application rates, 130 and 190 lb N/ac; each having 0, 40, 60, 80 and 100 percent of total N applied in fall, and the balance in the spring. Trials were laid out in randomized block design with four replicates. Each plot was 30 ft long and 11 ft wide. During fall 2016, soil samples were collected from 0-6", 6-24" and 24-48" and analyzed for soil nitrate-N. Required amount of urea-N after adjusting for soil residual N were broadcast. Recommended rates of P and K fertilizer were also applied. Spring fertilizer-N treatments were applied just before planting and incorporated. Standard Roundup Ready® cultivar was planted at 22 inch row spacing. Middle two rows of six row plots were harvested and quality traits were evaluated by American Crystal Lab, Grand Forks. Planting occurred on May 4 and April 29 and harvested on September 21st and September 19th, at Crookston and Downer, respectively. Economic return was calculated using the beet payment formula used by the American Crystal Sugar. Statistical analyses were conducted using PROC ANOVA in SAS 9.4 and significant mean separation identified using Fisher's LSD at 95% significance level.

Table 1. Initial soil parameters of Downer and Crookston field experimental sites in fall, 2016.

Depth	NO ₃ -N (lb/ac)			Olsen-P (ppm)	K (ppm)	Soil OM%	Soil pH
	0-6"	6-24"	24-48"	0-6"	0-6"		
Downer	19	21	30	10.5	97	3.1	8.6
Crookston	17	30	24	42	132	3.8	8.4

Table 2. Sugarbeet yield, quality and economic return in response to urea-N application rate and timing during the 2017 growing season

N Rate	Split%		Crookston				Downer			
	Fall	Spring	Tons/ac	Sugar %	RSA (lb ac ⁻¹)	Return (\$ ac ⁻¹)	Tons/ac	Sugar %	RSA	Return (\$ ac ⁻¹)
Check	0	0	32.7 ^{AB}	18.6	11437 ^{AB}	1570.91 ^{AB}	26.8 ^B	19.6 ^A	9998 ^B	1462.22 ^B
130	0	100	31.4 ^{AB}	19.0	11417 ^{AB}	1633.69 ^{AB}	34.5 ^A	19.2 ^{AB}	12549 ^A	1795.26 ^A
	40	60	32.1 ^{AB}	18.5	11260 ^{AB}	1554.28 ^{AB}	33.9 ^A	19.1 ^{ABC}	12310 ^A	1756.14 ^A
	60	40	30.1 ^{AB}	18.5	10356 ^{AB}	1401.82 ^{AB}	33.0 ^A	19.2 ^{AB}	12044 ^A	1727.25 ^A
	80	20	31.5 ^{AB}	19.1	11511 ^{AB}	1653.68 ^{AB}	32.8 ^A	19.1 ^{ABC}	11799 ^A	1670.05 ^{AB}
190	100	0	31.7 ^{AB}	18.8	11374 ^{AB}	1605.44 ^{AB}	31.6 ^A	19.0 ^{ABCD}	11268 ^{AB}	1577.73 ^{AB}
	0	100	35.0 ^A	18.4	12209 ^A	1674.65 ^A	34.7 ^A	18.7 ^{BCD}	12171 ^A	1680.61 ^{AB}
	40	60	29.4 ^{AB}	18.6	10328 ^{AB}	1428.95 ^{AB}	34.9 ^A	18.9 ^{ABCD}	12367 ^A	1725.19 ^A
	60	40	35.4 ^A	18.6	12403 ^A	1706.51 ^A	32.2 ^A	18.8 ^{BCD}	11301 ^{AB}	1558.64 ^{AB}
LSD	80	20	33.0 ^{AB}	18.5	11674 ^{AB}	1621.62 ^{AB}	34.1 ^A	18.5 ^{CD}	11785 ^A	1599.93 ^{AB}
	100	0	26.3 ^B	18.8	9417 ^B	1326.25 ^B	34.1 ^A	18.3 ^D	11682 ^A	1569.19 ^{AB}
Significance (P<0.05)			8.09	0.68	2717	377.9	4.31	0.73	1588	258
			*	NS	*	*	*	*	*	*

At Crookston, the lowest observed yield was associated with the high fertilizer N rate (190 lb N/ac), 100 % applied in the fall. This application scheme yielded significantly less than 100% and 40% spring application of 190 lb N/ac. Spring application of 100% and 40% of 190 lb N/ac also resulted in the higher recoverable sugar per acre (RSA) and economic return; RSA and return calculations involve yield and percent sugar. At Downer, all the N fertilizer treatments resulted in significantly higher yield than the check, irrespective of N rate and application time. Sugar percent was lowest with 100% fall application of 190 lb N/ac. RSA and economic return was lowest for the check plot, although not significantly different from several other N application patterns. These results show that high N application rate in fall might reduce sugarbeet yield and percent sugar while reducing economic return.

**SUGARBEET
PHYSIOLOGY / STORAGE /
PRODUCTION PRACTICES / ECONOMICS**

NOTES

THE EFFECT OF CLOSING WHEEL AND SEED TUBE CONFIGURATION ON SUGARBEET YIELD AND QUALITY

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Introduction : Uniform seed spacing of sugarbeet plants is important to reduce weed competition and maximize sugarbeet production. The last several years, with the introduction of numerous seed tube configurations and modifications, and closing wheels, growers are asking which seed tube types, seed tube sensors or closing wheel configurations are recommended to best optimize sugarbeet yield and quality in the field and on their farms.

Objectives : With the introduction of John Deere MaxEmerge 2 Planter, many different seed tube sizes and shapes, closing wheel configurations and planter attachments are being marketed. Most were developed or used in corn and soybean growing areas of the United States, but over the year's sugarbeet growers' have adopted these different planter configurations into sugarbeet production as well. Past research has shown that the standard straight sugarbeet tube produced the highest recoverable sugar. Since that research many new and potentially improved seed tubes have been introduced. The objective of this research is to evaluate the affect these new tubes and attachments have on uniformity of seed spacing and yield and quality using various options of these planter attachments or combinations of them. Some of these seed tubes have been evaluated on the planter test stand and have not performed satisfactorily and should be evaluated in the field.

Materials and Methods: One sugarbeet field experiment was established on a Colvin silty clay loam location near Ada, MN in 2017. Planting was arranged in a randomized complete block design with five replications. Individual treatment plots measured 11 feet wide and 30 feet long. A Roundup Ready Regular Pellet sugarbeet variety with a good disease resistance package was planted on May 4/2017 with a John Deere MaxEmerge II planter. Large sugarbeet plates were used and vacuum set as recommended. Sugarbeet was placed 1.25 inches deep with 4.5-inch in-row spacing. A 22-inch row spacing was used. The trial was planted into wheat residue and a fairly wet soil seedbed. Roundup herbicide was applied twice for weed control, plots were not cultivated. Soil nitrogen, Phosphorous and Potassium levels were adjusted with fertilizer to approximately 130 lbs/acre of available residual soil test plus added fertilizer N. Treatments included in the experiment were (1) Straight tube – reg. closing wheels (2) Straight tube – modified insert regular closing wheels (3) Curved tube - regular closing wheels (4) Curved tube - modified insert - regular closing wheels (5) Precision planting tube – regular closing wheels (6) Straight tube – no insert – spiked wheels (7) Straight tube – 1 schlagel 1 smooth closing wheel. Three fungicide applications, Inspire (July 21 @ 7 fl. oz/A), Supertin/Topsin (Aug 8 @ 6 fl. oz/A & 7.6 fl. oz/A) and Proline (August 22 @ 7 fl. oz/A) were applied for Cercospora leafspot control. Total monthly rainfall for April was 1.05 inches, May 1.36 inches, June 2.91 inches, July 2.68 inches, August 1.27 inches, September 5.76 inches and October 0.69 inches. The middle two rows were treated and harvested on October 9/2017. Yield determinations were made and quality analysis performed at American Crystal Sugar Quality Tare Lab, East Grand Forks, MN.

Results and Discussion: The field or research plot area, due to nearly 11 inches of rainfall October of 2016 was only tilled once that fall. The plot area was fertilized and tilled with a field cultivator in the spring of 2017. High amounts of wheat residue and fairly wet soil conditions may have affected some germination in certain plots. Since rainfall occurred soon after planting no significant results were observed in the seed tube/planter attachment study in 2017. The measurements between plants (target spacing of 4.5 inches / Regular Pellets) obtained around the four-leaf stage of growth were analyzed and histograms of distance distributions were constructed for each treatment (Figure 1). Generally the inclusion of an insert into any style of seed tube reduces the number of plants at the 4.5 inch target spacing (histograph 3&4), as was observed in past field and grease belt tests, similar treatments with modified inserts seem to produce lower yields and recoverable sugar per acre. In 2017 the root yield was highest on the curved tube with regular closing wheels and the lowest yield treatment was the curved tube with modified insert and regular closing wheels (Table 1). The highest recoverable sugar per acre treatment was the curved tube with regular closing wheels (Table 1). Stand counts were lowest on the precision tube with regular closing wheel whereas the highest stand counts were straight tube with the modified insert although none of the treatments were significantly different.

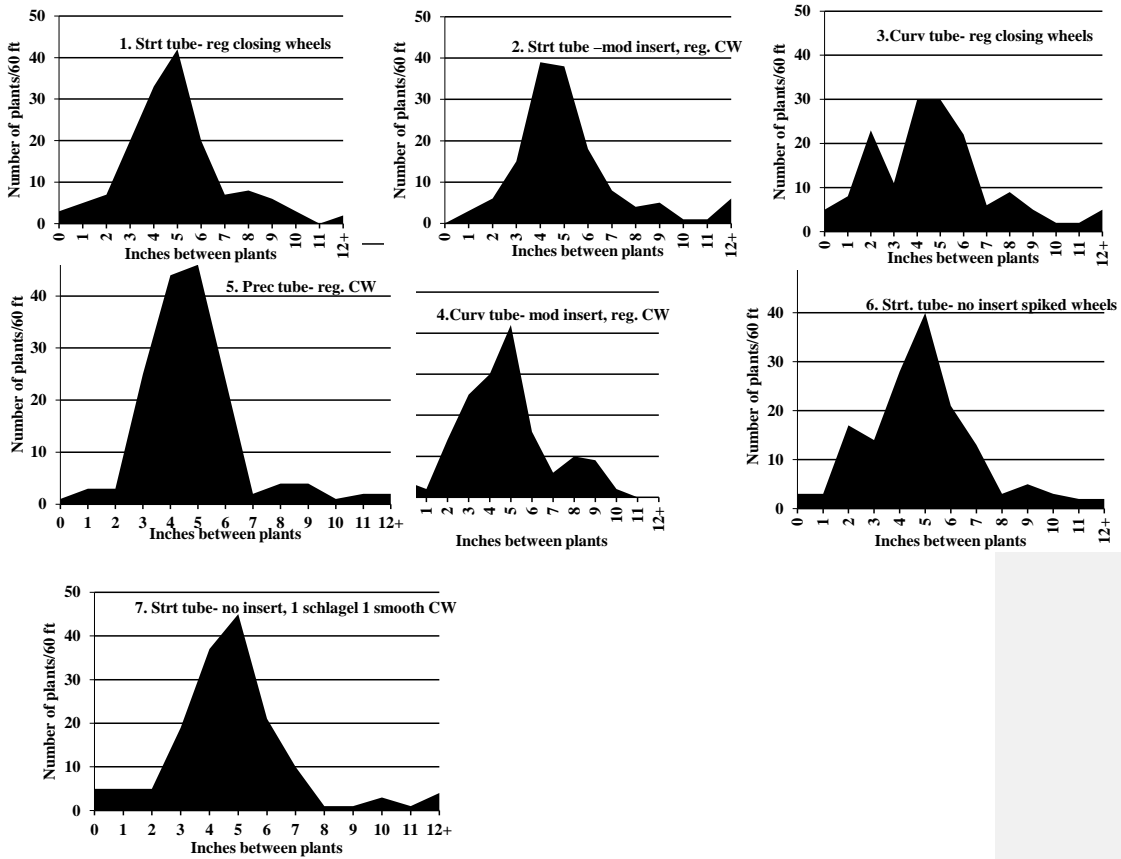
Use of different styles and combinations of closing wheels had little effect on uniformity of plant spacing. Again there was no significant difference of any of the parameters depending on what closing wheels or combination of closing wheels was used. However the two Schlagel closing wheels together had lower yield and recoverable sugar per acre than did the one smooth regular closing wheel and one Schlagel closing wheel treatment or the treatments where the standard smooth closing wheels were used (Table 1). It seems the standard smooth closing wheels tested are still as good as any other of the newer closing wheels examined in this trial for sugarbeet emergence. It is important to note that closing wheels be properly set at $\frac{3}{4}$ to 1 inch distance apart between wheels for sugarbeets and that the wheels are centered directly over the top of the planted row and also set at a proper down pressure.

This is one year and one location of data. Additional research trials both in the field and on the planter test stand comparing seed tube configurations and planter closing wheels with different size pellets should be examined to reinforce current sugarbeet grower recommendations.

Table 1. Effect of seed tube and closing wheel combinations on sugarbeet root yield, sucrose percentage, recoverable sugar production, population and gross \$ return. Ada, MN. 2017.

Treatment	Roots yield (Tons/a)	(%) Sucrose	Sim%	RSA (lb/ac)	RST (lb/ton)	Tare %	Gross (\$/acre)	Beet counts /60ft of row
1. Straight tube - regular closing wheels	36.5a	17.6a	0.94a	12161a	333a	3.5a	1526.72a	156a
2. Straight tube - mod. Insert, regular closing wheels	37.2a	17.5a	0.87a	12355a	332a	2.9ab	1547.29a	161.4a
3. Curve tube - regular closing wheels	38.1a	17.5a	0.87a	12628a	332a	3.2ab	1580.18a	161.2a
4. Curve tube - modified Insert regular closing wheels	35.5a	17.6a	0.81a	11905a	335a	2.9b	1506.00a	158.2a
5. Precision tube - regular closing wheels	36.0a	17.3a	0.92a	11764a	327a	3.3ab	1447.79a	152.2a
6. Straight tube - no insert spiked wheels	35.7a	17.5a	0.91a	11879a	333a	3.0ab	1489.81a	159.4a
7. Straight tube - no insert, 1 Schlagel 1 smooth cw.	37.5a	17.6a	0.88a	12558a	335a	3.3ab	1587.11a	158.2a
LSD (P<0.05)	NS	NS	NS	NS	NS	0.6145	NS	NS

Figure 1. Seed spacing as influenced by seed tubes and planter attachments



VARIATION IN PLANT TISSUE CONCENTRATION AMONG SUGARBEET VARIETIES

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Justification: Plant tissue analysis has increasingly been used for crops as a tool to fine tune nutrient management. Plant analysis was developed as a diagnostic tool and is generally not been used to determine nutrients to apply. For sulfur, analysis of sulfur in plant tissue is commonly determined using inductively coupled plasma emission spectroscopy (ICP) even though older data that is typically used to develop sufficiency ranges may have been determined by dry combustion. Recent work in Minnesota on corn and soybean has found differences in the assessment of sulfur concentration by ICP versus combustion. Comparison of methods of analysis for sulfur for additional crops such as sugarbeet would help to determine the accuracy of ICP and where additional research in correlation of plant tissue tests to crop yield should be conducted. If differences in the methods can be documented, it would indicate that sugarbeet growers should exercise extreme caution when interpreting plant tissue results for sulfur.

Plant tissue analysis has resulted in more recent questions on boron application than other micro-nutrients. Reports that list boron as being low typically suggest a foliar application of boron containing fertilizer sources. However, there is no documented evidence that tissue sufficiency ranges currently used are accurate and that when a low tissue boron concentration is reported that application will increase crop yield. Comparisons of yield response to tissue concentration are needed to provide evidence that a sufficiency range actually has meaning when deciding if fertilizer should be applied.

Recent surveys of corn, soybean, and hard red spring wheat plant tissue has shown significant variation in nutrient concentration when multiple hybrids/varieties are sampled in the same field at the same time. If taken at face value, tissue nutrient concentration should be reflective of soil nutrient status. Past research on corn, soybean, and wheat showed a significant portion of the variation in nutrient concentration was due to growth stage differences among hybrids/varieties at sampling. What needs to be addressed for sugarbeet is the degree of variation in tissue nutrient concentration in petioles and leaf blades for varieties grown at multiple locations and years and whether plant tissue analysis can be related to root or sugar yield. If there is significant variation in concentration that is reflective of genetics and not of yield potential, there should be a significant degree of caution when interpreting tissue results without further documentation of deficiencies with additional analysis such as soil tests.

Summary of Literature: Plant tissue analysis is being utilized more as a tool to determine whether nutrients should be applied in-season to maximize yield of crops. Plant analysis is only suggested for use for diagnosing problems that may occur in field (Kaiser et al., 2013). Fertilizer decisions should be made using soil samples which have been correlated and calibrated to crop response. Never the less, samples are being taken in fields and are being used to sell products which are likely not needed. Databases for “sufficient” levels for nutrients have been developed for use in diagnosing problem areas within fields (Bryson et al., 2014). It is not known whether these sufficiency values were generated using crop response data that documents that yield will be reduced when tissue concentrations are below the stated sufficiency level. It is more likely that the sufficiency values used currently for nutrients such as sulfur or boron are developed based on tissue concentration averages for plots where either nutrient was added but no yield response was achieved. Since both boron and sulfur can be taken up by plants in excess quantities, utilizing averages values of fertilized plots can result in the development of sufficiency ranges that are higher than what would actually be required for maximum crop yield. Most of the research previously cited has shown the effects of boron or sulfur on petiole or leaf blade boron or sulfur concentration the works have not taken the next step in correlating it to crop yield.

Understanding potential sources of variation is important when interpreting plant tissue analysis results. One major source of variation can be differences in uptake patterns among hybrids or varieties. In Minnesota, unpublished survey data for corn and soybean and published data for hard red spring wheat (Kaiser et al., 2014b) found significant variation among hybrids/varieties for a majority of the nutrients analyzed. For the wheat trials, the

majority of the variation in nutrient concentration across locations could be attributed to when the samples were collected and the stage of development of the plant at the time of sampling. For all crops the variation in yield could not be explained by one or more nutrients measured in the plant tissue. For sulfur, data collected from multiple crops has noted differences in the amount of sulfur reported in plant tissue based on how the samples are analyzed in the lab (Sterrett et al., 1987). These sources of variation indicate that varieties may have their own sufficiency range for nutrients and that ranges need to be developed based on specific laboratory methods used to determine the concentration of nutrients in plant tissue.

Objectives:

1. Compare nutrient concentration in petioles and leaf blades among varieties at three sampling times.
2. Determine if tissue nutrient concentration is predictive of root and sugar yield when sampling adequately fertilized fields.

Materials and Methods: Six sugarbeet varieties (listed below) were planted at four locations and tissue analysis samples was collected at three sampling times over the growing season. Varieties were planted in four replications at each site. Sampling times were early- to mid-June, early July, and late July to early August. The newest developed leaf was sampled. The petiole and leaf blade will be sampled at once then separated for individual analysis. All samples were dried, ground, and analyzed for nitrate N via extraction with 5% acetic acid, total N by combustion, and P, K, Ca, Mg, S, B, Cu, Fe, Mn, and Zn by ICP. A single composite soil sample consisting of six to eight cores was taken from the 0-6 and 6-24 inch depths from each site at each plant sampling date. Soil samples were analyzed using recommended procedures of N, P, K, Ca, Mg, S, B, Cu, Fe, Mn, and Zn and for pH, soil organic matter, and cation exchange capacity (CEC). Plant tissue nutrient concentration was correlated with yield and quality to determine what factors may be important for the prediction of root and sugar yield. All data was subject to an analysis of variance procedure assuming fixed effects of location, sampling time, and variety and random blocking effects.

Varieties used in the sampling trial:

Crystal RR018 – Check variety: Good disease tolerance, average yield but below average sugar.

Maribo 109 – Check variety: Good disease tolerance with average sugar content. Below average tons. Tends to have a smaller leaf canopy than other varieties.

Beta 92RR30 –Average tons and average sugar.

Beta 9475 –Good Cercospora leaf spot resistance, high yield, average sugar

Crystal M579 –High sugar content.

Crystal M509 – Good cercospora resistance, low sugar content and high yield.

Results: Sample timings were targeted to occur within three week intervals near the 50-80 day suggested for sugarbeet sampling. Actual sampling dates for Clara City, Lake Lillian, Murdock, and Renville, respectively, were 48, 44, 53, and 46 days after planting (DAP) for sample date 1; 69, 65, 74, and 66 DAP for sample date 2; and 89, 96, 96, and 87 DAP for sample date 3 (Table 1). Soil types, chemical properties, and cation exchange capacity was similar among soils at the four locations. Results for chemical soil tests for samples collected from each location at the time samples were collected are summarized in Table 2.

Root yield, sugar content per ton, and sugar content produced per acre varied among the six varieties across all four locations (Table 3). The four site average for each of the variables is given in Table 3. However, analysis indicated a significant interaction between site and variety providing evidence of variation in the ranking of varieties among the sites. Overall, root yield, sugar content, and sugar production followed anticipated patterns based on past varietal response data. Root yield and quality did vary allow for correlation between yield and quality and plant tissue concentration.

Results for the analysis of variance for leaf blade tissue concentration are summarized in Table 4. The effect of time and variety was significant for all nutrient concentrations Nutrient concentrations differed among locations for all nutrients except for calcium. The location by time interaction was significant for nearly all nutrients while the time

by variety and the three-way interaction of time x location x variety was consistently non-significant. Similar results were found for petiole concentration (Table 5).

Differences in leaf blade nutrient concentration among varieties, when averaged across time and location, are summarized in Table 6. While significant, the relative differences in plant nutrient concentrations among the varieties were relatively small. The ranking among varieties (maximum to minimum concentrations) were not consistent indicating that varieties with greater nutrient concentration were not greater for all nutrients. This indicates that plant nutrient uptake is not relatively greater for one variety versus another for all nutrients. Table 6 also lists the anticipated sufficiency range according to Bryson et al., 2014. The average for boron tissue concentration was the only instance where a concentration average was close to the low end of the sufficiency range, but the boron concentration in the leaf blade tissue did not necessarily indicate that boron was limiting yield.

Effects on all nutrient concentrations were similar for petioles (Table 7) as with leaf blades. However, the concentration of nutrients tended to be less in the petiole than in the leaf blade tissue. The major exception was potassium where the concentration was greater in the petiole than in the leaf blade. There is no identified sufficiency range for petiole tissue to compare results with established ranges.

The effect of time on macro- and micronutrient concentrations is summarized in Figures 1 and 2, respectively. Mobile nutrients (N, P, Ca, Mg) exhibited a general decrease in concentration for both leaf blade and petiole tissue over time except for potassium where the leaf blade tissue increased and the petiole potassium concentration decreased. The opposite effect was found for immobile nutrients (S, B, Cu, Mn, and Zn) where concentration increased over time. Iron did exhibit a decrease over time, but this decrease was likely due to less soil contamination on leaves later in the growing season. As more leaves developed it was less likely that rain drops would reach the soil surface resulting in splashing of soil particles onto plant tissue. Due to contamination, tissue iron concentration should not be used as a predictor of yield and quality parameters.

Simple correlation between individual nutrient concentration in the leaf blade and petiole at each sampling time and sugarbeet root yield is summarized in Table 8. There were significant positive and negative correlations among many of the nutrients studied. The only nutrient which consistently showed little to no correlation with root yield was tissue phosphorus concentration. There was not instance where a single nutrient always showed a positive correlation with root yield. For example, total nitrogen content in the leaf blade and petiole was positively correlated with root yield at T1 but was negatively correlated by T3. The greatest correlation was between leaf blade Fe and root yield ($r=0.69$). However, differences in Fe concentration early in the growing season can be impacted by the number and size of leaves on the plant which affects contamination of plant tissue by Fe splashed onto the leaves by raindrops hitting the soil.

Table 9 summarizes the correlation between plant tissue and sucrose content and Table 10 summarizes correlation with sugar production per acre. Similar to root yield, there were no instances where sugar content or yield showed a consistent correlation with any nutrient. It would be expected that if a nutrient is limiting or if yield or quality is a function of nutrient concentration then there should be consistent correlation over time between these factors and the concentration of nutrient in the plant tissue. Nutrient concentration in plant tissue does not necessarily account for variations in plant growth and differences in nutrient remobilization among varieties. The data overall indicates that some caution should be exercised when interpreting plant tissue results as a correlation between yield and quality and a concentration of a specific nutrient at a single point during the growing season does not prove that uptake of any nutrient is driving final yield or sugar production.

Correlations between individual nutrient concentrations and their respective soil test collected at the time of tissue sampling are summarized in Table 11. Significant positive correlations were found between soil test N, P, and K with leaf blade and petiole N, P, and K, respectively. The strongest correlations were for the 0 to 6-inch depth but significant positive correlations were also found between tissue N and K and the 6-24 inches N and K soil test values. For micronutrients, the only significant positive correlation was between leaf blade Cu and Zn and their soil test values and leaf blade boron and the boron soil test at 6-24 inches. Since the sites were maintained at high fertility levels it is not surprising that there was little correlation between soil test values and tissue nutrient concentration. Environmental factors such as temperature and precipitation and crop development at sampling have

been shown to influence variation in nutrient concentration among research sites for other crops. Further work is planned for the sugarbeet data but the 2017 data at four sites was not enough to conduct a correlation between outside factors and concentration. Further research is planned using the same varieties in subsequent years which will be needed to fully determine what factors can explain variations in tissue nutrient concentrations among sites and varieties.

Conclusions: The data presented in the reports is for the first year of a three-year study assessing the variation in tissue nutrient concentration among sugar beet varieties. The first year data showed that there were clear differences in yield and quality among the sugarbeet varieties used in the study. Tissue (leaf blade and petiole) nutrient concentration will vary among sugarbeet varieties sampled in the same field at the same time. The concentration of mobile nutrients will decrease while the concentration of immobile nutrients will increase when sampling the same leaf relative to the top part of the canopy over time. The decrease or increase will occur for each nutrient similar for the leaf blade and petiole sample. Due to this variation, a large range in the recommended sampling time for leaf blade samples (50-80 days after planting) should not be used. Data outlining a single sampling time is warranted to narrow down sufficiency levels for most nutrients. The data indicates that significant caution should be exercised when collecting a single sample from a well fertilized field as there is no evidence that the concentration of a nutrient in the leaf or petiole has a direct impact on yield or quality.

Literature Cited

Bryson, G.M., H.A. Mills, D.N. Sasseville, J. Benton Jones Jr., and A.V. Barker. 2014. Plant analysis handbook III: A guide to sampling, preparation, analysis, and interpretation for agronomic and horticultural crops. Micro-macro Publ. Inc., Athens, GA.

Kaiser, D.E., J.A. Lamb, and C.J. Rosen. 2013. Plant analysis sampling and interpretation. Ext. Publ. FO-3176-B. Univ. of MN Ext. St. Paul.

Kaiser, D.E., J.J. Wiersma, and J.A. Anderson. 2014b. Genotype and environment variation in elemental composition of spring wheat flag leaves. *Agron. J.* 106:324-336.

Sterrett, S.B., C.B. Smith, M.P. Mascianica, and K.T. Demchak. 1987. Comparison of analytical results from plant analysis laboratories. *Commun. Soil Sci. Plant Anal.* 18:287-299.

Table 1. Location, planting and sampling information, dominant soil series, and cation exchange capacity (CEC) for each location (CC, Clara City; LL, Lake Lillian; M, Murdock; R, Renville).

Location	Planting	Date of			Series	Soil		CEC	
		Sample 1	Sample 2	Sample 3		Texture†	Classification‡	0-6"	6-24"
								meq/100g	
CC	25-May	12-Jul	2-Aug	22-Aug	Colvin-Quam	SiCL	T Calciaquoll	31.6	25.5
LL	8-May	21-Jun	12-Jul	2-Aug	Nicollet	SiCL	A Hapludoll	33.7	28.7
M	29-Apr	21-Jun	12-Jul	2-Aug	Bearden-Quam	SiCL	Ae Calciaquoll	28.0	22.2
R	6-May	21-Jun	11-Jul	1-Aug	Chetomba	SiCL	T Endoquoll	31.1	24.4

† SiCL, silty clay loam.

‡ A, aquic; Ae, aeric; T, typic

Table 2. Summary of soil test results for samples collected with plant tissue samples at Clara City (CC), Lake Lillian (LL), Murdock (M), and Renville (R).

Time	Location	Depth in	NO ₃ -N	P	Ammonium Acetate			SO ₄ -S	DTPA					O.M.	pH			
					Ca	K	Mg		Cu	Fe	Mn	Zn	B			Cl		
					-----ppm-----													-%-
1	CC	0-6	17.5	12	5852	242	832	12	1.0	7.8	18.1	2.7	1.2	11.2	7.0	7.9		
		6-24	11.5	3	5058	153	1076	10	1.4	10.0	7.2	0.6	0.8	11.6	4.0	8.1		
	LL	0-6	31.0	36	4833	182	562	15	1.0	43.8	29.5	0.9	0.6	8.6	6.2	7.0		
		6-24	17.2	8	4679	153	548	11	1.2	43.5	17.3	0.6	0.6	8.6	4.7	7.0		
	M	0-6	9.3	8	5960	189	696	12	1.0	7.1	18.6	1.9	1.6	7.8	5.3	8.0		
		6-24	14.0	2	6330	163	869	133	1.2	6.4	8.0	0.8	1.0	6.7	3.1	7.8		
	R	0-6	6.9	8	5152	348	583	12	1.4	17.2	29.9	1.6	0.9	9.6	5.1	7.5		
		6-24	6.9	3	5581	217	608	8	1.4	9.2	11.3	0.5	0.6	7.7	3.1	7.9		
	2	CC	0-6	12.6	12	5938	249	817	11	1.0	7.3	14.7	2.7	1.3	6.9	6.6	8.0	
			6-24	3.4	3	5139	134	1016	10	1.5	8.2	7.4	0.8	0.7	7.8	4.3	8.2	
LL		0-6	16.4	35	4772	156	523	14	1.0	36.0	26.4	0.8	0.5	6.7	6.0	7.3		
		6-24	4.4	4	4480	138	543	10	1.3	40.7	16.3	0.4	0.5	6.9	4.2	7.1		
M		0-6	3.5	9	5877	163	657	11	1.1	7.6	15.3	1.9	1.5	8.0	5.2	8.1		
		6-24	3.0	3	6824	155	717	160	1.2	6.2	7.6	0.8	1.1	6.8	3.5	7.8		
R		0-6	3.4	9	5126	316	537	11	1.3	12.1	24.0	1.4	0.8	9.0	5.2	7.7		
		6-24	1.6	2	5280	147	693	6	1.4	8.2	8.2	0.3	0.6	9.8	2.9	8.0		
3		CC	0-6	4.5	16	5957	214	801	11	1.0	8.0	14.0	2.8	0.9	8.6	6.6	8.0	
			6-24	7.1	2	4835	138	1004	9	1.6	7.6	4.5	0.8	0.6	5.7	3.1	8.2	
	LL	0-6	4.3	34	4718	142	545	14	1.1	39.6	23.3	1.0	0.6	7.6	6.2	7.3		
		6-24	1.6	8	3552	135	550	12	1.2	46.0	20.7	0.4	0.7	7.4	4.7	6.8		
	M	0-6	3.5	7	5943	169	667	11	1.3	6.2	13.4	2.0	1.2	7.1	5.2	8.1		
		6-24	2.9	3	6236	156	723	61	1.3	5.8	6.5	1.0	1.1	7.5	3.5	7.9		
	R	0-6	3.4	8	5034	312	558	11	1.4	15.0	22.6	1.4	0.8	8.6	5.2	7.6		
		6-24	1.7	3	5539	188	688	8	1.4	10.0	10.0	0.4	0.6	8.4	3.2	7.8		

Table 3. Summary of analysis of variance for the main effect of sugarbeet variety by and across location. Numbers within rows which are followed by the same letter are not significantly different at $P \leq 0.10$.

Location	Variety						P>F
	Crystal RR018	Maribo 109	Beta 92RR30	Beta 9475	Crystal M579	Crystal M509	
	-----Root Yield (tons/acre)-----						
Clara City	26.8a	23.0ab	19.2b	26.6a	26.2a	25.1a	0.06
Lake Lillian	33.6b	29.0c	28.0c	33.9b	35.0b	38.2a	<0.001
Murdock	37.4b	36.7b	33.2c	37.6b	35.5bc	41.7a	<0.001
Renville	32.6b	29.1c	30.0c	34.3ab	35.0a	36.3a	<0.001
Average	32.5b	29.3c	27.8d	33.1b	32.9b	35.4a	<0.001
	-----Recoverable Sugar (lbs/ton)-----						
Clara City	266bc	278ab	272b	272bc	289a	260c	0.01
Lake Lillian	269a	268a	257b	263ab	270a	249c	<0.001
Murdock	294ab	289bc	297ab	288bc	305a	280c	0.04
Renville	285cd	295b	302a	293b	289bc	280d	<0.01
Average	280b	283b	281b	279b	288a	267c	<0.001
	-----Recoverable Sugar (lbs/acre)-----						
Clara City	7130ab	6413bc	5278c	7254ab	7561a	6555ab	0.05
Lake Lillian	9056a	7789b	7185b	8912a	9421a	9526a	<0.001
Murdock	11011b	10614b	9837c	10820b	10832b	11673	<0.01
Renville	9282bc	8590c	9067c	10014ab	10125a	10173a	<0.01
Average	9110a	8300b	7873c	9265a	9489a	9490a	<0.001

Table 4. Summary of analysis of variance for leaf blade nutrient concentration averaged across four locations in 2017 and three sampling times at each location.

Nutrient	Time (T)	Location (L)	T x L	Variety (V)	T x V	L x V	T x L x V
Nitrogen	***	***	***	***	*	**	0.17
Phosphorus	***	***	***	***	0.45	**	0.46
Potassium	***	***	***	***	***	0.16	0.17
Calcium	***	0.21	**	***	***	*	0.11
Magnesium	***	***	***	***	0.39	0.07	0.54
Sulfur	***	***	***	***	**	0.31	0.60
Boron	***	***	***	***	0.06	*	0.31
Copper	***	***	***	*	***	0.06	*
Iron	***	***	***	***	***	0.37	0.06
Manganese	***	0.08	***	***	***	0.62	0.96
Zinc	***	***	***	***	***	*	***

†Asterisks represent significance at $P < 0.05$, *, 0.01, **, and 0.001, ***.

Table 5. Summary of analysis of variance for petiole nutrient concentration averaged across four locations in 2017 and three sampling times at each location.

Nutrient	Time (T)	Location (L)	T x L	Variety (V)	T x V	L x V	T x L x V
Nitrogen	***	***	***	***	*	0.17	0.07
Phosphorus	***	**	***	***	0.34	0.06	0.07
Potassium	***	***	***	***	**	0.06	*
Calcium	***	0.23	***	***	**	**	*
Magnesium	***	***	***	***	***	***	***
Sulfur	***	***	***	***	0.23	0.18	0.40
Boron	***	***	***	***	***	0.61	0.79
Copper	***	**	***	***	0.13	0.24	0.24
Iron	***	***	***	***	*	0.96	0.98
Manganese	***	0.37	***	***	0.22	0.93	0.92
Zinc	***	0.78	***	***	*	0.65	0.81

†Asterisks represent significance at $P < 0.05$, *, 0.01, **, and 0.001, ***.

Table 6. Varietal differences in leaf blade nutrient concentration across four locations in 2017 and three sampling times at each location. Within rows, numbers followed by the same letter are not significantly different at $P \leq 0.10$.

Nutrient	Variety						Suffic. †
	Crystal RR018	Maribo 109	Beta 92RR30	Beta 9475	Crystal M579	Crystal M509	
	-----%-----						
Nitrogen	5.45a	5.02b	4.99bc	4.98bc	4.90c	5.03b	4.3-5.0
Phosphorus	0.53a	0.54a	0.45d	0.47c	0.44d	0.51b	0.45-1.1
Potassium	3.81a	3.61bc	3.47d	3.50cd	3.65b	3.41d	2.0-6.0
Calcium	0.59b	0.69a	0.67a	0.59b	0.59b	0.61b	0.5-1.5
Magnesium	0.45c	0.54a	0.56a	0.50b	0.50b	0.51b	0.25-1
Sulfur	0.39a	0.36b	0.34c	0.37b	0.36b	0.38a	0.21-0.5
	-----ppm-----						
Boron	29b	32a	31a	28c	29b	27c	31-200
Copper	26ab	24abc	24bc	23bc	27a	21c	11-40
Iron	443b	366c	436b	437b	517a	541a	60-140
Manganese	72c	85b	87b	72c	94a	77c	26-360
Zinc	47a	41d	45b	44bc	42cd	48a	10-80

†Suffic, sufficiency range identified by Bryson et al., 2014.

Table 7. Varietal differences in petiole nutrient concentration across four locations in 2017 and three sampling times at each location. Within rows, numbers followed by the same letter are not significantly different at $P \leq 0.10$.

Nutrient	Variety					
	Crystal RR018	Maribo 109	Beta 92RR30	Beta 9475	Crystal M579	Crystal M509
	-----%-----					
Nitrogen	2.50b	2.64a	2.62a	2.42b	2.42b	2.66a
Phosphorus	0.34c	0.42a	0.35c	0.33d	0.33d	0.37b
Potassium	4.28b	4.28b	4.07c	4.20bc	4.12c	4.53a
Calcium	0.34e	0.47b	0.41c	0.37d	0.41c	0.52a
Magnesium	0.26c	0.31a	0.31a	0.26c	0.27b	0.27b
Sulfur	0.12c	0.14a	0.12c	0.12c	0.12c	0.13b
	-----ppm-----					
Boron	25d	29a	27b	26bc	25.5cd	29a
Copper	9.0a	8.5b	7.7c	8.9a	7.7c	8.6ab
Iron	218c	302a	245bc	225c	262b	270b
Manganese	27d	32b	29c	26d	36a	32b
Zinc	16c	21a	16c	17c	18b	20a

Table 8. Simple correlation (r) between sugarbeet root yield and leaf blade and petiole nutrient concentration for the newest fully developed leaf sampled the third week in June, first week in July, and fourth week in July. Correlation r values when between -0.15 and 0.15 are not considered significant at $P \leq 0.10$.

	N	P	K	Ca	Mg	S	B	Cu	Fe	Mn	Zn
Time 1 Blade	0.58	0.16	0.20	0.52	0.29	0.05	0.27	-0.43	0.69	0.47	0.31
Time 1 Petiole	0.59	-0.28	0.38	0.51	0.32	0.26	0.42	-0.14	0.57	0.48	0.47
Time 2 Blade	0.11	0.03	-0.18	-0.50	-0.65	0.56	0.28	0.40	-0.42	-0.48	0.07
Time 2 Petiole	-0.46	-0.07	-0.55	-0.39	-0.64	0.01	-0.29	0.08	-0.61	-0.54	-0.35
Time 3 Blade	-0.27	-0.40	0.19	-0.11	-0.36	0.22	0.47	0.10	-0.41	-0.04	-0.50
Time 3 Petiole	-0.51	0.05	-0.38	0.03	-0.57	-0.18	0.42	-0.05	0.04	-0.12	-0.30

Table 9. Simple correlation (r) between sugarbeet sugar content (pounds per ton) and leaf blade and petiole nutrient concentration for the newest fully developed leaf sampled the third week in June, first week in July, and fourth week in July. Correlation r values when between -0.15 and 0.15 are not considered significant at $P \leq 0.10$.

	N	P	K	Ca	Mg	S	B	Cu	Fe	Mn	Zn
Time 1 Blade	-0.02	-0.27	0.41	-0.10	-0.38	-0.54	0.52	0.08	0.07	-0.05	0.33
Time 1 Petiole	-0.07	-0.44	0.30	-0.20	-0.32	-0.25	-0.18	0.15	-0.05	-0.10	0.04
Time 2 Blade	-0.47	-0.58	0.26	0.01	-0.40	-0.21	0.62	0.33	-0.43	-0.15	0.01
Time 2 Petiole	-0.62	-0.45	-0.03	-0.13	-0.62	-0.27	-0.16	0.07	-0.40	-0.26	-0.12
Time 3 Blade	-0.64	-0.59	0.57	0.46	-0.21	-0.47	0.51	0.38	-0.01	0.32	0.02
Time 3 Petiole	-0.59	-0.38	0.23	0.32	-0.59	-0.23	0.30	0.19	0.45	0.30	0.01

Table 10. Simple correlation (r) between sugarbeet sugar production (pounds per acre) and leaf blade and petiole nutrient concentration for the newest fully developed leaf sampled the third week in June, first week in July, and fourth week in July. Correlation r values when between -0.15 and 0.15 are not considered significant at $P \leq 0.10$.

	N	P	K	Ca	Mg	S	B	Cu	Fe	Mn	Zn
Time 1 Blade	0.52	0.06	0.29	0.43	0.14	-0.12	0.40	-0.35	0.64	0.41	0.38
Time 1 Petiole	0.51	-0.39	0.42	0.39	0.19	0.15	0.32	-0.09	0.49	0.40	0.43
Time 2 Blade	-0.04	-0.15	-0.10	-0.45	-0.71	0.43	0.43	0.46	-0.50	-0.47	0.08
Time 2 Petiole	-0.59	-0.20	-0.50	-0.38	-0.77	-0.07	0.30	0.11	-0.66	-0.56	-0.34
Time 3 Blade	-0.43	-0.53	0.33	0.03	-0.39	0.05	0.58	0.20	-0.37	0.05	0.43
Time 3 Petiole	-0.63	-0.07	-0.28	0.12	-0.69	-0.23	0.47	0.01	0.17	-0.02	-0.26

Table 11. Correlation between leaf blade and petiole nutrient concentration across locations and sample time with the soil test concentration for the same nutrient for soil samples collected at 0-6 and 6-24 inch soil depths.

Nutrient	Plant Part	0-6" Soil Test	6-24" Soil Test
Nitrogen	Leaf Blade	0.56	0.64
	Petiole	0.69	0.69
Phosphorus	Leaf Blade	0.52	0.26
	Petiole	0.65	0.52
Potassium	Leaf Blade	0.72	0.69
	Petiole	0.63	0.49
Calcium	Leaf Blade	-0.12	0.13
	Petiole	-0.06	0.13
Magnesium	Leaf Blade	-0.27	-0.36
	Petiole	-0.08	-0.20
Sulfur	Leaf Blade	0.40	-0.21
	Petiole	0.45	0.31
Boron	Leaf Blade	0.30	0.59
	Petiole	-0.01	-0.13
Copper	Leaf Blade	0.54	0.23
	Petiole	0.17	0.40
Iron	Leaf Blade	0.10	0.09
	Petiole	0.20	0.16
Manganese	Leaf Blade	-0.01	0.13
	Petiole	0.20	0.13
Zinc	Leaf Blade	0.67	0.44
	Petiole	0.03	0.17

Correlations between -0.50 and 0.50 are not significant at $P \leq 0.10$

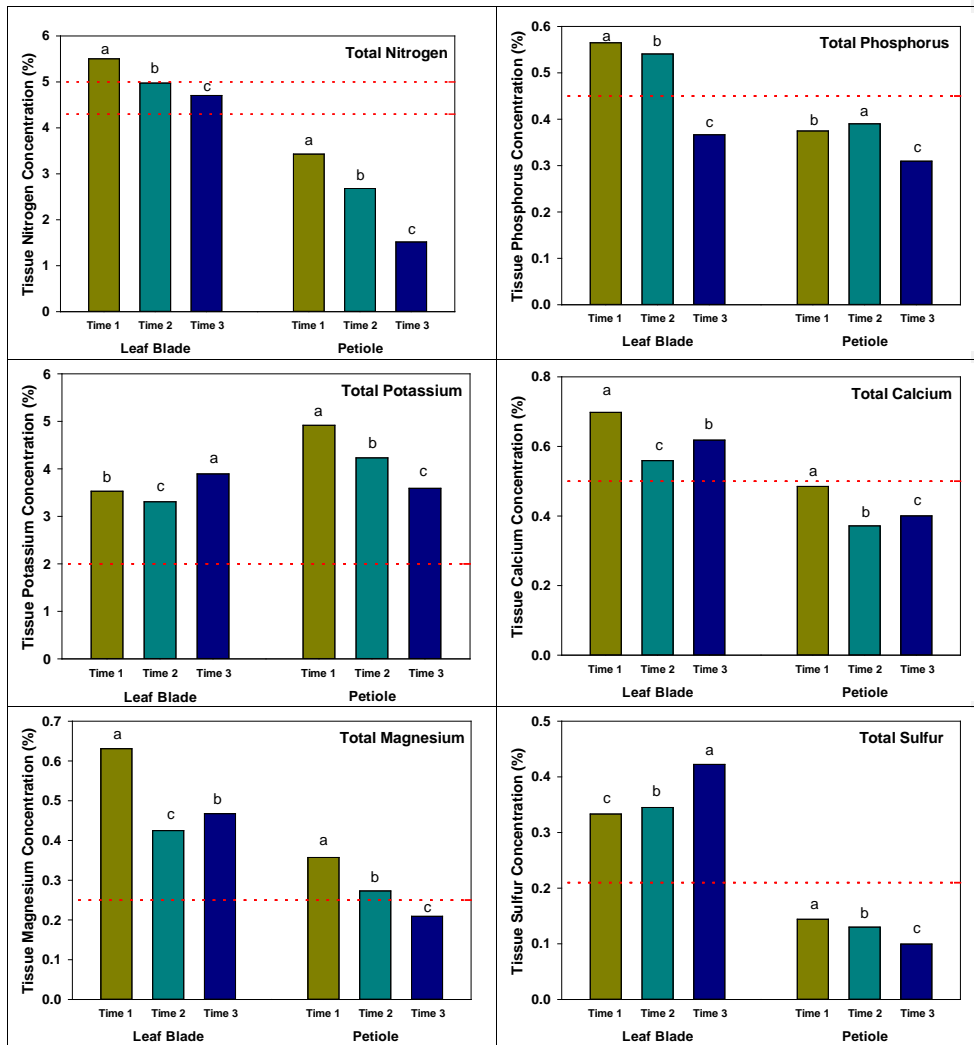


Figure 1. Summary of the impact of time on sugarbeet total macronutrient concentrations for leaf blade and petiole samples collected from six sugarbeet varieties. Letters denote significance among sampling times for leaf blade or petiole samples at $P \leq 0.10$. Horizontal dashed lines represent the upper and lower end of the sufficiency range for leaf blade samples according to Bryson et al., 2014. A single dashed line represents the low end of the sufficiency range.

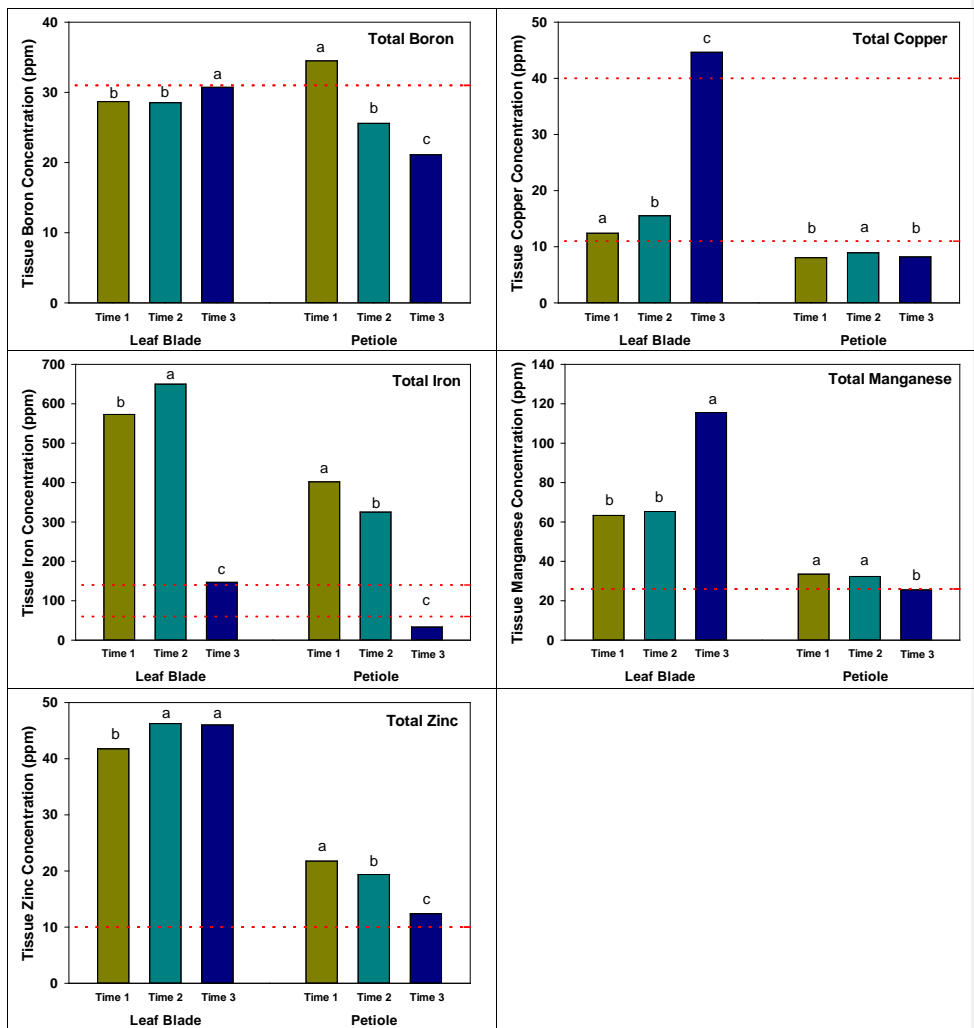


Figure 2. Summary of the impact of time on sugarbeet total micronutrient concentrations for leaf blade and petiole samples collected from six sugarbeet varieties. Letters denote significance among sampling times for leaf blade or petiole samples at $P \leq 0.10$. Horizontal dashed lines represent the upper and lower end of the sufficiency range for leaf blade samples according to Bryson et al., 2014. A single dashed line represents the low end of the sufficiency range.

EFFECT OF METHYL JASMONATE, SALICYLIC ACID, HEADLINE™ AND STADIUM™ ON ROOT YIELD, SUCROSE YIELD, AND STORAGE PROPERTIES

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INTRODUCTION

Methyl jasmonate (MeJA) and salicylic acid (SA) are increasingly being investigated for their ability to enhance yield and protect crop plants and products from environmental stress and disease (Rohwer and Erwin, 2008; Hayat et al., 2010). For a number of crop species and plant products, the application of these compounds improves resistance against a range of pathogens and insect pests and provides protection against environmental stresses including cold temperature, drought, and high soil salinity. MeJA and SA can also affect plant development, growth, and metabolism, and increases in biomass (Pelacho and Mingo-Caster, 1991; Khan et al., 2003; Loutfy et al., 2012), alterations in carbohydrate partitioning (Khodary, 2004; Wang and Zheng, 2005), and improvements in water and nitrogen use efficiency (Kumar et al., 2000; Singh et al., 2010) have been attributed to their use. Previous research established that sugarbeet roots respond to these compounds and documented the ability of postharvest MeJA treatments to reduce rot from three storage pathogens (Fugate et al., 2012; 2013). The effect of preharvest MeJA and SA treatments on sugarbeet production and storage properties, however, has not previously been examined.

Research was initiated in 2014 to determine the effects of an early season MeJA treatment, a late season MeJA treatment, or an early season SA treatment on sugarbeet root yield, sucrose content, and storage properties. A late season SA treatment was not included since preliminary studies indicated a detrimental effect of this treatment on storage properties. All treatments were applied singly or in combination with a late season Headline treatment. At the time these experiments were initiated, Headline was a commonly used fungicide for control of *Cercospora* leaf spot (causal agent *Cercospora beticola*) and was used by some for possible plant health benefits. Headline treatments were included in this study because of its potential to interact with MeJA or SA treatments due to its purported hormone-like attributes (Köhle et al., 2003).

In 2014, significant increases in root yield and recoverable sugar per acre were observed for plants that received an early MeJA treatment + a late Headline treatment (Fugate et al., 2016). Plants that received the early MeJA + Headline treatment yielded 3.5 tons acre⁻¹ more than untreated controls. Recoverable sugar per acre (RSA) for the early MeJA + Headline treatment was 1856 lbs acre⁻¹ greater than the RSA of controls. No statistically significant effects on storage traits including root respiration rate, sucrose loss in storage, invert sugar accumulation, or root firmness were observed due to early MeJA + Headline treatment.

In a 2015 repetition of this experiment, MeJA had no beneficial effects on root yield, sucrose content, or sucrose yield at time of harvest. The experiment, however, was compromised by a late season *Cercospora* infection, and Headline-containing treatments outperformed treatments without Headline. An early season MeJA + Headline treatment, however, affected storage traits, and roots that received this treatment had reduced respiration rates after 30 days in storage, reduced loss to molasses after 30 and 90 days in storage, and improved recoverable sugar per ton after 30 days in storage (Fugate et al., 2017). Postharvest Stadium™ treatments, with or without Headline treatment, were also included in the 2015 experiment. Stadium is a commercial mixture of three fungicides (fludioxonil, azoxystrobin, and difenoconazole) that is marketed for the postharvest protection of potato and other tuber and corm products. Beneficial effects due to Stadium were only observed with roots that received both Stadium and Headline treatments. Roots receiving this treatment had lower respiration rates and reduced sucrose loss to molasses after 90 days in storage, relative to controls (Fugate et al., 2017).

In 2016, the MeJA/SA/Headline field and storage experiments were repeated a third time and results of these experiments are reported here. Field and storage experiments were also carried out in 2017. For 2017 experiments,

the early MeJA treatments, with or without Headline, were expanded to include two application times and two rates. SA treatments were eliminated since beneficial effects for these treatments were not found in the previous three years.

MATERIALS AND METHODS

Field studies were conducted in Fargo, ND in 2016 and 2017 and at a location near Mooreton, ND in 2017. For Fargo experiments, seed of Crystal ACH 817 was planted using a randomized complete block design with 4 replicates. In 2016, treatments included (1) an untreated control, (2) an early season MeJA treatment, (3) a late season MeJA treatment, (4) an early season SA treatment, (5) a late season Headline treatment, (6) an early season MeJA treatment + a late season Headline treatment, (7) a late season MeJA treatment + a late season Headline treatment, (8) an early season SA treatment + a late season Headline treatment, (9) a postharvest Stadium treatment, and (10) a late season Headline treatment + a postharvest Stadium treatment. MeJA, SA, Headline, and Stadium were applied at rates of 0.01 μM , 10 μM , 9 oz/acre, and 1.6% (v/v), respectively. MeJA and SA solutions contained 10 ppm (v/v) Tween 20 and were applied as foliar sprays. For the Fargo, ND 2017 experiment, treatments included (1) an untreated control, (2) a late season Headline treatment, (3) an early season MeJA treatment of 0.01 μM , (4) an early season MeJA treatment of 1.0 μM , (5) a late season MeJA treatment of 0.01 μM , (6) a late season MeJA treatment of 1.0 μM , (7) an early season MeJA treatment of 0.01 μM + a late season Headline treatment, (8) an early season MeJA treatment of 1.0 μM + a late season Headline treatment, (9) a late season MeJA treatment of 0.01 μM + a late season Headline treatment, and (10) a postharvest Stadium treatment. Headline and Stadium were applied using the same rates as in 2016. Planting, treatment, and harvest dates for 2016 and 2017 are reported in Table 1.

The 2017 Mooreton, ND experiment was planted to two varieties, Hilleshög 4062 and Betaseed 73MN, as a split plot design with 6 replications, using varieties as the main plots. Treatments included (1) an untreated control, (2) a late season Headline treatment, (3) an early June MeJA treatment of 0.01 μM , (4) an early June MeJA treatment of 10 μM , (5) a mid-July MeJA treatment of 0.01 μM , (6) a mid-July MeJA treatment of 10 μM , (7) an early June MeJA treatment of 0.01 μM + a late season Headline treatment, (8) an early June MeJA treatment of 10 μM + a late season Headline treatment, (9) a mid-July MeJA treatment of 0.01 μM + a late season Headline treatment, and (10) a mid-July MeJA treatment of 10 μM + a late season Headline treatment. MeJA and Headline were applied as described above, and planting, treatment, and harvest dates are reported in Table 1.

Table 1. Planting, treatment, and harvest dates for the 2016 and 2017 field studies conducted in Fargo, ND and the 2017 study near Mooreton, ND. In 2016, methyl jasmonate (MeJA) was applied as an early season or late season treatment, and salicylic acid was applied as an early season treatment. In 2017, only MeJA was applied.

	2016, Fargo	2017, Fargo	2017, Mooreton
Planting date	4 May	6 June	9 May
Early season treatments			
date	29 June	13 July	8 Jun; 14 July
days after sowing	56	37	30; 66
Headline & late season treatments			
date	26 Aug	30 Aug	21 Aug
days before harvest	33	30	46
Harvest date	28 Sept	29 Sept	6 Oct

For all experiments (Fargo in 2016; Fargo and Mooreton in 2017), plants were mechanically defoliated and the roots were hand-harvested, washed, and stored at 5°C (41°F) and 95% relative humidity for up to 100 days. Respiration rate, sucrose content, loss to molasses, recoverable sugar yield, and invert sugar concentration were determined after 30 and 100 days in storage using established protocols (Campbell et al., 2012).

Data were analyzed using the GLM procedure of SAS (ver. 9.1, SAS Institute, Inc., Cary, NC) with $\alpha = 0.05$. Fisher's LSD was used to identify significant differences between treatment means.

RESULTS

In 2016, MeJA and SA treatments had little effect on root yield or sucrose yield at harvest, or storage properties at 30 or 100 days after harvest (Tables 2, 3, and 4). Relative to the untreated control, no statistical differences were noted except for an increase in root respiration rate after 30 days in storage for roots that received the early MeJA treatment (Table 3) and an increase in recoverable sugar per ton after 100 days in storage for roots that received a late MeJA treatment + Headline (Table 4). Stadium had no statistically significant effects on any storage property after 30 or 100 days storage (Tables 3 and 4).

In 2017, poor germination for the Fargo, ND field experiment required that the field be replanted. The Fargo field was replanted on 6 June and no treatments were applied until mid-July to allow plants sufficient time to become established. At harvest, no significant differences in root yield, sucrose content, recoverable sugar per ton, recoverable sugar per acre, or sucrose loss to molasses were found for any treatments (Table 5). In the 2017 Mooreton, ND experiment, all treatments had similar root yield, sucrose content, recoverable sugar per ton, and sucrose loss to molasses at time of harvest (Table 6). A significant 1149 lbs/acre increase in recoverable sugar per acre, however, was found for plants receiving a mid-June treatment of 0.01 μ M MeJA + a late August Headline treatment. Storage studies for roots from both locations are ongoing.

Table 2. Harvest data from 2016 Fargo, ND field experiment. Means within a column followed by different letters are significantly different based upon Fisher's LSD, with $\alpha = 0.05$.

Treatment	yield (tons/acre)	root weight (g/root)	sucrose (%)	loss to molasses (%)	Recoverable sugar	
					per ton (lbs/ton)	per acre (lbs/acre)
control--untreated	17.4 a	609 a	16.8 ab	1.66 a	303 ab	5287 a
early MeJA	17.5 a	712 a	16.7 ab	1.73 a	299 ab	5212 a
late MeJA	20.1 a	667 a	16.6 ab	1.68 a	298 ab	6011 a
early SA	18.0 a	701 a	16.1 b	1.88 a	285 b	5049 a
late Headline	19.5 a	690 a	17.2 ab	1.62 a	311 ab	6082 a
early MeJA + Headline	18.1 a	784 a	17.0 ab	1.41 a	311 ab	5643 a
late MeJA + Headline	18.1 a	619 a	17.6 a	1.63 a	318 a	5723 a
early SA + Headline	18.7 a	667 a	16.5 ab	1.84 a	294 ab	5491 a

Table 3. Respiration rate and invert sugar concentration 30 and 100 days after harvest (DAH) for the 2016 Fargo, ND field experiment. Means within a column followed by different letters are significantly different based upon Fisher's LSD, with $\alpha = 0.05$. Treatment means that are significantly different from the control are highlighted in red.

Treatment	respiration (mg CO ₂ /kg/h)		inverts (g/100 g sucrose)	
	30 DAH	100 DAH	30 DAH	100 DAH
	control--untreated	3.67 a	3.88 b	0.75 ab
early MeJA	3.49 a	4.82 a	0.93 ab	0.55 a
late MeJA	3.90 a	3.93 ab	0.84 ab	0.53 a
early SA	3.71 a	4.25 ab	1.02 ab	0.52 a
late Headline	3.60 a	4.18 ab	0.84 ab	0.56 a
early MeJA + Headline	3.75 a	3.80 b	0.80 ab	0.43 a
late MeJA + Headline	4.01 a	4.17 ab	0.58 b	0.53 a
early SA + Headline	3.66 a	3.96 ab	0.66 ab	0.73 a
Stadium	3.85 a	4.43 ab	1.06 a	0.54 a
Stadium + Headline	3.59 a	3.94 ab	0.67 ab	0.56 a

Table 4. Sucrose content, loss to molasses and recoverable sugar per ton 30 and 100 days after harvest (DAH) for the 2016 Fargo, ND field experiment. Means within a column followed by different letters are significantly different based upon Fisher's LSD, with $\alpha = 0.05$. Treatment means that are significantly different from the control are highlighted in red.

Treatment	sucrose (%)		loss to molasses (%)		recoverable sugar per ton (lbs/ton)							
	30 DAH	100 DAH	30 DAH	100 DAH	30 DAH	100 DAH						
control--untreated	17.4	a	17.2	ab	1.73	a	1.94	a	313	a	308	b
early MeJA	17.0	a	16.7	b	1.80	a	1.77	a	305	a	303	b
late MeJA	17.6	a	17.3	ab	1.71	a	2.04	a	317	a	308	b
early SA	17.0	a	16.7	b	1.82	a	2.01	a	304	a	301	b
late Headline	17.6	a	17.2	ab	1.76	a	1.92	a	316	a	311	ab
early MeJA + Headline	17.5	a	17.2	ab	1.84	a	1.91	a	314	a	310	ab
late MeJA + Headline	18.1	a	18.3	a	1.85	a	1.92	a	325	a	331	a
early SA + Headline	17.2	a	16.7	b	1.66	a	1.87	a	312	a	301	b
Stadium	16.8	a	16.7	b	1.85	a	2.00	a	301	a	300	b
Stadium + Headline	17.0	a	17.0	b	1.78	a	1.99	a	306	a	306	b

Table 5. Harvest and storage data for the 2017 Fargo, ND field experiment. Means within a column followed by different letters are significantly different based upon Fisher's LSD, with $\alpha = 0.05$. Determination of storage properties for these roots is in progress.

Treatment	yield		recoverable sugar		recoverable sugar		loss to molasses		sucrose content		respiration rate			
	tons/acre		lbs/acre		lbs/ton		%		0 DAH	30 DAH	30 DAH			
										%		mg CO ₂ /kg/h		
control--untreated	16.8	abcd	5552	abc	330	a	1.42	a	17.9	a	18.7	a	4.24	ab
Headline (HDL)	16.1	bcd	5052	cd	316	a	1.90	a	17.5	a	18.8	a	3.80	b
Jul MeJA, 0.01 μ M	16.7	abcd	5534	abc	331	a	1.51	a	18.1	a	18.7	a	4.31	ab
Jul MeJA, 10 μ M	16.1	bcd	5150	bcd	319	a	1.65	a	17.6	a	19.3	a	4.40	ab
Jul MeJA, 0.01 μ M + HDL	17.5	abc	5703	ab	326	a	1.54	a	17.8	a	19.3	a	4.16	ab
Jul MeJA, 10 μ M + HDL	15.9	cd	5060	cd	318	a	1.59	a	17.5	a	19.0	a	4.38	ab

Table 6: Harvest and storage data for the 2017 Mooreton, ND field experiment. Means within a column followed by different letters are significantly different based upon Fisher's LSD, with $\alpha = 0.05$. Values that are statistically different from untreated controls are highlighted in red. Determination of storage properties for these roots is in progress.

Treatment	yield		recoverable sugar		recoverable sugar		loss to molasses		sucrose content		respiration rate			
	tons/acre		lbs/acre		lbs/ton		%		0 DAH	30 DAH	30 DAH			
										%		mg CO ₂ /kg/h		
control--untreated	32.4	ab	7993	bc	293	a	1.58	a	16.2	ab	16.2	abc	4.32	a
Headline (HDL)	29.9	b	7454	c	285	a	1.66	a	15.9	ab	15.9	bc	4.21	a
Jun MeJA, 0.01 μ M	30.1	b	7497	c	292	a	1.62	a	16.2	ab	16.1	abc	4.14	a
Jun MeJA, 10 μ M	31.4	b	7644	bc	286	a	1.49	a	15.8	b	15.9	c	4.09	a
Jul MeJA, 0.01 μ M	32.4	ab	8520	ab	297	a	1.45	a	16.3	ab	16.6	a	4.03	a
Jul MeJA, 10 μ M	30.8	b	7646	bc	287	a	1.53	a	15.9	b	16.1	abc	4.09	a
Jun MeJA, 0.01 μ M + HDL	35.4	a	9142	a	299	a	1.18	a	16.4	a	16.5	ab	4.06	a
Jun MeJA, 10 μ M + HDL	33.4	ab	8438	abc	295	a	1.43	a	16.2	ab	16.3	abc	4.02	a
Jul MeJA, 0.01 μ M + HDL	31.8	ab	8045	bc	291	a	1.46	a	16.0	ab	16.3	abc	4.34	a
Jul MeJA, 10 μ M + HDL	30.8	b	7678	bc	291	a	1.53	a	16.1	ab	16.3	abc	4.19	a

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REFERENCES

- Campbell, L.G., Fugate, K.K., Smith, L.J. (2012). Effect of pyraclostrobin on postharvest storage and quality of sugarbeet harvested before and after a frost. *J. Sugar Beet Res.* 49:1-25.
- Fugate, K., Campbell, L., Eide, J., Lafta, A., Khan, M. (2017). Effect of methyl jasmonate, salicylic acid, Headline and Stadium on sucrose yield and storage properties. 2016 Sugarbeet Res Ext. Rep. 47:88-92.
- Fugate, K., Campbell, L., Eide, J., Ribeiro, W., de Oliveira, L. (2016). Effect of methyl jasmonate, salicylic acid, Headline and Stadium on sucrose yield and storage properties. 2015 Sugarbeet Res Ext. Rep. 46:73-76.
- Fugate, K.K., Ferrareze, J.P., Bolton, M.D., Deckard, E.L., Campbell, L.G. (2012). Postharvest jasmonic acid treatment of sugarbeet roots reduces rot due to *Botrytis cinerea*, *Penicillium claviforme*, and *Phoma betae*. *Postharvest Biol. Technol.* 65:1-4.
- Fugate, K.K., Ferrareze, J.P., Bolton, M.D., Deckard, E.L., Campbell, L.G., Finger, F.L. (2013). Postharvest salicylic acid treatment reduces storage rots in water-stressed but not unstressed sugarbeet roots. *Postharvest Biol. Technol.* 85:1-4.
- Hayat, Q., Hayat, S., Ifaran, M., Ahmad, A. (2010). Effect of exogenous salicylic acid under changing environment: a review. *Environ. Exp. Bot.* 68:162-166.
- Khan, W., Prithviraj, B., Smith, D.L. (2003). Photosynthetic responses of corn and soybean to foliar application of salicylates. *J. Plant Physiol.* 160:485-492.
- Khodary, S.F.A. (2004). Effect of salicylic acid on the growth, photosynthesis and carbohydrate metabolism in salt stressed maize plants. *Int. J. Agric. Biol.* 6:5-8.
- Köhle, H., Grossmann, K., Jabs, T., Stierl, R., Gerhard, M., Kaiser, W., Glaab, B., Conrath, U., Seehaus, K., Herms, S. (2003). Physiological effects of the strobilurin fungicide F 500 on plants. In: Dehne, H.W., Gisi, U., Juck, K.H., Russel, P.E., Lyr, H. (Eds.). *Modern fungicides and antifungal compounds III*. Bonn, Germany: Agroconcept GmbH.
- Kumar, P., Lakshmi, N.J., Mani, V.P. (2000). Interactive effects of salicylic acid and phytohormones on photosynthesis and grain yield of soybean (*Glycine max* L. Merrill). *Ind. J. Plant Physiol.* 165:920-931.
- Loutfy, N., El-Tayeb, M.A., Hassanen, A.M., Moustafa, M.F.M., Sakuma, Y., Inouhe, M. (2012). Changes in the water status and osmotic solute contents in response to drought and salicylic acid treatments in four different cultivars of wheat (*Triticum aestivum*). *J. Plant Res.* 125:173-184.
- Pelacho, A.M., Mingo-Castel, A.M. (1991). Jasmonic acid induces tuberization of potato stolons cultured in vitro. *Plant Physiol.* 97:1253-1255.
- Rohwer, C.L., Erwin, J.E. (2008). Horticultural applications of jasmonates: A review. *J. Hort. Sci. Biotech.* 83:283-304.
- Singh, P.K., Chaturvedi, V.K., Bose, B. (2010). Effects of salicylic acid on seedling growth and nitrogen metabolism in cucumber (*Cucumis sativus* L.). *J. Stress Physiol. Biochem.* 6:102-113.
- Wang, S.Y., Zheng, W. (2005). Preharvest application of methyl jasmonate increases fruit quality and antioxidant capacity in raspberries. *Internatl. J. Food Sci. Technol.* 40:187-195.

TERMINATING FALL-SEEDED COVER CROPS

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SUMMARY

1. Seed cereal rye at no more than 25 pounds per acre.
2. Winter wheat is easier to kill than cereal rye in the spring.
3. Use full herbicides rates. Apply SelectMax at 12 to 16 fl oz/A or PowerMax at 32 to 64 fl oz/A.
4. Apply herbicides as early as possible following cover crop green-up with consideration to the weather forecast 5 to 7 days after application.
5. Herbicides work much slower in early spring and may require 2 to 3-weeks to reach 85% burndown control.
6. Cereal rye stubble may suppress emergence and development of broadleaf weeds including nightshade, lambsquarters, and pigweed.

INTRODUCTION

Sugarbeet farmers have adopted the practice of seeding nurse crops as a companion crop with sugarbeet to reduce stand losses from wind and blowing soils. Spring-seed nurse crops are seeded at sugarbeet planting and are terminated when sugarbeet is at the 4-leaf stage or when small grains are 4 to 5 leaves (tillering). Many farmers have stated they desire to implement cover crops for a longer length of time. That is, seeding cover crops after wheat harvest and prior to sugarbeet planting or after sugarbeet harvest to reduce the chances and amount of blowing soil during the winter and early spring.

Soil health is currently a popular topic in agriculture. The topic is complicated, but the goal essentially is to protect our land resource. Cover crops in sugarbeet production is often discussed since fields are very smooth and contain very little surface crop residue after sugarbeet harvest. In addition, primary and secondary fall tillage is done on fields to be planted to sugarbeet to lessen spring tillage and to conserve moisture in advance of planting next year's sugarbeet crop. Once again, tillage often creates smooth fields that are susceptible to soil erosion, especially in dry and windy conditions.

A probe experiment was initiated in September 2016 with multiple objectives including: a) how effective is spring-applied Roundup PowerMax (glyphosate) or Select Max (clethodim) for killing fall-seeded cover crops; b) when should herbicides be applied to optimize cover crop control and sugarbeet stand establishment; and c) do cover crops provide additional benefits, for example, weed suppression? The goal was to better understand how and when fall-seeded cover crops must be terminated so that sugarbeet can be planted in mid- to late April.

MATERIALS AND METHODS

Prosper, ND. Stubble was chisel plowed following wheat harvest at the Prosper Experiment Station, near Prosper, ND. Secondary tillage was done using a Kongskilde 's-tine' field cultivator with rolling baskets on September 6, 2017. Experiment was a split plot design with 4 replications. The main (whole) plot was fall seeded cover crop; the subplot was herbicide, herbicide rate, and timing of herbicide application.

Winter wheat at 60 lb/A, cereal rye at 50 lb/A, and a mixture of oat at 40 lb/A and tillage radish at 5 lb/A were spread by hand across respective whole plots in each replication and shallow tilled to incorporate seeds into soil on September 6, 2017. One main plot was left with no cover crop.

Select Max at 6 fl oz/A + 1.5 pt/A methylated seed oil (MSO) and Roundup PowerMax at 28 fl oz/A + Prefer 90 non-ionic surfactant (NIS) at 0.25% v/v with ammonium sulfate (N-Pak-AMS) at 2.5% v/v were applied as treatments on April 17, April 21, and April 29, 2017 when winter wheat was 5, 5, and 7-inches, respectively, and cereal rye was 8, 9, and 10 inches, respectively (Table 1). All herbicide treatments were applied with a bicycle sprayer (without the customary hood) in 17 gpa spray solution through 110002 Turbo TeeJet nozzles pressurized with CO₂ at 40 psi across

plots. Percent visual control or burndown of winter wheat and cereal rye was evaluated on October 27, 2016 and April 13, April 29, May 5, May 12, and May 23, 2017.

Table 1. Application Information – Prosper, ND 2017

Date	April 17	April 21	April 29
Time of Day	3:00 PM	3:00 PM	4:00 PM
Air Temperature (F)	49	62	58
Relative Humidity (%)	33	38	16
Wind Velocity (mph)	4	2	6
Wind Direction	NW	W	NE
Soil Temp. (F at 6")	54	56	46
Soil Moisture	Good	Good	Good
Cloud Cover (%)	80	10	30
Winter Wheat	5 inch	5 inch	7 inch
Cereal Rye	8 inch	9 inch	10 inch

'SV36272RR' sugarbeet, treated with NipsIt Suite, Tachigaren at 45g per unit, and Kabina at 7g per unit, was seeded in 22-inch rows at 60,560 seeds per acre on May 26, 2017. Roundup PowerMax at 32 fl oz per acre + ClassAct NG at 2.5% v/v was applied on June 19 and July 10, 2017 to control weed escapes in the trial.

Renville, MN. Cereal rye at 100 lb/A was seeded into a preharvest sugarbeet field on September 12, 2016. Rye was harrowed into the soil following seeding using a field cultivator. Roundup PowerMax at 22, 32, and 64 fl oz/A plus Class Act NG at 2.5% v/v or SelectMax at 6 fl oz/A plus Class Act NG at 2.5% v/v was applied to the center 7.3 ft of an 11 ft plot by 30 feet long on April 7, 2017. Herbicide was applied with a bicycle sprayer at 17 GPA through TeeJet 8002XR nozzles at 40 psi.

Evaluations were a visual assessment of cereal rye control (visual reduction in ground cover) on April 17, April 21 and April 28, 2017.

Data were analyzed with the ANOVA procedure of ARM, version 2017.4 software package.

RESULTS AND DISCUSSION

Cover Crop Establishment and Overwintering at Prosper. A visual assessment of cover crop establishment was collected on October 27, 2016. In general, cover crop emergence and percent visual ground cover was very good, perhaps exceeding expectations (Table 2). Favorable moisture conditions and warm temperatures in the fall of 2016 promoted cover crop growth. Cereal rye growth was most uniform while winter wheat was the least uniform. Tillage radish emerged but were small, ranging from 0.5 to 1 inch in diameter and 2 to 4 inches long. Ground cover in the no-cover crop main plot was a uniform cover of volunteer spring wheat.

Table 2. Percent visual ground cover and range of observations across replications, October 27, 2016 at Prosper, ND

Cover Crop	Visual Ground Cover	Range of Visual Ground Cover Observations
	%	%
Winter Wheat	60	40-70
Cereal Rye	85	80-90
Oat and Tillage Radish	68	50-80
No Cover Crop ¹	38	30-40

¹Block contained volunteer wheat from previous crop

Cover crop establishment was evaluated April 6 and April 13, 2017 following snow melt. On April 6, the cereal rye whole plots were greening up, but there was very little visual evidence of living winter wheat. Spring green-up and early season growth changed quickly in one week. On April 13 the number of green cereal rye or winter wheat plants per meter square were counted and a visual assessment of green-up was taken in 1m² quadrats at three evenly spaced points within the cover crop whole plot. Cereal rye ground cover and uniformity were greater than winter wheat which

may have suffered some winter-kill damage (Table 3). However, the number of rye or winter wheat plants per m² were similar. This may be attributed to the aggressive behavior of cereal rye which was well tillered on April 13 and was in general, much more robust than winter wheat.

Seeding rates were determined from the literature and through personal communication. In both cases, there was a wide range of opinions regarding seeding rates. Cereal rye seeding rate of 50 lb/A was much too great as the rye whole plots resembled sod.

Table 3. Percent visual ground cover, number of plants per square meter and range of observations across replications, April 13, 2017 at Prosper, ND

Cover Crop	Visual Ground Cover	Range of Visual Ground Cover Observations	Number of Plants per Square Meter	Range of count Observations per Square Meter
	%	%	Number	Number
Winter Wheat	46	0-80	16	0-44
Cereal Rye	73	40-100	17	6-32

Cereal Rye and Winter Wheat Control at Prosper. Percent visual control or burndown was collected April 29 (data not presented), May 5, May 12, and May 23, 2017. In general, winter wheat burndown was faster than cereal rye. Roundup PowerMax at 28 fl oz/A applied on April 17 or April 21 controlled 70% or 75% winter wheat on May 5 or 18 or 14 DAT (days after treatment), respectfully. PowerMax gave only 45% and 25% cereal rye control (Table 4). Winter wheat control from PowerMax ranged from 83 to 98% control by May 12 or 17 to 25 DAT. A minimum of 90% burndown control of cereal rye did not occur until May 23 or 32 to 28 DAT and following PowerMax application on April 21 or April 25. Roundup PowerMax provided greater overall cereal rye and winter wheat control and speed of kill than SelectMax. However, herbicide rate for both Roundup PowerMax and SelectMax probably were not sufficient, especially for early spring application. These results support the recommendation of full herbicide rates, including PowerMax at 32 to 43 fl oz/A and SelectMax at 12 to 16 fl oz/A. The use of appropriate adjuvants will also accentuate herbicide efficacy.

Cereal rye early-season growth and development was very rapid. Herbicide burndown application should be timed as early as possible or immediately after green-up in early spring. However, application timing is a compromise between growth and development of target species and environmental conditions. For example, the April 17 application was followed by wintry weather including 2 to 3 inches of snow and low temperatures. The cereal rye and winter wheat control data suggests herbicides and cover crop efficacy including speed of kill were influenced by environmental conditions.

Table 4. Percent visual cereal rye and winter wheat control, across herbicide, application timing, and evaluation date, Prosper, ND

Herbicide ¹	Appl Date	May 5		May 12		May 23	
		c rye %	w wheat %	c rye %	w wheat %	c rye %	w wheat %
PowerMax	April 17	55 cd	70 ab	65 c	83 b	75 c	85 b
Select Max	April 17	20 ef	45 d	5 f	60 c	0 g	20 f
PowerMax	April 21	60 bc	75 a	83 b	98 a	100 a	99 a
Select Max	April 21	5 g	25 e	25 e	50 d	0 g	55 d
PowerMax	April 25	20 ef	30 e	70 c	88 b	98 a	100 a
Select Max	April 25	0 g	10 fg	20 e	25 e	20 f	45 e
LSD (0.05)		10		7		7	

¹Roundup PowerMax at 28 fl oz/A + Prefer 90 NIS at 0.25% v/v + N-Pak AMS at 2.5% v/v; Select Max at 6 fl oz/A + Noble MSO at 1.5 pt/A
Cereal Rye Control at Renville. Cereal rye control (burndown) was dependent on Roundup PowerMax rate and number of days between application and evaluation. Roundup PowerMax at 64 fl oz/A gave 95% cereal rye control 21 DAT (Table 5). Cereal rye control from PowerMax at 32 fl oz/A was similar to control from PowerMax at 64 fl oz/A on April 21 and April 28 or 14 and 21 DAT. However, numbers of days to achieve similar numeric control from PowerMax at 64 fl oz/A was approximately 7 days faster than from PowerMax at 32 fl oz/A. PowerMax at 64 fl oz/A provided greater rye burndown control than PowerMax at 22 fl oz/A. Cereal rye control from SelectMax at 6 fl oz/A was less than control from PowerMax, regardless of rate.

Table 5. Percent visual cereal rye control, across herbicide, herbicide rate, and evaluation date, Renville, MN

Herbicide ¹	Herbicide Rate	April 17	April 21	April 28
	fl oz/A	-----% control-----		
PowerMax	22	41 b	61 b	76 b
PowerMax	32	41 b	73 a	85 ab
PowerMax	64	69 a	86 a	95 a
SelectMax	6	10 c	17 c	31 c
LSD (0.05)		16	12	10

¹Roundup PowerMax at 28 fl oz/A + Class Act NG at 2.5% v/v; SelectMax at 6 fl oz/A + Class Act NG at 2.5% v/v

Weed Suppression at Prosper. There is some evidence suggesting cover crop stubble suppresses germination and emergence of broadleaf weeds. Percent weed suppression across cover crop and burndown herbicide combination was collected visually on June 6 and June 12 and was collected using stand counts per unit area on June 12. Cereal rye stubble suppressed emergence and growth of hairy nightshade, lambsquarters, and pigweed better than winter wheat stubble or the no stubble blocks, but weed suppression was confounded by incomplete cover crop burndown control in some treatments (Table 6). Cover crop termination date did not affect weed suppression from cereal rye but delaying winter wheat termination to April 25 improved weed suppression. Winter wheat did not suppress hairy nightshade, lambsquarters, and pigweed. However, there were numeric differences in suppression when wheat cover crop termination date was delayed from April 21 to April 25 and from April 17 to April 21. Both visual and stand count data (data not presented) collected June 6 and 12 suggest that cereal rye stubble suppresses broadleaf weeds even after rye was killed with April applications of Roundup PowerMax.

Table 6. Visual weed suppression from cereal rye and winter wheat stubble, by cover crop termination date

Cover Crop	Cereal rye	Winter wheat
	%	%
April 17	91 a	39 c
April 21	96 a	51 c
April 25	93 a	71 b
No Cover Crop	55 b	54 b
LSD (0.05)		18

ENTOMOLOGY

NOTES

TURNING POINT® SURVEY OF SUGARBEET INSECT PEST PROBLEMS AND MANAGEMENT PRACTICES IN MINNESOTA AND EASTERN NORTH DAKOTA IN 2016

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Attendees of the 2017 Sugarbeet Winter Grower Seminars answered survey questions about their 2016 insect pest management issues and associated production practices in a live polling questionnaire that was conducted using Turning Point® interactive personal response technology. Initial questioning identified the county in which the majority of each respondent's sugarbeet crop was produced (Tables 1, 2, and 3).

Table 1. 2017 Fargo Grower Seminar – number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Barnes	3	9
Cass	7	21
Clay	11	32
Norman ¹	8	24
Richland	1	3
Traill	3	9
Wilkin ²	1	3
Total	34	100

¹Includes Mahnomon County

²Includes Otter Tail County

Table 2. 2017 Grafton Grower Seminar – number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Grand Forks	1	2
Kittson	4	7
Marshall	5	9
Pembina	19	35
Polk	1	2
Walsh	23	43
Other	1	2
Total	54	100

Table 3. 2017 Wahpeton Grower Seminar – number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Cass	2	4
Clay	3	7
Grant	5	11
Otter Tail	1	2
Richland	7	16
Stevens	1	2
Traverse	5	11
Wilkin	21	47
Total	45	100

NOTE: acreage estimates provided in this report do not include data from the Willmar Seminar location because that survey did not include questions involving insect pest incidence or insect pest management practices.

An estimated 99,491 acres were reported on by a total of 128 respondents at the Fargo, Grafton, and Wahpeton Winter Grower seminars (Table 4). The majority (35%) of respondents reported growing sugarbeet on between 300 and 599 acres in the 2016 production season. An additional 18% produced sugarbeet on 100 to 299 acres and another 32% grew the crop on a reported range of between 600 and 1,499 acres in 2016.

Table 4. Ranges of sugarbeet acreage operated by respondents in 2016.

Location	Number of Responses	Acres of sugarbeet									
		<99	100-199	200-299	300-399	400-599	600-799	800-999	1000-1499	1500-1999	2000+
		-----% of responses-----									
Fargo	33	3	0	15	18	18	6	9	12	6	12
Grafton	53	6	15	11	9	17	9	11	9	2	9
Wahpeton	42	2	7	2	10	33	17	12	10	5	2
Total	128	4	9	9	12	23	11	11	10	4	8

From a total of 127 respondents in the Fargo, Grafton, and Wahpeton Grower seminars, 26% reported that the sugarbeet root maggot was their worst insect pest problem during the 2016 growing season (Table 5). The root maggot was reported as the worst insect pest problem by respondents at both the Fargo (21%) and Grafton (47%) locations. Other significant insect pest problems reported included cutworms (6 and 7% of respondents at Fargo and Wahpeton, respectively), wireworms (6 and 5% of respondents at Fargo and Wahpeton, resp.), and white grubs (5% of respondents at the Wahpeton seminar).

Table 5. Worst insect pest problem in sugarbeet in 2016.

Location	Number of Responses	Springtails	Cutworms	Lygus bugs	Wireworms	Root maggot	White grubs	None
Fargo	33	3	6	3	6	21	0	61
Grafton	51	0	0	0	0	47	2	51
Wahpeton	43	2	7	0	5	5	5	77
Total	127	2	4	1	3	26	2	62

The majority (47%) of respondents that attended the Fargo, Grafton, and Wahpeton Winter Grower Seminars indicated that they used seed treated with Poncho Beta insecticidal seed treatment, whereas Cruiser and NipsIt Inside seed treatment insecticides were only reported as being used by 5 and 3% of respondents, respectively (Table 6). A relatively large number (45%) of respondents at these events reported that they did not use any insecticidal seed treatment in 2016. Most of the use of seed treatment insecticides was reported by attendees of the Fargo and Grafton Grower Seminars.

Table 6. Seed treatment insecticide use for sugarbeet insect pest management in 2016.

Location	Number of Responses	Poncho Beta	Cruiser	NipsIt Inside	None
Fargo	30	57	3	3	37
Grafton	49	67	8	6	18
Wahpeton	40	15	3	0	82
Total	119	47	5	3	45

Planting-time granular insecticides were used by a combined average of 29% of grower attendees of the Fargo, Grafton, and Wahpeton seminars (Table 7). An overall average of 24% of growers at these meetings reported using Counter 20G at planting time, whereas only 5% of attendees reported applying Lorsban 15G for planting-time protection of their sugarbeet crop from insect pests. Thirty-one percent of Fargo seminar respondents reported using Counter 20G at planting time, whereas 21 and 22% of respondents at the Grafton and Wahpeton seminars, respectively, reported applying Counter 20G at planting to protect their sugarbeet crop. Overall, 66% of respondents across all three grower seminars reported that they did not use a granular insecticide product at planting in 2016.

Table 7. Planting-time granular insecticide use for sugarbeet insect pest management in 2016.

Location	Number of Responses	% of responses				
		Counter 20G	Lorsban 15G	Thimet 20G	Other	None
Fargo	29	31	3	0	0	66
Grafton	47	21	2	9	2	66
Wahpeton	40	22	10	0	3	65
Total	116	24	5	3	2	66

Overall results from this survey across all three seminar locations indicated that 22% of all respondents used low to moderate rates (5.25 to 7.5 lb product/ac) of Counter 20G, while only 6% used the high rate of this material (Table 8). At the Fargo seminar, the majority of respondents that reported using Counter 20G indicated that they applied it at the 7.5-lb rate, whereas, at the Grafton seminar, the majority reported using Counter at its high (9 lb product/ac) rate in 2016. The majority of grower respondents at the Fargo seminar location that reported using Lorsban 15G at planting time indicated that they applied it at the low labeled rate of 6.7 lb product/ac. Attendees of the Grafton seminar that reported using Lorsban 15G were split evenly between using it at its high (13.4 lb/ac) and low (6.7 lb) application rates. At the Wahpeton location, 100% of attendees that reported using Lorsban 15G indicated that they applied it at a moderate rate of 10 lb of product per acre.

Table 8. Application rates of planting-time granular insecticides used for sugarbeet insect pest management in 2016.

Location	Number of Responses	Counter 20G			Lorsban 15G			Other	None
		9 lb	7.5 lb	5.25 lb	13.4 lb	10 lb	6.7 lb		
Fargo	31	0	23	16	0	0	3	6	52
Grafton	49	14	4	8	2	0	2	0	69
Wahpeton	42	0	12	12	0	7	0	5	64
Total	122	6	11	11	1	2	2	3	63

Most of the postemergence insecticide use for sugarbeet root maggot management was reported by growers that attended the Grafton Growers Seminar (Table 9). At that location, the majority (44%) of respondents indicated that they used either Lorsban Advanced or Lorsban 4E (or a generic equivalent material), and an additional 13% reported using Thimet 20G. Similarly, the majority of respondents at the Fargo seminar that reported using a postemergence insecticide for root maggot control indicated that they used either Lorsban Advanced or Lorsban 4E (or a generic equivalent material). An average of 60% of the respondents across all locations indicated that they did not apply a postemergence insecticide to manage the sugarbeet root maggot. The majority of those respondents were attendees of the Fargo and Wahpeton locations, where a respective 82 and 73% of the respondents reported no use of a postemergence insecticide for root maggot control.

Table 9. Postemergence insecticide use for sugarbeet root maggot management in 2016.

Location	Number of Responses	Lorsban 4E	Lorsban Advanced	Mustang	Asana	Other liquid	Counter 20G	Lorsban 15G	Thimet 20G	None
		% of responses								
Fargo	34	9	6	3	0	0	0	0	0	82
Grafton	45	40	4	4	0	0	0	2	13	36
Wahpeton	40	8	0	12	0	0	2	2	2	73
Total	122	20	3	7	0	0	1	2	6	60

Overall satisfaction with insecticide applications made for root maggot management was rated as good to excellent by 78% of respondents when averaged across the Fargo, Grafton, and Wahpeton seminar locations (Table 10). At the Fargo location, 82% of respondents rated their satisfaction with root maggot management efforts as being good to excellent. Similarly, 91% of respondents at the Grafton location rated their satisfaction with root maggot management practices as being good to excellent. The percentages of respondents that indicated good to excellent satisfaction with performance of root maggot management practices were lower at the Wahpeton location; however, that is likely a product of a large portion (55%) of those respondents responding with an answer of “unsure”.

Table 10. Satisfaction with insecticide treatments for sugarbeet root maggot management in 2016.

Location	Number of Responses	Excellent	Good	Fair	Poor	Unsure
Fargo	16	44	38	0	0	19
Grafton	32	19	72	6	0	3
Wahpeton	11	27	9	0	9	55
Total	59	27	51	3	2	17

At the Fargo Growers Seminar, 16% of respondents indicated that their insecticide use in sugarbeet had decreased in comparison to the previous five years, and 74% of respondents at that location reported no change in insecticide use (Table 11). However, 33% of grower attendees at the Grafton location indicated that their insecticide use had increased when compared to the previous five years. This finding is probably due to recent the increases in root maggot populations that reached extremely high levels in 2015 and continued into the 2016 growing season. At the Wahpeton seminar location, 49% of attendees indicated that their insecticide use either did not change or had decreased in comparison to the previous five years. Attendees at that location also had the highest percentage (44%) of no reported insecticide use in 2016.

Table 11. Insecticide use in sugarbeet during 2016 compared to the previous 5 years.

Location	Number of Responses	Increased	Decreased	No Change	No Insecticide Use
Fargo	31	3	16	74	6
Grafton	49	33	6	57	4
Wahpeton	41	7	15	34	44
Total	121	16	12	54	18

At the Fargo Sugarbeet Growers Seminar, 47% of attendees indicated using an online decision-making tool for sugarbeet insect pest management in 2016 (Table 12). Similarly, 66% of the attendees at the Grafton location indicated that they used some form of online information or tool for assistance or guidance with their insect management decision-making procedures. Conversely, only 12% of the attendees at the Wahpeton seminar location indicated use of an online decision-making tool. The majority of respondents at the Grafton location that indicated use of an online insect management tool responded that they used NDSU's online posting of root maggot fly counts for guidance with management decisions. An additional 19% of the Grafton attendees reported using the NDSU root maggot model application on the North Dakota Agricultural Weather Network (NDAWN) website.

Table 12. Use of online decision-making tools for sugarbeet insect management in 2016.

Location	Number of Responses	NDSU					
		Crop & Pest Report	NDAWN Root Maggot Model	Root Maggot Fly Counts (online)	Root Maggot Mobile App	Other	None
-----% of responses-----							
Fargo	38	10	10	3	0	24	53
Grafton	62	5	19	31	3	8	34
Wahpeton	41	2	2	2	0	5	88
Total	141	6	12	15	1	11	55

SUGARBEET ROOT MAGGOT FLY MONITORING IN THE RED RIVER VALLEY IN 2017

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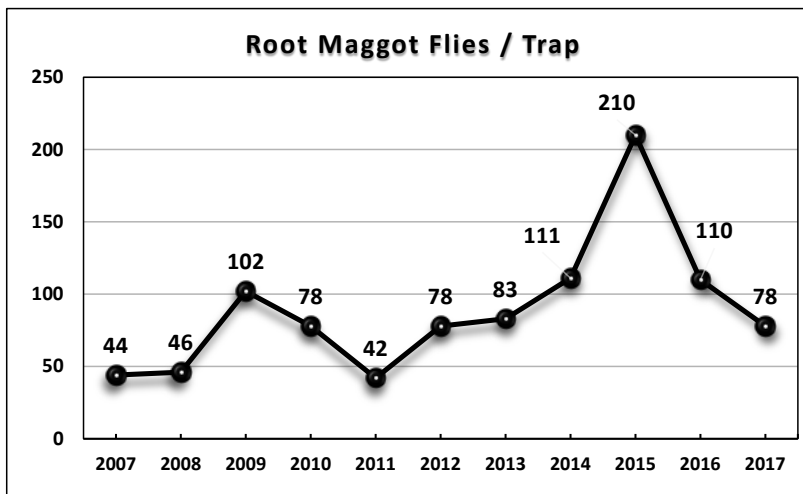
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Sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder), fly activity was monitored at 36 grower field sites throughout the Red River Valley during the 2017 growing season. The monitoring program was a result of a collaborative effort between the North Dakota State University Entomology Department and the Minn-Dak Farmers Cooperative. Additionally, the project was jointly funded by the Sugarbeet Research & Education Board of Minnesota and North Dakota and the American Crystal Sugar Company.

For the second consecutive year, fly activity in 2017 were significantly lower than those in 2015, which was the third-highest activity year in the past decade (Figure 1). Valley-wide fly counts for the whole season were about 63% lower than in 2015. This may suggest that control efforts between 2015 and 2017 were effective in reducing overall population levels throughout the Valley. However, it should be noted that a severe hailstorm occurred just two days before expected peak fly activity at South St. Thomas Township (TWP), which usually has some of the highest fly activity levels in the region. The storm is estimated to have killed 40 to 60% of the SBRM fly population at that sampling site and in the surrounding area within the path of the storm. This severe weather event likely contributed to the overall reduction in SBRM fly counts shown in Fig. 1.

Figure 1. Yearly averages of sugarbeet root maggot flies captured on sticky-stake traps



(Blickenstaff and Peckenpaugh, 1976) in the Red River Valley from 2007 to 2017.

The highest levels of SBRM fly activity were observed near Merrifield/Grand Forks, St. Thomas, and Thompson, ND, as well as Euclid and East Grand Forks, MN. Moderately high levels of activity were recorded near Auburn, Bathgate, Glasston, and Reynolds, ND, and also near Crookston, MN. Fly activity in most of the southern portion of the Valley remained at relatively low or undetectable levels throughout the growing season.

Figure 2 presents SBRM fly monitoring results from three representative sites (i.e., Reynolds, St. Thomas, and Grand Forks [Merrifield], ND). The onset of root maggot fly activity began a few days later than average, with the first captures of flies on sticky stakes occurring on June 2. Significant increases in fly activity occurred during the second week of June, with main peaks in activity occurring between June 7th and 11th in most sites.

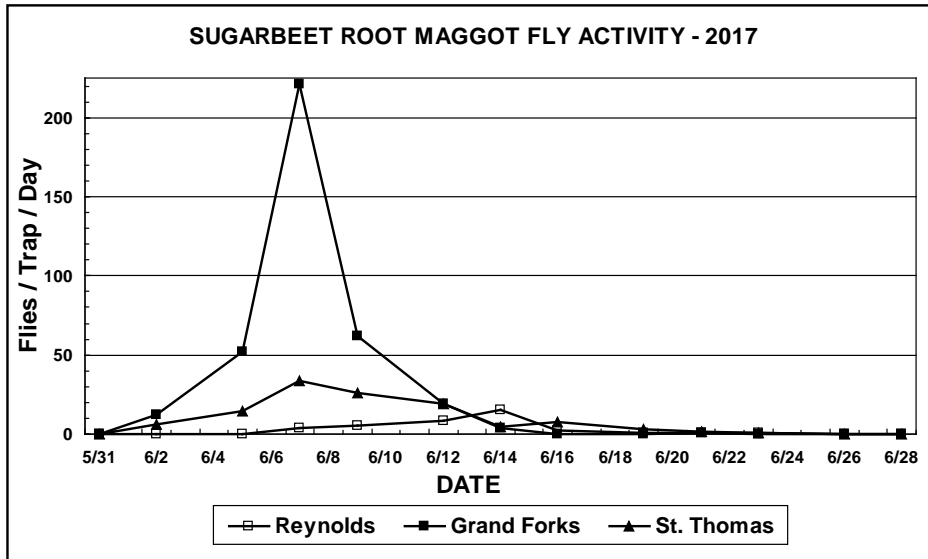


Fig. 2. Sugarbeet root maggot flies captured on sticky-stake traps at selected sites in the Red River Valley.

After the larval feeding period ended in August, all 36 fly monitoring sites were rated for sugarbeet root maggot feeding injury in accordance with the 0-9 scale of Campbell et al. (2000). This is carried out on an annual basis as a means of determining whether fly outbreaks and larval infestations were managed effectively.

Root maggot larval feeding injury in most fields was again lower than that observed in the past few years. The highest root injury ratings were observed near Grand Forks (Grand Forks TWP), Merrifield (Brenna TWP), Thompson (Walle TWP), St. Thomas (S. St. Thomas TWP), and Auburn (Martin TWP), ND, with respective average damage ratings of 2.7, 2.0, 1.5, 1.3, and 1.2. Areas where low to moderate feeding injury levels were observed, but still could produce isolated damaging infestations next year included Glasston and Reynolds, ND, and Argyle, Crookston, E. Grand Forks, and Euclid, MN. Feeding injury observed in all other sampled fields was very low. The nearly universal low root injury in those fields, despite the occurrence of moderate to high fly activity levels earlier in the season, suggests that control efforts were effective at managing SBRM infestations in 2016 and 2017. Careful monitoring of fly activity in moderate- and high-risk areas (see Forecast Map [Fig. 1] in subsequent report) will be critical in 2018 to detect unanticipated flare-ups of SBRM fly activity and to prevent economic loss. Vigilant monitoring and effective SBRM management on an individual-field basis by sugarbeet producers may also help prevent significant population increases from one year to another because even moderate levels of root maggot survival in one year can be sufficient to result in economically damaging populations in the following year.

References Cited:

Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000. Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. J. Sugar Beet Res. 37: 57-69.

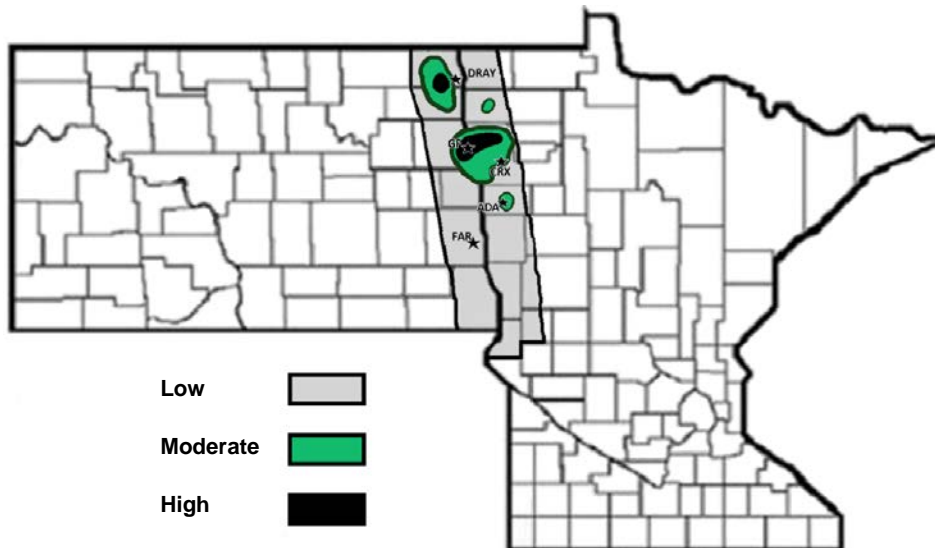
Blickenstaff, C.C., and R.E. Peckenpaugh. 1976. Sticky-Stake traps for monitoring fly populations of the sugarbeet root maggot and predicting maggot population and damage ratings. *J. Am. Soc. Sugar Beet Technol.* 19: 112-117.

SUGARBEET ROOT MAGGOT FORECAST FOR THE 2018 GROWING SEASON

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The 2018 sugarbeet root maggot (SBRM) forecast map for the Red River Valley is shown in the figure below. Areas at highest risk include rural Grand Forks, Merrifield, St. Thomas, and Thompson, ND, as well as East Grand Forks and Euclid, MN. Moderate risk is expected near Auburn, Bathgate, Buxton, and Reynolds, ND, and in the vicinity of Argyle and Climax, MN. Other areas that should be monitored closely this year include Glasston and Oakwood, ND, and Ada, Fisher, and Stephen, MN. The remainder of the area is at lower risk. Root maggot infestations are expected to be lower in 2018 than in the past few years. However, some fields will still be at high risk of damaging infestations this year. SBRM populations can increase rapidly from year to year. Proximity to previous-year beet fields where SBRM populations were high and/or control was unsatisfactory during the previous year increases risk. Sugarbeet fields near those where high fly activity occurred in 2017 should be closely monitored in 2018. Growers in high-risk areas should use an aggressive form of at-plant insecticide treatment (i.e., granular insecticide) and a postemergence rescue insecticide (i.e., banded granules or peak-fly spray). Those in moderate-risk areas using insecticidal seed treatments for at-plant protection should monitor fly activity levels in their area, and be ready to apply additive protection if needed. All growers in known SBRM areas should pay close attention to fly activity levels in late-May through June to decide if postemergence treatment is needed. NDSU Entomology will continue to inform growers regarding SBRM activity levels and hot spots each year through radio reports, the NDSU "Crop & Pest Report", and notification of sugar cooperative agricultural staff when appropriate. Root maggot fly count information for the current season and from previous years can be viewed at: <http://www.ndsu.edu/entomology/people/faculty/boetel/flycounts/>.



SUGARBEET ROOT MAGGOT CONTROL USING SINGLE- DUAL- AND TRIPLE-COMPONENT INSECTICIDE PROGRAMS

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Introduction:

Severe infestations of the sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder), occur on a frequent basis in central and northern portions of the Red River Valley (RRV) of North Dakota and Minnesota. Published research has demonstrated that this pest is capable of causing more than 45% yield losses in the absence of effective control measures (Boetel et al. 2010). High population levels of this pest often require aggressive pest management programs to ensure adequate protection of the sugarbeet crop. Control programs in areas at high risk of damaging SBRM infestations usually consist of either a granular insecticide or an insecticidal seed treatment at planting, followed by an additive postemergence insecticide application when SBRM populations warrant it. Broadcast applications of sprayable liquid insecticides, applied on an as-needed, rescue basis, are the most commonly used postemergence tools for SBRM control in the RRV. However, the use of postemergence granular insecticide products has increased in recent years. An advantage of postemergence sprays is that growers can use a “wait and see” approach, and make informed decisions on whether rescue insecticide treatments are needed based on current fly activity levels in their fields. This research was carried out to determine the most effective combinations of planting-time and postemergence insecticides to optimize sugarbeet root maggot control.

This project involved two experiments. The objectives of Study I were to: 1) compare Counter 20G granular insecticide with Poncho Beta seed treatment for at-plant SBRM control; 2) assess the efficacy of combining Poncho Beta with Counter 20G at planting time for a one-pass SBRM control system; 3) determine the impacts of additive postemergence applications of Thimet 20G to plots initially treated with either Counter 20G or Poncho Beta seed treatment for SBRM control; 4) measure the performance of Counter 20G as a postemergence control option; and 5) determine if SBRM control can be maximized by employing a three-component (i.e., seed treatment insecticide + at-plant or postemergence granular insecticide + postemergence liquid spray) management program.

The objectives of Study II were to: 1) measure the impact of Lorsban Advanced liquid insecticide spray applications on plots initially treated at planting time with Poncho Beta seed treatment or Counter 20G for root maggot control; and 2) assess the effect of application rate on performance of Lorsban Advanced for postemergence root maggot control.

Materials and Methods:

Both experiments were established on a commercial sugarbeet field site near St. Thomas (Pembina County), ND. Betaseed 89RR52 glyphosate-resistant seed was used for all entries in both experiments, and a professional seed preparation company (Germaines Seed Technology, Fargo, ND) applied Poncho Beta insecticide to seed for all seed treatment entries. Both experiments were planted on 10 May. All plots were planted using a 6-row Monosem NG Plus 4 7x7 planter set to plant at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Plots were six rows (22-inch spacing) wide with the four centermost rows treated. The outer “guard” rows (i.e., rows one and six) on each side of the plot served as untreated buffers. Each plot was 35 feet long, and 35-foot alleys between replicates were maintained weed-free throughout the growing season through tillage operations. The experiment was arranged in a randomized complete block design with four replications of the treatments.

Planting-time insecticide applications. Counter 20G was applied by using band (B) placement (Boetel et al. 2006), which consisted of 5-inch swaths of granules delivered through Gandy™ row banders. Granular application rates were regulated by using a planter-mounted SmartBox™ computer-controlled insecticide delivery system that was calibrated on the planter immediately before all applications.

Postemergence insecticide applications (Studies I and II). Postemergence insecticides in Study I consisted of two granular materials (i.e., Counter 20G and Thimet 20G) and one liquid spray product (i.e., Lorsban Advanced). Postemergence granules (Post B) were applied on 5 June, or about 6 days before peak SBRM fly activity. Band placement of postemergence granules was achieved by using Kinze™ row banders attached to a tractor-mounted tool bar and adjusted to a height needed to deliver the insecticides in 4-inch bands. Similar to at-plant insecticide applications, postemergence granular output rates were also regulated by using a SmartBox™ system mounted on a tractor-drawn four-row toolbar. Postemergence granules were delivered in 4-inch bands by using Kinze™ row banders. All postemergence granular applications were incorporated using two pairs of rotary tines that straddled each row on the tool bar. A paired set of tines was positioned ahead of each bander, and a second pair was mounted behind the granular drop zone. This system effectively stirred soil around the bases of sugarbeet seedlings and incorporated granules as the unit passed through each plot.

The postemergence spray applications of Lorsban Advanced were broadcast-applied on 8 June (i.e., about 3 days before peak SBRM fly activity). Sprays were applied from a tractor-mounted CO₂-propelled spray system equipped with an 11-ft boom that was calibrated to deliver a finished spray volume output of 10 GPA through TeeJet™ 110015VS nozzles.

In Study II, all postemergence insecticide treatments involved Lorsban Advanced spray applications that were applied in the same manner as described for Study I. Sprays were applied on 8 June (i.e., about 3 days before peak SBRM fly activity).

Root injury ratings: Sugarbeet root maggot feeding injury was assessed in both studies on 31 July by randomly collecting ten beet roots per plot (five from each of the outer two treated rows), hand-washing them, and scoring them in accordance with the 0 to 9 root injury rating scale (0 = no scarring, and 9 = over ¾ of the root surface blackened by scarring or dead beet) of Campbell et al. (2000).

Harvest: Treatment performance was also compared on the basis of sugarbeet yield parameters. Plots for both studies were harvested on 10 October. Foliage was removed from plots immediately before harvest by using a commercial-grade mechanical defoliator. All beets from the center two rows of each plot were extracted from soil using a mechanical harvester, and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All data from root injury ratings and harvest samples were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

Study I. Sugarbeet root maggot feeding injury rating results for Study I are presented in Table 1. The level of root injury that occurred in the untreated check plots (mean = 5.48 on the 0 to 9 scale of Campbell et al. [2000]) suggested that a moderate SBRM infestation was present for this study. This is due, in large part, to a hailstorm that occurred on 9 June (2 days before peak fly activity). It is estimated that the storm killed at least 40 to 60% of the SBRM fly population in the plot area and surrounding fields. Despite that reduction in the local population, there were several significant differences among treatments in this trial. All insecticide-protected plots had significantly lower levels of SBRM feeding injury than the untreated check, regardless of whether they involved a seed treatment, single at-plant granular application, dual-, or triple-application insecticide combination was used for SBRM control.

The lowest overall root injury rating mean (i.e., highest root protection level) in Study I occurred in plots that received the triple-insecticide application treatment comprised of Poncho Beta-treated seed, combined with an at-plant application of Counter 20G at its high (8.9 lb product/ac) rate, and followed by a postemergence application of Lorsban Advanced at 1 pt/ac. Root maggot feeding injury in those plots was significantly lower than that in all other treatments, except a similar treatment that included Poncho Beta plus the at-plant application of Counter 20G at 8.9 lb, but without the post application of Lorsban Advanced. Similarly, there was no significant difference in root protection between a triple-component program consisting of Poncho Beta-treated seed plus a postemergence application of Thimet 20G, followed by a postemergence spray of Lorsban, and similar plots that received Poncho Beta and Thimet, but were not treated with the additional application of Lorsban Advanced. These results suggest that there was no significant improvement in root protection from the postemergence spray of Lorsban Advanced.

In dual insecticide programs, adding a postemergence granular product consistently provided significant improvements in root protection from SBRM feeding injury, irrespective of whether plots were initially protected at planting by Poncho Beta or any rate of Counter 20G. The lowest average SBRM feeding injury for dual-insecticide treatments was observed in plots protected by Poncho Beta-treated seed plus a planting-time application of Counter 20G at its high (8.9 lb product/ac) rate. Root maggot feeding injury in plots treated with this combination was the second-lowest in the entire trial, and it was significantly lower than that observed in plots treated with the similar dual-insecticide program that included Poncho Beta plus Counter applied at its low (5.25 lb) rate at planting.

Table 1. Larval feeding injury in an evaluation of sugarbeet root maggot control by combining planting-time insecticide granules or seed treatments with postemergence insecticides, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Poncho Beta + Counter 20G + Lorsban Advanced	Seed B 3 d Pre-peak Broadcast	8.9 lb 1 pt	68 g a.i./unit seed 1.8 0.5	0.88 g
Poncho Beta + Counter 20G	Seed B	8.9 lb	68 g a.i./unit seed 1.8	1.48 fg
Counter 20G + Thimet 20G	B 6 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	1.55 f
Poncho Beta + Counter 20G	Seed 6 d Pre-peak Post B	8.9 lb	68 g a.i./unit seed 1.8	1.68 ef
Poncho Beta + Thimet 20G + Lorsban Advanced	Seed 6 d Pre-peak Post B 3 d Pre-peak Broadcast	7 lb 1 pt	68 g a.i./unit seed 1.4 0.5	1.73 ef
Poncho Beta + Counter 20G	Seed 6 d Pre-peak Post B	5.25 lb	68 g a.i./unit seed 1.05	1.75 ef
Counter 20G + Thimet 20G	B 6 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	1.78 ef
Poncho Beta + Thimet 20G	Seed 6 d Pre-peak Post B	7 lb	68 g a.i./unit seed 1.4	1.98 def
Counter 20G	B	8.9 lb	1.8	2.23 cde
Poncho Beta + Counter 20G	Seed B	5.25 lb	68 g a.i./unit seed 1.05	2.30 cde
Counter 20G	B	7.5 lb	1.5	2.50 cd
Counter 20G	B	5.25 lb	1.05	2.85 c
Poncho Beta	Seed		68 g a.i./unit seed	4.13 b
Check	---	---	---	5.48 a
LSD (0.05)				0.667

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).
^aB = banded at planting; Post B = postemergence band; Seed = insecticidal seed treatment

Although trends suggested that higher rates of both at-plant and postemergence granular insecticides provided improved protection from SBRM feeding injury, there were no statistical differences among the three rates of Counter 20G when applied as single at-plant treatments. There also was no significant difference in root protection between at-plant and postemergence applications of when Counter 20G was applied at the low (5.25 lb/ac) rate to plots planted with Poncho Beta-treated seed.

Yield data from Study I are presented in Table 2. Most treatments in this experiment resulted in exceptionally high yields, and relative differences in treatment performance generally followed patterns observed in root maggot feeding injury data for this trial. There were very few significant differences among treatments in relation to recoverable sucrose and root tonnage yield, and there were no significant differences in percent sucrose content among treatments. The infrequent statistical differences in yields is probably due to two factors: 1) the unusually moderate SBRM feeding pressure; and 2) there was a considerable amount of variability within and between replications in this trial due to a couple of heavy rainfall during the growing season that resulted in standing water in the plots. The standing water would have added variability to root yields, but also could have precluded SBRM females from laying eggs in the affected plots.

As observed in the SBRM feeding injury data for this trial, trends suggested better performance with increasing rates of both at-plant and postemergence applications of Counter 20G, although significant rate-related differences were rare. The top-performing entries with regard to both recoverable sucrose and root tonnage included the following: 1) Poncho Beta + Counter 20G applied at planting time at 8.9 lb/ac; 2) the triple-component program consisting of Poncho Beta seed treatment, combined with an at-plant application of Counter 20G at its high (8.9 lb product/ac) rate and a postemergence spray application of Lorsban Advanced at its moderate rate of 1 pt/ac; 3)

Poncho Beta + postemergence Thimet 20G at 7 lb/ac + Lorsban Advanced applied postemergence at 1 pt/ac; and 4) at-plant Counter 20G + postemergence Thimet 20G, both applied at their respective high labeled rates of 8.9 and 7 lb/ac. Yields from these treatments were not statistically different from the single planting-time application of Counter at 8.9 lb/ac or most of the dual-insecticide programs in this trial. However, these top-performing treatments generated between \$97 and \$159/ac more gross revenue than the at-plant application of Counter at 8.9 lb/ac, and between \$201 and \$263/ac more revenue than the untreated check plots. These economic benefits would have easily paid for the costs of their use, and provided significant amounts of additional revenue per acre.

The gross economic return generated by using stand-alone planting-time applications of Counter 20G ranged between \$104 and \$172/ac, which would have significantly exceeded the treatment cost and provided additional net revenue. The use of Poncho Beta as a stand-alone form of protection generated an increase of \$57/ac in gross return, which also would have easily paid for the cost of the treatment.

Although these results demonstrate the economic benefits of at-plant protection against SBRM feeding injury and associated yield/revenue loss, they also clearly demonstrate the economic value of applying an additive insecticide, either in the form of a planting-time insecticide (if insecticide-treated seed is used), or a postemergence insecticide application (whether the initial at-plant protection consists of a seed treatment or a conventional insecticide).

Table 2. Yield parameters from an evaluation of sugarbeet root maggot control by combining planting-time insecticide granules or seed treatments with postemergence insecticides, St. Thomas, ND, 2017

Treatment/ form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Poncho Beta + Counter 20G	Seed B	8.9 lb	68 g a.i./unit seed 1.8	12,433 a	40.2 ab	16.70 a	1,477
Poncho Beta + Counter 20G + Lorsban Advanced	Seed B 3 d Pre-peak Broadcast	8.9 lb 1 pt	68 g a.i./unit seed 1.8 0.5	12,400 a	41.0 a	16.48 a	1,427
Poncho Beta + Thimet 20G + Lorsban Advanced	Seed 6 d Pre-peak Post B 3 d Pre-peak Broadcast	7 lb 1 pt	68 g a.i./unit seed 1.4 0.5	12,388 a	41.0 a	16.35 a	1,423
Counter 20G + Thimet 20G	B 6 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	12,173 ab	40.0 abc	16.40 a	1,415
Poncho Beta + Counter 20G	Seed 6 d Pre-peak Post B	8.9 lb	68 g a.i./unit seed 1.8	12,083 abc	39.3 a-d	16.60 a	1,425
Poncho Beta + Counter 20G	Seed B	5.25 lb	68 g a.i./unit seed 1.05	12,045 abc	40.1 ab	16.35 a	1,375
Counter 20G + Thimet 20G	B 6 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	11,905 abc	39.8 abc	16.20 a	1,348
Counter 20G	B	7.5 lb	1.5	11,720 abc	38.0 cde	16.70 a	1,386
Poncho Beta + Thimet 20G	Seed 6 d Pre-peak Post B	7 lb	68 g a.i./unit seed 1.4	11,710 abc	40.2 ab	16.00 a	1,275
Counter 20G	B	8.9 lb	1.8	11,637 abc	38.9 b-e	16.20 a	1,318
Counter 20G	B	5.25 lb	1.05	11,468 bcd	37.5 de	16.55 a	1,343
Poncho Beta	Seed		68 g a.i./unit seed	11,372 bcd	37.0 e	16.68 a	1,341
Poncho Beta + Counter 20G	Seed 6 d Pre-peak Post B	5.25 lb	68 g a.i./unit seed 1.05	11,192 cd	38.8 b-e	15.78 a	1,199
Check	---	----	---	10,560 d	35.0 f	16.38 a	1,214
LSD (0.05)				917.6	2.01	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Post B = postemergence band; Seed = insecticidal seed treatment

The following treatments failed to provide statistically significant increases in recoverable sucrose yield when compared to the untreated check plots: 1) Counter 20G applied at planting at its low (5.25 lb/ac) rate; 2) Poncho Beta seed treatment; and 3) Poncho Beta + Counter 20G applied postemergence at 5.25 lb/ac.

It should be noted that Counter insecticide can only be applied once per year. Therefore, if Counter is applied at planting, it cannot be applied to the same field at postemergence. It also bears noting that the Counter 20G label has been revised to include a 90-day preharvest interval (i.e., PHI, the number of days that must elapse after application before a crop can be harvested) for sugarbeet. This makes Counter 20G a much more feasible product as a postemergence option for sugarbeet root maggot control than before, as it previously was labeled with a 110-day PHI.

The 90-day PHI should work well for Red River Valley growers choosing to use Counter 20G for SBRM management. Postemergence granule applications for SBRM control in the growing area are typically most effective if made in late-May to early-June. If this product were to be applied to a field on June 1, the 90-day PHI would expire before September 1, which is typically the earliest that preliminary sugarbeet harvest operations begin in the Valley.

Study II. Results from evaluations of sugarbeet root maggot larval feeding injury in Study II are shown in Table 3. Moderate larval feeding pressure developed in this trial, as was evidenced by the moderate root injury rating mean recorded for the untreated check plots (5.98 on the 0 to 9 scale). All insecticide-treated entries provided significant reductions in SBRM feeding injury when compared to the injury recorded in the untreated check plots.

Table 3. Larval feeding injury in an evaluation of sugarbeet root maggot control by combining planting-time insecticide granules or seed treatments with postemergence liquid sprays, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 1 pt	1.8 0.5	1.25 e
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 2 pts	1.5 1.0	1.73 de
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 2 pts	1.8 1.0	1.90 cde
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	2.23 cd
Counter 20G	B	8.9 lb	1.8	2.63 bc
Counter 20G	B	7.5 lb	1.5	2.65 bc
Poncho Beta + Lorsban Advanced	Seed 3 d Pre-peak Broadcast	1 pt	68 g a.i./unit seed 0.5	3.10 b
Poncho Beta + Lorsban Advanced	Seed 3 d Pre-peak Broadcast	2 pts	68 g a.i./unit seed 1.0	3.25 b
Poncho Beta	Seed		68 g a.i./unit seed	3.38 b
Check	---	---	---	5.98 a
LSD (0.05)				0.837

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Seed = insecticidal seed treatment

The following treatments provided the highest levels of root protection in Study II: 1) Counter banded at 8.9 lb product/ac + Lorsban Advanced postemergence at 1 pt/ac; 2) Counter banded at 7.5 lb product/ac + Lorsban Advanced at 2 pts/ac; and 3) Counter banded at 8.9 lb product/ac + Lorsban Advanced postemergence at 2 pts/ac. There were no significant differences in levels of SBRM feeding injury among these three treatments.

In plots initially treated with at-plant applications of Counter 20G at its high (8.9 lb) rate, the addition of a postemergence application of Lorsban Advanced at 1 pt/ac resulted in a significant improvement in root protection from SBRM feeding injury when compared to plots that only received the at-plant application of Counter. Similarly, applying Lorsban Advanced at 2 pts/ac to plots initially treated with Counter at its moderate (7.5 lb) rate resulted in a significant reduction in SBRM feeding injury when compared to plots that only received the moderate rate of Counter. In contrast, there were no significant improvements in protection from SBRM feeding injury by adding postemergence applications of Lorsban Advanced to plots initially protected with Poncho Beta-treated seed.

Yield results for Study II (Table 4) were somewhat supportive of the root maggot feeding injury rating results. As observed in Study I of this project, recoverable sucrose and root tonnage yields were exceptionally high for most treatments. This was partly due to the comparatively low root maggot infestation, but also a result of good growing conditions in the St. Thomas area during 2017.

The top-performing treatments with regard to recoverable sucrose yield in Study II included the following: 1) Counter banded at 8.9 lb product/ac + Lorsban Advanced postemergence at 1 pt/ac; 2) Counter banded at 7.5 lb product/ac + Lorsban Advanced at 1 pt/ac; 3) Counter banded at 8.9 lb product/ac + Lorsban Advanced postemergence at 2 pts/ac; and 4) Counter banded at 7.5 lb product/ac + Lorsban Advanced at 2 pts/ac. There were no significant differences among these treatments with respect to recoverable sucrose yield. The best treatment overall, regarding recoverable sucrose and root yield, and gross economic return, was Counter banded at 8.9 lb product/ac + Lorsban Advanced postemergence at 1 pt/ac. Plots protected by this treatment produced significantly more recoverable sucrose per acre than most other treatments, and significantly more root yield than all treatments, except the combination of Counter banded at 7.5 lb product/ac + Lorsban Advanced postemergence at 2 pts/ac.

Table 4. Yield parameters from an evaluation of sugarbeet root maggot control by combining planting-time insecticide granules or seed treatments with postemergence liquid sprays, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 1 pt	1.8 0.5	12,357 a	38.9 a	16.98 a	1,519
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	11,595 ab	35.7 bcd	17.33 a	1,469
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 2 pt	1.8 1.0	11,487 abc	35.8 bcd	17.20 a	1,432
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 2 pts	1.5 1.0	11,425 abc	36.9 ab	16.70 a	1,358
Counter 20G	B	7.5 lb	1.5	11,300 bc	36.5 bc	16.75 a	1,346
Counter 20G	B	8.9 lb	1.8	11,233 bc	35.5 bcd	16.90 a	1,374
Poncho Beta + Lorsban Advanced	Seed 3 d Pre-peak Broadcast	2 pts	68 g a.i./unit seed 1.0	10,642 bcd	34.6 cde	16.58 a	1,256
Poncho Beta + Lorsban Advanced	Seed 3 d Pre-peak Broadcast	1 pt	68 g a.i./unit seed 0.5	10,572 cd	33.9 de	16.80 a	1,273
Poncho Beta	Seed		68 g a.i./unit seed	10,133 d	32.4 ef	16.83 a	1,224
Check	---	----	---	9,768 d	31.1 f	16.85 a	1,187
LSD (0.05)				972.8	2.24	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Seed = insecticidal seed treatment

Estimated gross revenue from treatment combinations that included Counter 20G at planting, followed by a postemergence application of Lorsban Advanced, ranged from \$1,358 to \$1,519/ac, which translated to revenue increases of between \$171 and \$332/ac when compared to revenue from the untreated check plots. Plots protected by single, planting-time applications of Counter 20G generated revenue increases of between \$159 and \$187. Plots planted with Poncho Beta-treated seed (i.e., without an additive postemergence insecticide application) generated a revenue increase of \$37/ac; however, applying a postemergence application of Lorsban Advanced to Poncho Beta plots resulted in additional revenue increases ranging from \$69 to \$86/ac.

In general, the results from Study II indicate that effective root maggot control, even under moderate infestation levels such as those that developed in this trial, can result in significant yield increases. These findings also demonstrate that single-component insecticide programs may not provide sufficient protection from yield losses associated with SBRM larval feeding injury, even under such moderate infestations. Although the returns generated by single control tool entries in this study would easily justify their use, these results demonstrate that more aggressive approaches, combining at-plant and postemergence rescue insecticide protection, can contribute substantially to maximizing economic returns from sugarbeet production in areas affected by this pest.

References Cited:

- Boetel, M.A., R.J. Dregseth, and A.J. Schroeder. 2010.** Economic benefits of insecticide applications for root maggot control in replanted sugarbeet. *J. Sugar Beet Res.* 47: 35-49.
- Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006.** Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47-63.

Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000. Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. *J. Sugar Beet Res.* 37: 57–69.

SAS Institute. 2008. *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

APPLICATION TIMING AND RATE EFFECTS ON POSTEMERGENCE INSECTICIDE SPRAYS FOR ROOT MAGGOT CONTROL

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Introduction:

Severe infestations of the sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder), occur commonly in central and northern portions of the Red River Valley (RRV) growing area of North Dakota and Minnesota. As such the SBRM is an ongoing threat to farm profitability for producers in the area. This intense insect pressure typically requires aggressive pest management programs to ensure adequate protection of the sugarbeet crop. Pest management programs in areas at high risk for damaging SBRM infestations usually consist of either a granular insecticide or an insecticidal seed treatment at planting, followed by an additive postemergence insecticide application when the localized infestation level warrants it. The most commonly used approach for postemergence root maggot control in the RRV is a broadcast application of a sprayable liquid insecticide product.

Beginning with the 2010 growing season, federal label changes resulted in a 10-day reapplication interval for all sprayable liquid insecticide products containing the active ingredient chlorpyrifos (e.g., Lorsban 4E, Lorsban Advanced, and all generic versions). The label revision lengthened the reapplication interval by three days. This change may have compromised the ability of sugarbeet growers to effectively manage the SBRM with chlorpyrifos-based products, because fly activity peaks usually rise and fall relatively quickly, often subsiding in about seven days. In an effort to address this potential problem, research was undertaken to achieve the following objectives regarding postemergence SBRM management: 1) determine the most effective timing schemes for repeated applications of Lorsban Advanced sprays that adhere to its 10-day reapplication restriction; 2) assess the impact of application rate on Lorsban Advanced performance; and 3) evaluate Mustang Maxx as a single postemergence tool and as rotated with Lorsban Advanced applications for postemergence SBRM control.

Materials and Methods:

This experiment was conducted on a commercial sugarbeet field site near St. Thomas in rural Pembina County, ND. Betaseed 89RR52 glyphosate-resistant seed was used for all treatments. Plots were planted on 11 May. All plots were planted using a 6-row Monosem NG Plus 4 7x7 planter set to deliver seed at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Plots were six rows (22-inch spacing) wide with the four centermost rows treated. The outer “guard” row on each side of the plot served as an untreated buffer. Each plot was 35 feet long, and 35-foot tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design with four replications of the treatments.

Planting-time insecticide applications. Planting-time applications of Counter 20G were applied by using band (B) placement (Boetel et al. 2006), which consisted of 5-inch swaths of granules delivered through Gandy™ row banders. Granular application rates were regulated by using planter-mounted SmartBox™ computer-controlled insecticide delivery system that had been calibrated on the planter before all applications.

Postemergence insecticide applications. Additive postemergence insecticides used included Lorsban Advanced and Mustang Maxx. Treatments that included postemergence applications involved both single and double postemergence spray applications at varying rates. Treatment timings compared included six, five, and three days pre-peak SBRM fly activity (i.e., 5, 6, and 8 June, respectively, and five, four, and eight days after peak fly (i.e., 15, 16, and 19 June, resp.). Liquid insecticide solutions were delivered with a tractor-mounted CO₂-propelled spray system equipped with TeeJet™ 110015VS nozzles calibrated to deliver applications in a finished output volume of 10 GPA.

Root injury ratings: Sugarbeet root maggot feeding injury was assessed in this experiment on 1 August, by randomly collecting ten beet roots per plot (five from each of the outer two treated rows), hand-washing them, and scoring them in accordance with the 0 to 9 root injury rating scale (0 = no scarring, and 9 = over ¾ of the root surface blackened by scarring or dead beet) of Campbell et al. (2000).

Harvest: Treatment performance was also compared on the basis of sugarbeet yield parameters. Plots were harvested on 2 October. Foliage was removed from plots immediately before harvest by using a commercial-grade mechanical defoliator. All beets from the center two rows of each plot were extracted from soil using a mechanical harvester, and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All data from root injury ratings and harvest samples were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

NOTE: Results of this trial should be interpreted with some degree of discretion, because a strong hailstorm occurred in the plot area just two days before the anticipated peak in SBRM fly activity. As a result, we estimate that at least 40-60% of the fly population in the immediate vicinity of this trial site were likely killed.

Sugarbeet root maggot feeding injury ratings in the untreated check plots averaged 4.8 on the 0 to 9 scale of Campbell et al. (2000) (Table 1). This suggested that a moderate SBRM infestation was present for the experiment.

Table 1. Larval feeding injury in an assessment of postemergence insecticide spray timing, rate, and frequency impacts on sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 5 d Pre-peak Broadcast 5 d Post-peak Broadcast	7.5 lb 2 pts 2 pts	1.5 1.0 1.0	1.28 c
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 2 pts	1.8 1.0	1.33 c
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 5 d Pre-peak Broadcast 5 d Post-peak Broadcast	7.5 lb 1 pts 1 pts	1.5 1.0 1.0	1.35 c
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 3 d Pre-peak Broadcast 8 d Post-peak Broadcast	7.5 lb 1 pt 1 pt	1.5 0.5 0.5	1.38 c
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 6 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 2 pts 2 pts	1.5 1.0 1.0	1.40 bc
Counter 20G + Lorsban Advanced + Mustang Maxx	B 3 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 1 pt 4 fl oz	1.5 0.5 0.025	1.60 bc
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 6 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 1 pt 1 pt	1.5 0.5 0.5	1.70 bc
Counter 20G + Mustang Maxx + Lorsban Advanced	B 3 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 4 fl oz 1 pt	1.5 0.025 0.5	1.80 bc
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 2 pts	1.5 1.0	1.98 bc
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	2.03 bc
Counter 20G	B	8.9 lb	1.8	2.10 bc
Counter 20G + Mustang Maxx	B 3 d Pre-peak Broadcast	7.5 lb 4 fl oz	1.5 0.025	2.25 bc
Counter 20G	B	7.5 lb	1.5	2.48 b
Check	---	---	---	4.80 a
LSD (0.05)				1.091

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting

The moderate feeding pressure resulted in very few significant differences among treatments in this experiment. All insecticide treatments, whether involving single at-plant applications, or at-plant/postemergence combinations, provided significant reductions in feeding injury when compared to the untreated check. There were no statistically significant differences in root protection among any of the treatments in this trial that included both a planting-time application of Counter 20G plus at least one postemergence spray of either Lorsban Advanced or

Mustang Maxx. No rate-related differences in performance were observed either, although general patterns indicated that the best protection from root maggot feeding injury was provided by entries that involved combining planting-time with aggressive postemergence control programs comprised of two spray applications.

The following treatments provided the best average protection from SBRM feeding injury in this trial:

- 1) planting-time Counter 20G at 8.9 lb/ac + two 2-pt/ac applications of Lorsban Advanced at 5 days pre-and 5 days post-peak;
- 2) planting-time Counter at 8.9 lb/ac + one 2-pt/ac postemergence application of Lorsban Advanced at 3 days pre-peak;
- 3) planting-time Counter at 7.5 lb/ac + two 1-pt/ac postemergence applications of Lorsban Advanced at 5 days pre-and 5 days post-peak; and
- 4) planting-time Counter at 7.5 lb/ac + two 1-pt/ac postemergence applications of Lorsban Advanced at 3 days pre-peak and 8 days post-peak.

Treatment timing, in relation to the required 10-day reapplication interval, did not have a significant impact on performance of Lorsban Advanced applications in relation to preventing SBRM feeding injury. Another trend observed suggested that the high (2 pt/ac) rate of Lorsban Advanced tended to provide slightly better root protection than the 1 pt/ac rate. In treatment combinations that included postemergence applications of both Lorsban Advanced and Mustang Maxx, control appeared to be slightly improved by applying the Lorsban Advanced during the pre-peak application, and following with a post-peak application of Mustang Maxx. Another pattern observed was that splitting 2 pts of Lorsban Advanced into two 1-pt applications spaced 10 days apart appeared to provide a slight improvement in root protection, although the difference was not statistically significant.

Yield results and associated gross economic returns from this trial are presented in Table 2. All insecticide treatments provided significant increases in both recoverable sucrose yield and root tonnage. As observed with root injury rating data, there were also very few significant differences among insecticide treatments with respect to recoverable sucrose yield. This was probably a product of the atypically moderate SBRM larval feeding pressure that occurred following the pre-peak-fly hailstorm. Variability within and between replicates in the plot area due to standing water in some plots during the SBRM egg-laying period could have also contributed to the relatively low incidence of significant differences in this experiment.

The best overall treatments in this trial with regard to recoverable sucrose yield included the following:

- 1) planting-time Counter 20G at 7.5 lb/ac + two 2-pt/ac applications of Lorsban Advanced at 6 days pre-and 4 days post-peak;
- 2) planting-time Counter at 7.5 lb/ac + two 1-pt/ac postemergence applications of Lorsban Advanced at 6 days pre-and 4 days post-peak;
- 3) planting-time Counter at 7.5 lb/ac + two 2-pt/ac postemergence applications of Lorsban Advanced at 5 days pre-and 5 days post-peak; and
- 4) planting-time Counter at 7.5 lb/ac + postemergence Lorsban Advanced (1 pt/ac) at 3 days pre-peak + Mustang Maxx (4 fl oz/ac) at 4 days post-peak.

There were no significant different differences among these four treatments with regard to either recoverable sucrose yield or root tonnage. Although significant yield differences were rare in this study, it should be noted that these top four treatments all included a planting-time application of Counter 20G at its moderate rate (7.5 lb/ac). These results may suggest that, under moderate SBRM feeding pressure, a moderate rate of at-plant protection, followed by more aggressive postemergence control strategy (i.e., two split applications of a postemergence liquid insecticide spray), may allow growers to optimize sucrose yield and revenue.

In comparisons among dual- and triple-insecticide component programs, there were two key findings. First, the top-yielding treatment consisted of Counter 20G applied at planting at 7.5 lb product per acre combined with two postemergence applications of Lorsban Advanced at 2 pts/ac. In that program, adding a second application of Lorsban Advanced generated significantly greater recoverable sucrose yield (1,401 lb increase) and root tonnage (3.5-ton increase), and \$223 more in gross revenue than a similar program that only included a single 2-pt/ac application of Lorsban Advanced. Second, the program that included the same moderate rate of Counter (7.5 lb/ac) at planting, followed by two split applications of postemergence Lorsban Advanced at 1 pt/ac (6 days pre- and 4 days post-peak), also produced significantly more sucrose and root yield than when the Lorsban Advanced was applied in a single 2-pt/ac application. The program involving two split postemergence applications of Lorsban Advanced at 1 pt/ac generated \$249 more gross economic return than when the Lorsban was applied as a single 2-pt application.

Table 2. Yield parameters from an assessment of postemergence insecticide spray timing, rate, and frequency impacts on sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 6 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 2 pts 2 pts	1.5 1.0 1.0	12,187 a	39.7 ab	16.60 a	1,436
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 6 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 1 pt 1 pt	1.5 0.5 0.5	12,015 a	38.9 ab	16.70 a	1,427
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 5 d Pre-peak Broadcast 5 d Post-peak Broadcast	7.5 lb 2 pts 2 pts	1.5 1.0 1.0	12,007 a	40.0 a	16.43 a	1,367
Counter 20G + Lorsban Advanced + Mustang Maxx	B 3 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 1 pt 4 fl oz	1.5 0.5 0.025	11,827 ab	40.2 a	16.10 a	1,310
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 3 d Pre-peak Broadcast 8 d Post-peak Broadcast	7.5 lb 1 pt 1 pt	1.5 0.5 0.5	11,697 ab	38.4 abc	16.50 a	1,363
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	11,544 ab	37.5 bc	16.60 a	1,363
Counter 20G	B	8.9 lb	1.8	11,499 ab	38.9 ab	16.18 a	1,283
Counter 20G + Mustang Maxx	B 3 d Pre-peak Broadcast	7.5 lb 4 fl oz	1.5 0.025	11,441 ab	37.5 bc	16.50 a	1,333
Counter 20G + Lorsban Advanced + Lorsban Advanced	B 5 d Pre-peak Broadcast 5 d Post-peak Broadcast	7.5 lb 1 pts 1 pts	1.5 1.0 1.0	11,347 ab	39.1 ab	15.93 a	1,229
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	8.9 lb 2 pts	1.8 1.0	11,295 ab	39.8 a	15.70 a	1,178
Counter 20G + Mustang Maxx + Lorsban Advanced	B 3 d Pre-peak Broadcast 4 d Post-peak Broadcast	7.5 lb 4 fl oz 1 pt	1.5 0.025 0.5	11,264 ab	37.5 bc	16.30 a	1,285
Counter 20G	B	7.5 lb	1.5	11,258 ab	37.9 abc	16.20 a	1,264
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 2 pts	1.5 1.0	10,781 b	36.2 c	16.20 a	1,213
Check	---	---	---	9,182 c	30.9 d	16.10 a	1,030
LSD (0.05)				1,198.6	2.29	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting

Meaningful trends observed in this trial involved treatment timing and order. For example, numerically (i.e., not statistically significant) greater recoverable sucrose yield was produced when the first applications of Lorsban Advanced in dual postemergence sprays were applied earlier (6 days pre- + 4 days post-peak vs. 5 days pre- + 5 days post-peak and 3 days pre- + 8 days post-peak). Also, in postemergence spray programs where Lorsban Advanced and Mustang Maxx were alternated, applying the Lorsban on the pre-peak spray and following it with Mustang Maxx resulted in numerically greater recoverable sucrose yield and significantly more root tonnage than when the Mustang was applied first.

Despite the moderate SBRM feeding pressure that was present during this experiment, most of the SBRM control programs evaluated in this experiment provided effective SBRM control that translated to major yield benefits. Another general conclusion that can be drawn is that the root protection, yield, and revenue benefits from additive postemergence insecticides demonstrate that they are cost-effective tools to use in areas where damaging SBRM populations occur.

References Cited:

- Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006.** Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.
- Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000.** Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. *J. Sugar Beet Res.* 37: 57–69.
- SAS Institute. 2008.** *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

DOES APPLICATION RATE OR TIMING IMPACT PERFORMANCE OF THIMET 20G FOR POSTEMERGENCE SUGARBEET ROOT MAGGOT CONTROL?

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Introduction:

In recent years, sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder), populations have been at alarmingly high levels in central and northern portions of the Red River Valley. This has provided the impetus to refine postemergence tools for more effective SBRM management. The key objective of this experiment was to assess the impacts of application timing and rate on the performance of Thimet 20G insecticide when applied as a postemergence rescue insecticide for SBRM control in the Red River Valley. A secondary objective was to compare moderate and high rates of Counter 20G (i.e., 7.5 and 8.9 lb product/acre, respectively) as planting-time components in dual-insecticide (i.e., planting-time + postemergence) programs for root maggot control.

Materials and Methods:

This study was planted on 10 May at a commercial field site near St. Thomas (Pembina County), ND. Plots were planted using a 6-row Monosem NG Plus 7x7 planter set to plant at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Plots were six rows (22-inch spacing) wide with the four centermost rows treated. The outer two rows of each plot served as untreated buffers. Individual plots were 35 feet long, and 35-foot tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design with four replications of the treatments. Counter 20G was applied as a base planting-time insecticide for all plots that received insecticide protection, and it was applied at either the moderate (7.5 lb product/ac) or high (8.9 lb/ac) labeled rate. Band (B) placement (Boetel et al. 2006), which consisted of 5-inch swaths of granules delivered through Gandy™ row banders, was used for all applications of Counter 20G. Granular output rates were regulated by using a planter-mounted SmartBox™ computer-controlled insecticide system that was calibrated on the planter before planting.

Postemergence Thimet 20G granules were applied at either 11 or five days before peak fly activity (i.e., 31 May or 6 June, respectively), and rates of Thimet 20G included 4.9 and 7 lb product/ac. As with at-plant applications, granular output rates were regulated by using a SmartBox™ system mounted on a tractor-drawn four-row toolbar, and placement of insecticide in 4-inch bands was achieved by using Kinze™ row banders. Granules were incorporated by using two pairs of metal rotary tines that straddled each row. A set of tines was positioned ahead of each bander, and a second pair was mounted behind the granular drop zone. Lorsban Advanced, applied in a broadcast at 1 pt product/ac using TeeJet™ 110015VS nozzles, was also included in this experiment for comparative purposes. This application was made on 8 June, which was two days before the initial peak in SBRM fly activity.

Root injury ratings: Root maggot feeding injury assessments were carried out on 31 July by randomly collecting ten beet roots per plot (five from each of the outer two treated rows), hand-washing them, and scoring them in accordance with the 0 to 9 root injury rating scale (0 = no scarring, and 9 = over ¾ of the root surface blackened by scarring or dead beet) of Campbell et al. (2000).

Harvest: Performance was also compared using sugarbeet yield parameters derived by harvesting roots from all treatment plots. All foliage was removed from plots immediately before harvest on 3 October by using a commercial-grade mechanical defoliator. On the same day, all beets from the center two rows of each plot were extracted from soil by using a mechanical harvester, and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All data from root injury ratings and yield/quality analyses were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008). Treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

Root maggot feeding injury results from this trial are presented in Table 1. The SBRM infestation present for this experiment was classified as moderate, as was evidenced by the moderate average feeding injury rating of 5.2 (0 to 9 scale of Campbell et al. 2000) in the untreated check plots. Although all insecticide entries in the experiment provided significant reductions in SBRM feeding injury when compared to the untreated check, the moderate infestation resulted in very few statistically significant differences among insecticide treatments. Most of the dual (i.e., planting-time plus postemergence) insecticide programs that included a planting-time application of Counter 20G at its moderate rate of 7.5 lb product/ac rate, followed by a postemergence application of Thimet 20G provided significant improvements in root protection from SBRM feeding injury when compared to those that only received the single, 7.5-lb application of Counter at planting time. Exceptions to this were the 11-day pre-peak fly applications of Thimet that followed the moderate rate of Counter. When the full 8.9-lb rate of Counter was applied at planting, there were numerical reductions in SBRM feeding injury in plots that received a postemergence application of Thimet, but none of the differences were statistically significant. As observed in previous years of testing, there were no significant differences in root protection from SBRM feeding injury in relation to timing of the Thimet applications, regardless of the rate of the initial at-plant rate of Counter. There also was no significant application rate response in feeding injury ratings between the single, at-plant applications of 7.5 and 8.9 lb of Counter 20G, thus suggesting that the higher rate is probably not necessary in a dual-insecticide program under low to moderate SBRM pressure such as that which occurred during this trial.

Table 1. Larval feeding injury in an evaluation of Thimet 20G application timing and rate on sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	1.13 c
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	7.5 lb 4.9 lb	1.5 1.0	1.20 c
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	1.20 c
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	1.20 c
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	1.48 bc
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	7.5 lb 4.9 lb	1.5 1.0	1.60 bc
Counter 20G	B	8.9 lb	1.8	1.90 bc
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	2.00 b
Counter 20G	B	7.5 lb	1.5	2.23 b
Check	-----	----	-----	5.20 a
LSD (0.05)				0.785

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Post B = postemergence band

The postemergence spray of Lorsban Advanced, applied at its moderate labeled rate (1 pt product/ac) did not provide a significant improvement in root protection when added to plots initially treated with the 7.5-lb rate of Counter 20G at planting. This result may have been caused by the hailstorm and associated heavy rainfall that occurred on June 9, which was just one day after the Lorsban Advanced was applied.

Yield data from this experiment are presented in Table 2. All insecticide-treated entries in this trial, except the single planting-time application of Counter 20G at its moderate rate (7.5 lb product/ac) of resulted in significant increases in recoverable sucrose yields when compared to the untreated check. Plots treated with the combination of Counter 20G at its high (8.9 lb product/ac) rate plus a postemergence application of the high (7 lb/ac) rate of Thimet 20G at 11 days before peak fly generated the highest average recoverable sucrose and root yield in the trial. Roots harvested from that treatment also had the highest percentage sucrose content in the study; however, very few of the differences were statistically significant. There were no significant differences in recoverable sucrose, root tonnage,

or percent sucrose between the single planting-time applications of Counter 20G. Similarly, there were no significant differences for any yield parameter between Thimet application rates or timings tested.

Table 2. Impacts of Thimet 20G application timing and rate on yield parameters in an evaluation of sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	11,179 a	33.3 a	17.73 a	1,472
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	7.5 lb 4.9 lb	1.5 1.0	10,689 ab	32.6 a	17.25 a	1,367
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	8.9 lb 7 lb	1.8 1.4	10,636 ab	33.2 a	16.93 a	1,322
Counter 20G + Lorsban Advanced	B 3 d Pre-peak Broadcast	7.5 lb 1 pt	1.5 0.5	10,595 ab	32.7 a	17.10 a	1,337
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	10,582 ab	32.4 a	17.30 a	1,349
Counter 20G + Thimet 20G	B 5 d Pre-peak Post B	7.5 lb 4.9 lb	1.5 1.0	10,514 ab	33.0 a	16.95 a	1,300
Counter 20G + Thimet 20G	B 11 d Pre-peak Post B	7.5 lb 7 lb	1.5 1.4	10,349 ab	32.7 a	16.73 a	1,266
Counter 20G	B	8.9 lb	1.8	10,332 ab	33.0 a	16.68 a	1,249
Counter 20G	B	7.5 lb	1.5	9,737 bc	32.9 a	16.05 a	1,086
Check	-----	----	-----	8,595 c	27.6 b	16.55 a	1,029
LSD (0.05)				1,204.6	2.41	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Post B = postemergence band

Yield trends in this experiment suggested an advantage to using the higher rate of Counter 20G (8.9 lb product/ac) at planting time plus postemergence Thimet 20G earlier (11 days ahead of peak fly activity). This treatment generated \$150/ac more gross revenue per acre than when the Thimet was applied at 5 days pre-peak. Similarly, when lower rates of both Counter 20G (7.5 lb/ac) and Thimet 20G (4.9 lb/ac) were used for SBRM management, applying the postemergence Thimet 11 days pre-peak resulted in an increase in gross revenue by \$67/ac when compared to revenue from the same program if the Thimet was applied at 5 days ahead of peak fly.

Adding postemergence applications of Thimet 20G to plots initially treated with a planting-time application of Counter 20G at its high (8.9 lb/ac) labeled rate generated gross economic return increases that ranged from \$73 to \$223 per acre above the revenue from planting-time-only applications of Counter at 8.9 lb per acre. Similarly, plots initially treated at planting with Counter at the moderate (7.5 lb product/ac) rate produced revenue increases of between \$180 and \$281/ac when a postemergence application of Thimet was added. Plots that received 7.5 lb of Counter at planting and a postemergence rescue application of Lorsban Advanced three days ahead of peak fly generated an increase in gross economic return of \$251/ac.

As observed in previous years of testing, the results of this experiment showed that combining at-plant Counter 20G with postemergence applications of Thimet 20G provides effective control of the sugarbeet root maggot, and that Thimet performance is not significantly impacted by application timing (i.e., seven days pre-peak vs. peak fly) or rate. This allows growers a wide window of flexibility in relation to when the Thimet must be applied to achieve satisfactory SBRM control. The additional economic returns from postemergence insecticide applications in this experiment provide ample justification for the use of these materials to provide additive control of the sugarbeet root maggot. The fact that insecticide protection, in the form of either a single at-plant insecticide or a dual-insecticide program, increased gross economic returns by between \$57 and \$443/ac above the untreated check provides strong evidence regarding the economic importance of the sugarbeet root maggot as a serious pest of sugarbeet. Effective SBRM management programs, such as the dual-insecticide programs tested in this experiment, will be essential to ensuring the profitability of sugarbeet production in areas affected by moderate to high infestations of this pest.

References Cited:

- Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006.** Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.
- Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000.** Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. *J. Sugar Beet Res.* 37: 57–69.
- SAS Institute. 2008.** *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

EVALUATION OF EXPERIMENTAL INSECTICIDES FOR MANAGEMENT OF THE SUGARBEET ROOT MAGGOT

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Introduction:

The sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder) is a major pest of sugarbeet in the Red River Valley (RRV). Observations during the past 15+ years suggest that economically significant SBRM infestations frequently develop on between 50,000 and 85,000 acres within the RRV production area each year. Sugarbeet producers in the U.S. have a limited number of insecticides that are currently registered by the U.S. Environmental Protection Agency (EPA) for root maggot management. With so few options available for SBRM control, RRV sugarbeet producers have had to rely heavily on the same insecticide mode of action (i.e., acetylcholinesterase [ACHE] inhibition) to manage this pest for over 40 years.

In areas affected by severe SBRM infestations, many fields frequently require two to three applications of these materials each growing season to achieve satisfactory control. This long-term pattern of repeated use of ACHE-inhibiting insecticides has exerted intense selection pressure for the development of insecticide resistance in root maggot populations in the RRV. Therefore, research is critically needed to develop alternative strategies for root maggot management to ensure the long-term sustainability and profitability of sugarbeet production for growers affected by this pest. This experiment was carried out to achieve the following objectives: 1) test several natural and/or botanical insecticides for efficacy at managing the sugarbeet root maggot; and 2) evaluate commercially available, EPA-labeled conventional chemical insecticides that are currently not registered for use in sugarbeet to determine if their performance would warrant future pursuit of labeling for use in the crop for SBRM control

Materials and Methods:

This experiment was carried out on a commercial sugarbeet field site near St. Thomas (Pembina County), ND. The experiment was planted on 11 May using Betaseed 89RR52 glyphosate-resistant seed. All plots were planted using a 6-row Monosem NG Plus 4 7x7 planter set to plant at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Plots were six rows (22-inch spacing) wide with the four centermost rows treated. The outer “guard” rows (i.e., rows one and six) on each side of the plot served as untreated buffers. Each plot was 35 feet long, and 35-foot tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design with four replications of the treatments. All insecticide treatments were single, stand-alone (i.e., planting-time or postemergence) applications. For example, there was no at-plant insecticide in plots assigned to receive a postemergence insecticide, and vice versa.

Planting-time insecticide applications. Counter 20G was used for comparative purposes as a planting-time standard chemical insecticide in this experiment. It was applied by using band (B) placement (Boetel et al. 2006), which consisted of 5-inch swaths of granules delivered through Gandy™ row banders. Granular application rates were regulated by using a planter-mounted SmartBox™ computer-controlled insecticide delivery system calibrated on the planter immediately before all applications. Planting-time liquid insecticides included the following: 1) Aza-Direct (active ingredient: azadirachtin, a neem tree-derived insect antifeedant and growth disruptor); 2) Knack 0.86EC (an insect growth regulator insecticide); Endigo (a combination insecticide containing lambda-cyhalothrin [a pyrethroid insecticide] and thiamethoxam [a neonicotinoid]), and Manticor LFR (a combination product comprised of Capture LFR insecticide and Headline fungicide). Planting-time liquid products in this experiment were delivered in 3-inch T-bands over the open seed furrow by using a planter-mounted, CO₂-propelled spray system calibrated to deliver a finished spray volume output of 5 GPA through TeeJet™ 400067E nozzles.

Postemergence insecticide applications. Postemergence insecticide treatments in this experiment included the following sprayable liquid products: Captiva (an insect repellent comprised of capsicum [pepper] extract, garlic oil, and soybean oil), Dibrom Emulsive (a conventional organophosphate insecticide), Ecozin Plus 1.2%ME (azadirachtin), Evergreen Crop Protection 60-6EC (pyrethrum + a synergist), Veratran D (a botanical material containing insecticidal alkaloids from the *Sabadilla* plant), Warrior II (a pyrethroid insecticide with Zeon U.V.

protection), and Vydate C-LV (a carbamate), and all were compared with Lorsban Advanced as a postemergence chemical insecticide standard. All postemergence spray treatments were broadcast-applied on 9 June (i.e., about 1 day before peak SBRM fly activity). Sprays were applied from a tractor-mounted, CO₂-propelled spray system equipped with an 11-ft boom that was calibrated to deliver a finished spray volume output of 10 GPA through TeeJet™ 110015VS nozzles.

Root injury ratings: Sugarbeet root maggot feeding injury was assessed in this trial on 1 August by randomly collecting ten beet roots per plot (five from each of the outer two treated rows), hand-washing them, and scoring them in accordance with the 0 to 9 root injury rating scale (0 = no scarring, and 9 = over ¾ of the root surface blackened by scarring or dead beet) of Campbell et al. (2000).

Harvest: Treatment performance was also compared on the basis of sugarbeet yield parameters. Plots were harvested on 2 October. Foliage was removed from plots immediately before harvest by using a commercial-grade mechanical defoliator. All beets from the center two rows of each plot were extracted from the soil using a mechanical harvester, and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All data from root injury ratings and harvest samples were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance

Results and Discussion:

It is important to note that all insecticide entries in this trial were single-component control tools, which are not recommended in high-risk areas such as St. Thomas, where severe SBRM infestations are common. Another important aspect of this trial was that a hailstorm, including high winds and locally heavy rainfall, occurred on 9 June. This occurred just 2 days before peak fly was expected, and just a few hours after all postemergence spray treatments were applied. As such, the results of this trial should be interpreted with discretion.

Sugarbeet root maggot feeding injury results for this experiment are presented in Table 1. The average level of SBRM larval feeding injury recorded for the untreated check plots was only 5.33 on the 0 to 9 scale of Campbell et al. [2000]), which indicated that a moderate root maggot infestation developed in the plot area for this experiment.

Table 1. Larval feeding injury in an evaluation of experimental at-plant and postemergence sprays for sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Manticor LFR (bifenthrin + pyraclostrobin)	3" T-band	19 fl oz	0.2 lb bifenthrin + 0.1 lb pyraclostrobin	2.25 e
Counter 20G	B	7.5 lb	1.5	2.55 de
Endigo ZC	3" TB	4.5 fl oz		3.38 cd
Ecozin Plus 1.2% ME	2 d Pre-peak Broadcast	56 fl oz		4.00 bc
Lorsban Advanced	2 d Pre-peak Broadcast	1 pt	0.5	4.30 abc
Dibrom	2 d Pre-peak Broadcast	1 pt		4.33 abc
Evergreen Crop Protection	2 d Pre-peak Broadcast	16 fl oz		4.53 ab
Warrior II	2 d Pre-peak Broadcast	1.92 fl oz	0.03	4.68 ab
Knack 0.86 EC	3" TB	10 fl oz		4.70 ab
Vydate CLV	2 d Pre-peak Broadcast	34 fl oz	1.0	4.78 ab
Captiva	2 d Pre-peak Broadcast	2 pts		4.90 ab
Aza-Direct	3" TB	56 fl oz		5.08 ab
Veratran D	2 d Pre-peak Broadcast	20 lb	0.04	5.30 a
Check	---	---	---	5.33 a
LSD (0.05)				1.104

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; TB = T-band over open seed furrow

Entries that provided the greatest levels of root protection (i.e., lowest SBRM feeding injury ratings) included planting-time treatments of Manticor LFR (19 fl oz/ac) and Counter 20G at its moderate rate of 7.5 lb product/ac. Manticor outperformed all treatments, except Counter with regard to protection from larval feeding injury, and Endigo ZC (4.5 fl oz/ac) was the only other treatment that provided a level of root protection that was not

significantly different from Counter 20G. The only other treatment that provided a significant reduction in root maggot larval feeding injury when compared to the untreated check plots was Ecozin Plus, applied at 56 fl oz/ac.

Yield data from this trial are shown in Table 2. The highest-yielding treatments included the following: 1) Counter 20G, applied at a moderate rate of 7.5 lb product/ac; 2) Warrior II, applied as a postemergence broadcast at 1.92 fl oz/ac; 3) Manticor LFR, applied at 19 fl oz/ac in a 3-inch T-band at planting; and 4) Ecozin Plus, which was applied as a postemergence broadcast at 56 fl oz/ac. All of these treatments produced root yields of more than 34 tons/ac, which were all significantly greater than that recorded for the untreated check. The following treatments were not significantly outperformed by the top four treatments, and produced significantly more recoverable sucrose yield than the untreated check: 1) Vydate C-LV, applied postemergence at 34 fl oz/ac; 2) Dibrom Emulsive, broadcast-applied 1 pt product/ac; and 3) a postemergence spray of Veratran D at 20 lb product/ac.

It bears repeating that all insecticide-treated entries in this experiment were single-application treatments. Also, it should be noted that five of the top seven treatments in relation to recoverable sucrose and root yield are currently not registered for use in sugarbeet, and three of them represent alternative modes of action to the commonly used ACHE inhibitors. As such these results provide encouragement regarding the future of SBRM management. These alternatives, which included Warrior II and Manticor (both pyrethroid insecticides), Ecozin Plus (azadirachtin, a plant-derived insect antifeedant and growth disruptor), and Veratran D (a plant-derived insecticide containing Sabadilla alkaloids) generated recoverable sucrose yield increases ranging from 1,461 to 2,154 lb/ac above the average sucrose yield from the untreated check plots. Also, all of these treatments generated numerically (not statistically significant) more recoverable sucrose than Lorsban Advanced (the postemergence broadcast spray standard in this trial) and Counter 20G (the conventional planting-time standard). It should be noted that Counter 20G and Lorsban Advanced were both applied at their respective moderate rates, and not the maximum rates allowed on the respective labels of those products.

Table 2. Yield parameters from an evaluation of experimental at-plant and postemergence sprays for sugarbeet root maggot control, St. Thomas, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G	B	7.5 lb	1.5	11,446 a	37.1 a	16.58 a	1,358
Warrior II	2 d Pre-peak Broadcast	1.92 fl oz	0.03	10,917 ab	36.4 ab	16.28 a	1,244
Manticor LFR (bifenthrin + pyraclostrobin)	3" T-band	19 fl oz	0.2 lb bifenthrin + 0.1 lb pyraclostrobin	10,694 abc	33.8 abc	16.98 a	1,311
Ecozin Plus 1.2% ME	2 d Pre-peak Broadcast	56 fl oz		10,512 abc	34.2 abc	16.55 a	1,241
Vydate CLV	2 d Pre-peak Broadcast	34 fl oz		10,440 abc	33.2 bc	16.80 a	1,269
Dibrom	2 d Pre-peak Broadcast	1 pt		10,409 abc	34.2 abc	16.35 a	1,210
Veratran D	2 d Pre-peak Broadcast	20 lb	0.04	10,224 a-d	32.9 bc	16.60 a	1,223
Lorsban Advanced	2 d Pre-peak Broadcast	1 pt	0.5	10,070 b-e	32.8 bc	16.48 a	1,185
Captiva	2 d Pre-peak Broadcast	2 pts		10,069 b-e	33.4 abc	16.30 a	1,153
Evergreen Crop Protection	2 d Pre-peak Broadcast	16 fl oz		9,995 b-e	32.6 bc	16.45 a	1,175
Endigo ZC	3" TB	4.5 fl oz		9,988 b-e	33.0 bc	16.30 a	1,150
Knack 0.86 EC	3" TB	10 fl oz		9,500 cde	31.1 cd	16.43 a	1,112
Aza-Direct	3" TB	56 fl oz		8,965 de	28.7 d	16.70 a	1,080
Check	---	---	---	8,763 e	28.1 d	16.75 a	1,054
LSD (0.05)				1,353.7	3.82	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; TB = T-band over open seed furrow

It is encouraging that most of the alternative materials tested provided equivalent protection from SBRM feeding injury to that of the labeled chemical insecticides. Further testing should be carried out on these and other experimental materials to identify potential alternatives to the currently used insecticides. Alternative insecticide options could help prevent or delay the development of insecticide resistance in sugarbeet root maggot populations, and could also provide viable tools for growers to sustainably and profitably produce sugarbeet in SBRM-affected areas if the currently available conventional insecticides become unavailable due to regulatory action.

References Cited:

Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006. Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.

Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000. Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. *J. Sugar Beet Res.* 37: 57–69.

SAS Institute. 2008. *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

MOVENTO HL®: TWO YEARS OF PERFORMANCE TRIALS ON A NEWLY REGISTERED INSECTICIDE FOR SUGARBEET ROOT MAGGOT CONTROL

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Introduction:

The sugarbeet root maggot (SBRM), *Tetanops myopaeformis* (Röder) is a serious economic pest of sugarbeet in the Red River Valley (RRV) growing area. Sugarbeet producers in the RRV typically manage this pest by prophylactically applying granular insecticides to at-risk fields during planting operations. In areas where severe SBRM infestations frequently develop, planting-time control efforts are often augmented by one to two postemergence applications. As far back as the mid-1970s, most of these applications have involved the use of insecticides in the organophosphate and carbamate classes to manage the sugarbeet root maggot. Both of these classes cause mortality in insects through the same mode of action, acetylcholinesterase (ACHE) inhibition.

Grower dependence on a single mode of action for SBRM control in the Red River Valley has been largely due to two factors. First, a limited number of insecticide products have been registered for use in the crop for much of this time. Second, despite frequent screening efforts on a variety of insecticides belonging to alternative modes of action, very few insecticidal products tested in screening programs have shown promise as viable options for SBRM control. As a result of this long-term, repeated use of ACHE inhibitor insecticides, the threat of insecticide resistance development in RRV sugarbeet root maggot populations has been a looming concern for pest management advisors and producers for several years.

In July of 2017, the U.S. Environmental Protection Agency approved the registration of Movento HL insecticide for use in sugarbeet. The addition of this product is encouraging from an insect resistance management perspective, because the active ingredient in Movento (spirotetramat) belongs to the lipid biosynthesis inhibitors (LBIs), which will provide an alternative mode of action to the commonly used ACHE inhibitors. Thus far, after significant screening efforts have been conducted on insect species with known resistance to other insecticides, there is no evidence of cross resistance between the LBI insecticides and other classes.

This project was carried out to evaluate the efficacy of Movento HL as a postemergence tool for sugarbeet root maggot control. A secondary objective was to assess the performance of dual-insecticide programs for SBRM management that include Poncho Beta as the planting-time insecticide component and Movento HL as the postemergence rescue component.

Materials and Methods:

This experiment was conducted during the 2016 and 2017 growing seasons on commercial sugarbeet field sites near St. Thomas in rural Pembina County, ND. Betaseed 89RR52 glyphosate-resistant seed was used for all treatments in both study years. Plots were planted on 11 May in 2016 and 10 May in 2017. All plots were planted using a 6-row Monosem NG Plus 4 7x7 planter set to deliver seed at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Plots were six rows (22-inch spacing) wide with the four centermost rows treated. The outer “guard” row on each side of the plot served as an untreated buffer. Each plot was 35 feet long, and 35-foot tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design. Treatments were replicated four times in 2016 and three times in 2017.

Planting-time insecticide applications. Planting-time applications of Counter 20G were applied by using band (B) placement (Boetel et al. 2006), which consisted of 5-inch swaths of granules delivered through Gandy™ row banders. Granular application rates were regulated by using planter-mounted SmartBox™ computer-controlled insecticide delivery system that had been calibrated on the planter before all applications.

Postemergence insecticide applications. Additive postemergence insecticides applied in this trial included Movento HL, Movento 240SC, Lorsban Advanced, and Mustang Maxx. The original (i.e., 240SC) formulation of Movento was included in the trial for comparative purposes because it had been included in previous NDSU screening trials before the HL formulation was available for testing. Treatment timings evaluated included the following: 1) Lorsban Advanced and Mustang Maxx at two days before peak SBRM fly activity; 2) Movento 240SC and one Movento HL entry at seven days pre -peak; and 3) Movento HL on or within one day of peak fly activity. Liquid insecticide solutions were delivered with a tractor-mounted CO₂-propelled spray system equipped with TeeJet™ 110015VS nozzles calibrated to deliver applications in a finished output volume of 10 GPA. All postemergence Movento spray solutions included methylated seed oil at the recommended rate of 0.25% v/v.

Root injury ratings: Sugarbeet root maggot feeding injury was assessed in this experiment on 3 and 1 August in 2016 and 2017, respectively. Sampling consisted of randomly collecting ten beet roots per plot (five from each of the outer two treated rows), hand-washing them, and scoring them in accordance with the 0 to 9 root injury rating scale (0 = no scarring, and 9 = over ¾ of the root surface blackened by scarring or dead beet) of Campbell et al. (2000).

Harvest: Treatment performance was also compared on the basis of sugarbeet yield parameters. Plots were harvested on 20 September in 2016, and on 3 October in 2017. Foliage was removed from plots immediately before harvest by using a commercial-grade mechanical defoliator. All beets from the center two rows of each plot were extracted from soil using a mechanical harvester, and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All data from root injury ratings and harvest samples were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008). Treatment means were compared by using Fisher’s protected least significant difference (LSD) test at a 0.05 level of significance. Initial analyses indicated that there were no significant treatment × year interactions for root injury ratings ($P = 0.7445$), recoverable sucrose yield ($P = 0.2636$), root yield ($P = 0.1345$), or percent sucrose content data ($P = 0.4321$). As such, two-year combined analyses were performed on all data from this experiment.

Results and Discussion:

Sugarbeet root maggot feeding injury results from this two-year trial are presented in Table 1. The feeding injury rating mean for the untreated check (5.24 on the 0 to 9 scale of Campbell et al. [2000]) indicated the presence of a moderate SBRM larval infestation across both years. However, feeding injury recorded in all insecticide-protected plots was significantly lower than that in the untreated check.

Table 1. Larval feeding injury in a comparison of Movento HL®, Lorsban Advanced, and Mustang Maxx for postemergence sugarbeet root maggot control, St. Thomas, ND, 2016 – 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Root injury (0-9)
Counter 20G	B	7.5 lb	1.5	3.27 d
Poncho Beta + Mustang Maxx	Seed 2 d Pre-peak Broadcast	4 fl oz	68 g a.i./ unit seed 0.025	3.29 d
Poncho Beta + Movento 240SC + MSO	Seed 7 d Pre-peak Broadcast	5 fl oz	68 g a.i./ unit seed 0.078	3.51 cd
Poncho Beta + Lorsban Advanced	Seed 2 d Pre-peak Broadcast	2.0 pts	68 g a.i./ unit seed 1.0	3.59 bcd
Poncho Beta + Movento HL + MSO	Seed Peak fly	2.5 fl oz	68 g a.i./ unit seed 0.078	4.24 bc
Poncho Beta	Seed		68 g a.i./ unit seed	4.27 bc
Poncho Beta + Movento HL + MSO	Seed 7 d Pre-peak Broadcast	2.5 fl oz	68 g a.i./ unit seed 0.078	4.34 b
Check	----	----	----	5.24 a
LSD (0.05)				0.763

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher’s Protected LSD test).

^aB = banded at planting; Seed = insecticidal seed treatment

The lowest average root maggot feeding injury was observed in plots protected by the single at-plant application of Counter 20G at its moderate (7.5 lb product/ac) rate. Other entries that were not significantly outperformed by this treatment included the following: 1) Poncho Beta + a postemergence application of Mustang Maxx at 4 fl oz of product/ac; 2) Poncho Beta plus a postemergence application of Movento 240SC at 5 fl oz of product/ac (7 days pre-peak); and 3) Poncho Beta seed treatment plus a postemergence application of Lorsban Advanced at its high (2 pts product/ac) labeled rate. There was no significant difference in SBRM feeding injury between applications of Movento HL made at peak fly and seven days pre-peak.

Yield data from this experiment are shown in Table 2. Similar to the results from root ratings, all insecticide treatments provided significant increases in both recoverable sucrose yield and root tonnage. The top-performing treatment with regard to recoverable sucrose and root yield was the combination of Poncho Beta seed treatment plus a postemergence application of Lorsban Advanced. When compared to the untreated check, that entry produced 2,416 lb more recoverable sucrose and 7.4 additional tons per acre in root yield, and generated a revenue benefit of \$352/ac. Treatments that were not significantly different from the top treatment with regard to both recoverable sucrose yield and root tonnage included Poncho Beta plus Mustang Maxx and Poncho Beta plus Movento HL (applied at seven days ahead of peak SBRM fly activity).

Applying Movento HL at seven days ahead of peak fly to plots initially protected by Poncho Beta seed treatment generated an increase in revenue of \$79/acre when compared to Poncho Beta plots that did not receive a postemergence spray. Although there were no significant differences in coverable sucrose yield or root tonnage between the two Movento HL postemergence spray timings, applying this insecticide earlier (seven days pre-peak) generated \$69 more gross revenue than when it was applied at peak SBRM fly activity. Gross economic return increases from insecticide-based programs in this experiment ranged from \$165/ac for Poncho Beta plus Movento 240SC at postemergence to the aforementioned \$352/ac for the treatment that included Poncho Beta-treated seed plus a postemergence application of Lorsban Advanced.

Table 2. Yield parameters from a comparison of Movento HL[®], Lorsban Advanced, and Mustang Maxx for postemergence sugarbeet root maggot control, St. Thomas, ND, 2016 – 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Poncho Beta + Lorsban Advanced	Seed 2 d Pre-peak Broadcast	2.0 pts	68 g a.i./ unit seed 1.0	8,039 a	26.7 a	15.99 a	1,063
Poncho Beta + Mustang Maxx	Seed 2 d Pre-peak Broadcast	4 fl oz	68 g a.i./ unit seed 0.025	7,885 a	25.9 a	15.96 a	1,053
Poncho Beta + Movento HL + MSO	Seed 7 d Pre-peak Broadcast	2.5 fl oz	68 g a.i./ unit seed 0.078	7,409 ab	24.9 ab	15.69 a	961
Poncho Beta + Movento HL + MSO	Seed Peak fly Broadcast	2.5 fl oz	68 g a.i./ unit seed 0.078	6,923 b	23.4 b	15.43 a	892
Counter 20G	B	7.5 lb	1.5	6,877 b	23.1 b	15.66 a	894
Poncho Beta	Seed		68 g a.i./ unit seed	6,865 b	23.3 b	15.49 a	882
Poncho Beta + Movento 240SC + MSO	Seed 7 d Pre-peak Broadcast	5 fl oz	68 g a.i./ unit seed 0.078	6,841 b	23.3 b	15.49 a	876
Check	----	----	----	5,623 c	19.3 c	15.27 a	711
LSD (0.05)				755.5	2.22	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; Seed = insecticidal seed treatment

There were no significant differences in percent sucrose content between any of the treatments in this trial, but the untreated check had the lowest sucrose concentration, and roots from the treatment that generated the highest root tonnage and sucrose yield (Poncho Beta + Lorsban Advanced) also had numerically higher percent sucrose content than any other treatment in the experiment.

Overall, results from this two-year experiment demonstrate that, even under moderate SBRM infestation, major yield and revenue benefits can be achieved in insecticide-based control programs combining a neonicotinoid seed treatment insecticide and a postemergence sprayable insecticide such as Lorsban Advanced. Results also suggest that yields and revenue are markedly increased by adding a postemergence spray. Major yield increases were also achieved by applying Mustang Maxx at 2 days before peak fly and Movento HL at seven days pre-peak.

Although there were no significant differences in regard to root protection from SBRM feeding activity or resulting yield parameters between the two timings tested for Movento HL applications, results also suggest slight yield improvements by applying this product earlier. This pattern may have been associated with the fact that

Movento is a systemic insecticide. As such, applying it earlier may have resulted in higher concentrations of insecticide active ingredient in roots when SBRM larval feeding injury was occurring.

Further research is needed to evaluate Movento HL under higher SBRM infestation levels to determine its ability to effectively control this pest. Additional research should focus on optimizing the effectiveness of application timing and use rate. The EPA-approved label allows for a much higher application rate of 4.5 fl oz/ac. However, at this time, it is uncertain as to whether applying this product at its maximum labeled rate, even if shown to be effective, would be economically viable.

References Cited:

- Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006.** Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.
- Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000.** Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. *J. Sugar Beet Res.* 37: 57–69.
- SAS Institute. 2008.** *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

WIREWORM MANAGEMENT IN SUGARBEET USING PLANTING-TIME GRANULAR, LIQUID, AND SEED TREATMENT INSECTICIDES

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Introduction:

Wireworms occasionally cause significant plant stand and yield loss in Red River Valley (RRV) sugarbeet fields. They also can be problematic for producers in all other sugarbeet production areas of North America. Wireworms are the larval stage of insects commonly referred to as “click beetles”, and about three wireworm species are important pests of several North American field crops. Wireworm infestations are difficult to predict because the most common pest species of this group have between 3- and 5-year life cycles, and populations within an individual field can be at various stages within their life cycle.

For several decades, RRV sugarbeet producers mostly relied on prophylactic applications of planting-time granular insecticides to protect fields from a suite of soil-dwelling insects that threaten the profitability of sugarbeet production, including wireworms, the sugarbeet root maggot, springtails, and white grubs. More recently, growers have also had the option to use a seed-applied or sprayable liquid insecticide to protect crops from soil-inhabiting insect pests. Due to the aforementioned variability and unpredictability of wireworm infestations in North American field crop systems, the current body of literature lacks comprehensive data on the efficacy of insecticides against these pests. This experiment was carried out to compare at-plant granular, liquid, and seed-applied insecticides as tools to control wireworms in sugarbeet.

Materials & Methods:

The site chosen for this experiment was an established grower-owned sugarbeet field near Manvel, ND that had an infestation of about 1.2 wireworms per plant. Plots were planted on 20 June, 2017 by using a 6-row Monosem NG Plus 7x7 planter set to plant at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Betaseed 89RR52, a glyphosate-tolerant seed variety, was used for all treatments. Individual treatment plots were two rows (22-inch spacing) wide and 25 feet long, and 20-ft wide tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design with four replications of the treatments. Two-row plots are the preferred experimental unit size in wireworm trials because infestations of these pests are often patchy within a field. As such, a smaller test area increases the likelihood of having a sufficiently uniform wireworm infestation among plots within each block.

Insecticidal seed treatment materials were applied to seed by Germain’s Technology Group (Fargo, ND). Granular insecticide treatments were applied by using band placement (Boetel et al. 2006), which consisted of 5-inch swaths that were delivered through Gandy™ row banders. Output rates of the planting-time standard granular material used this experiment were regulated by using a planter-mounted SmartBox™ computer-controlled insecticide delivery system that was calibrated on the planter immediately before all applications. Mustang Maxx was delivered in 3-inch T-bands over the open seed furrow by using a planter-mounted, CO₂-propelled spray system calibrated to deliver a finished spray volume output of 5 GPA through TeeJet™ 400067E nozzles.

Treatment efficacy was compared for plant stand data and yield parameters because wireworm larval feeding injury causes stand losses that can lead to yield reductions. Stand counts involved counting all living plants within each 25-ft long row. Plant stand counts were taken on 30 June, and 7, 13, and 27 July, 2017, which were 10, 17, 23, and 37 days after planting (DAP), respectively. Raw stand counts were converted to plants per 100 linear row ft for the analysis. Plots were harvested on 9 October by using a 2-row mechanical harvester to collect all beets from both rows of each plot. Subsamples of 12-18 harvested beets were sent to the American Crystal Sugarbeet Quality Laboratory (East Grand Forks, MN) for quality analyses. Stand and yield data were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher’s protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

Results from plant stand counts for this trial are shown in Table 1. There were no significant differences among treatments at the initial stand count (10 DAP). However, at the second and third stand count dates (17 and 23 DAP), all insecticide-treated plots had significantly greater numbers of surviving plants than the untreated check plots, and there were no significant differences among insecticide-protected treatments.

Table 1. Plant stand counts from evaluation of planting-time granular, liquid, and seed treatment insecticides for wireworm control, Manvel, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb ai/ac)	Stand count ^b (plants / 100 ft)			
				10 DAP ^c	17 DAP ^c	23 DAP ^c	37 DAP ^c
Poncho Beta	Seed		68 g a.i./ unit seed	168 a	213 a	206 a	216 a
Counter 20G	B	5.9 lb	1.2	173 a	209 a	208 a	206 ab
Counter 20G	B	4.5 lb	0.9	173 a	209 a	203 a	196 ab
Mustang Maxx	3" T-band	4 fl oz	0.025	173 a	199 a	200 a	193 ab
NipsIt Inside	Seed	----	60 g a.i./ unit seed	170 a	205 a	200 a	190 b
Counter 20G	B	7.5 lb	1.5	159 a	192 a	199 a	194 ab
Cruiser 5FS	Seed	----	60 g a.i./ unit seed	148 a	190 a	198 a	193 ab
Check	---	----	---	126 a	151 b	148 b	134 c
LSD (0.05)				NS	24.6	25.1	24.2

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; T-band = 3" swath over open seed furrow at planting; Seed = insecticidal seed treatment

^bSurviving plant stands were counted on June 30 and on July 7, 13, and 27, 2017 (10, 17, 23, and 37 days after planting, respectively).

^cDAP = Days after planting

The effects of wireworm feeding on plant roots were more evident by the fourth stand count (37 DAP), when plots planted with Poncho Beta-treated seed had the highest average plant stands in the study. Poncho Beta plots had significantly greater plant stands than the untreated check plots and those planted with NipsIt Inside-treated seed, but they were not statistically different from any other insecticide-treated entry. All insecticide treatments, including NipsIt Inside seed treatment, had significantly greater plant densities per 100 row feet than the untreated check, irrespective of whether they were protected by a planting-time granular, sprayable liquid, or insecticidal seed treatment.

Yield results from this trial are presented in Table 2. All insecticide treatments provided significant increases in both recoverable sucrose yield and root tonnage when compared to yields recorded for the untreated check. There were no significant differences among any of the insecticide-protected treatments, however, plots treated with the lowest rate of Counter 20G (4.5 lb product/ac) generated numerically greater recoverable sucrose than any other insecticide-protected plots in the trial. Revenue benefits from Counter 20G, in comparison to revenue from the untreated check, ranged from \$58/ac for the 5.9-lb/ac rate to \$110/ac for the 4.5-lb rate. Seed treatment insecticides provided gross economic return increases that ranged from \$89/ac in Poncho Beta plots to \$111/ac for plots protected by NipsIt Inside. The gross economic return benefit from applying Mustang Maxx averaged \$76/ac.

Table 2. Yield parameters from evaluation of planting-time granular, liquid, and seed treatment insecticides for wireworm control, Manvel, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G	B	4.5 lb	0.9	6,692 a	26.7 a	14.68 a	544
Counter 20G	B	5.9 lb	1.2	6,516 a	26.7 a	14.40 a	492
Poncho Beta	Seed	----	68 g a.i./ unit seed	6,438 a	25.7 ab	14.63 a	523
Cruiser 5FS	Seed	----	60 g a.i./ unit seed	6,430 a	25.3 bc	14.70 a	538
NipsIt Inside	Seed	----	60 g a.i./ unit seed	6,396 a	25.0 bc	14.83 a	545
Counter 20G	B	7.5 lb	1.5	6,268 a	24.9 bc	14.73 a	515
Mustang Maxx	3" T-band	4 fl oz	0.025	6,146 a	24.3 c	14.73 a	510
Check	---	----	---	5,415 b	21.7 d	14.55 a	434
LSD (0.05)				562.7	1.29	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; T-band = 3" swath over open seed furrow at planting; Seed = insecticidal seed treatment

It should be noted that this trial was planted atypically late in the growing season because the trial was initiated subsequent to the grower detecting a wireworm infestation in an established sugarbeet field. As is typical with sugarbeet research plots, this study was also harvested over two weeks earlier in the season than a typical grower field would be harvested. As such, the resulting sucrose yield, root tonnage, and percent sucrose content values are much lower than would be experienced by a commercial producer. However, these findings provide an excellent window into the significance of wireworms as serious sugarbeet pests and effective tools with which to control them.

Overall, the findings from this trial clearly indicate that wireworms can cause significant harm to sugarbeet seedlings, and the effects result in major yield and revenue losses. Effective wireworm management in this late-planted trial resulted in major increases in gross revenue that would have easily paid for the associated investments and provided significant net revenue benefits. As such, growers managing fields with known wireworm infestation histories should consider the use of one of these prophylactic tools to protect their crops.

References Cited:

Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006. Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.

SAS Institute. 2008. *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

SPRINGTAIL CONTROL IN SUGARBEET: A COMPARISON OF GRANULAR, SPRAYABLE LIQUID, AND SEED-APPLIED INSECTICIDES

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Introduction:

Springtails belong to the order Collembola, an order of organisms that is so unique that they are considered by many experts to belong to a separate taxonomic group from that of true insects. Subterranean (soil-dwelling) springtails have been recognized as a serious pest threat of sugarbeet for many growers in the central and southern Red River Valley (RRV) of Minnesota and North Dakota since the late-1990s. Producers in western ND and eastern Montana also frequently have problems with springtails. These tiny, nearly microscopic, blind, and wingless insects spend their entire lives below the soil surface (Boetel et al. 2001).

Although subterranean springtails are present in many fields throughout the RRV, they only occasionally become a major pest problem. Subterranean springtails thrive in heavy soils with high levels of soil organic matter. Cool and wet weather can be conducive to buildups of springtail infestations because such conditions slow sugarbeet seed germination and seedling development, which renders plants extremely vulnerable to attack by springtails that are not negatively impacted by cool temperatures. Therefore, these pests can cause major stand and yield losses. We conducted a field experiment to evaluate the performance of a conventional granular insecticide, an experimental at-plant liquid insecticide, and three insecticidal seed treatments for springtail control in sugarbeet.

Materials & Methods:

This experiment was established on the NDSU experiment farm near Prosper, ND. Plots were planted on 19 May, 2017 using a 6-row Monosem NG Plus 7x7 planter set to plant at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Betaseed 89RR52, a glyphosate-tolerant seed variety, was used for all treatments. Individual treatment plots were two rows (22-inch spacing) wide and 25 feet long, and 20-ft wide tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a randomized complete block design with four replications of the treatments. Two-row plots are the preferred experimental unit size in springtail trials because infestations of these pests are typically patchy. A smaller test area increases the likelihood of having a sufficiently uniform springtail infestation among plots within each block.

Insecticidal seed treatment materials were applied to seed by Germain's Technology Group (Fargo, ND). Granular insecticide treatments were applied by using band placement (Boetel et al. 2006), which consisted of 5-inch swaths that were delivered through Gandy™ row banders. Output rates of the planting-time standard granular material used this experiment were regulated by using a planter-mounted SmartBox™ computer-controlled insecticide delivery system that was calibrated on the planter immediately before all applications. Manticor LFR was applied in 3-inch T-bands over the open seed furrow by using a planter-mounted, CO₂-propelled spray system calibrated to deliver a finished spray volume output of 5 GPA through TeeJet™ 400067E nozzles.

Treatment efficacy was compared by using plant stand counts and yield parameters because subterranean springtails cause stand losses that lead to yield reductions. Stand counts involved counting all living plants within each 25-ft long row. Plant stand counts were taken on June 1, 15 and 29, as well as 7 July, which were 13, 27, 41, and 49 days after planting (DAP), respectively. Raw stand counts were converted to plants per 100 linear row ft for the analysis. Plots were harvested on 18 September by using a 2-row mechanical harvester to collect all beets from both rows of each plot. Subsamples of 12-18 harvested beets were sent to the American Crystal Sugarbeet Quality Laboratory (East Grand Forks, MN) for quality analyses. All stand count and yield data were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

Plant stand count data for this trial are presented in Table 1. At the initial stand count date (13 DAP), the insecticide-protected plots had numerically greater numbers of surviving plants per 100 ft of row, but there were no significant differences among treatments, including the untreated check. However, at 27 DAP, all insecticide treatments except Cruiser 5FS resulted in significantly greater plant stands than the untreated check. The following

treatments had significantly greater plant stands than both Cruiser and the check at 27 DAP: 1) Poncho Beta; 2) Counter 20G at 4.5 lb product/ac; and 3) Mantikor LFR applied at 19 fl oz/ac.

Stand count comparisons for both 41 and 49 DAP generated the same results in that all insecticide treatments provided significant levels of protection from stand loss associated with springtail feeding injury when compared to the untreated check, irrespective of whether a granular, sprayable liquid, or seed treatment insecticide was used. Additionally, there were no significant differences among insecticide treatments at either 41 or 49 DAP.

Table 1. Plant stand counts from evaluation of planting-time granular, liquid, and seed treatment insecticides for springtail control, Prosper, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb ai/ac)	Stand count ^b (plants / 100 ft)			
				13 DAP ^c	27 DAP ^c	41 DAP ^c	49 DAP ^c
Poncho Beta	Seed		68 g a.i./ unit seed	166 a	176 a	198 a	203 a
Counter 20G	B	4.5 lb	0.9	159 a	171 a	191 a	183 a
Counter 20G	B	5.9 lb	1.2	158 a	169 ab	184 a	190 a
Mantikor LFR (bifenthrin + pyraclostrobin)	3" T-band	19 fl oz	0.2 lb bifenthrin + 0.1 lb pyraclostrobin	141 a	172 a	196 a	199 a
Cruiser 5FS	Seed		60 g a.i./ unit seed	122 a	129 bc	172 ab	169 ab
Check	---	----	---	117 a	127 c	150 b	148 b
LSD (0.05)				NS	39.7	30.7	34.6

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; T-band = 3" swath over open seed furrow at planting; Seed = insecticidal seed treatment

^bSurviving plant stands were counted on June 1, 15, and 29, and on July 7, 2017 (i.e., 13, 27, 41, and 49 days after planting, respectively).

^cDAP = Days after planting

Yield results from this experiment are presented in Table 2. General treatment performance patterns were similar to those observed in stand count results. Both rates of Counter 20G, as well as Poncho Beta seed treatment, resulted in significantly greater recoverable sucrose yield than the untreated check, and there were no significant differences among these three treatments with regard to recoverable sucrose. Cruiser 5FS seed treatment and Mantikor LFR were the only treatments that did not provide a significant increase in recoverable sucrose yield when compared to the untreated check. However, there were no significant differences in recoverable sucrose yield or root yield between Poncho Beta and Cruiser. Plots protected with the moderate rate of Counter 20G (5.9 lb product/ac) generated the highest tonnage in the trial, but that treatment was not significantly superior to the lower rate of 4.5 lb/ac. Additionally, both Counter 20G treatments were the only entries in this experiment that resulted in significant increases in root yield when compared to the untreated check.

Table 2. Yield parameters from evaluation of planting-time granular, liquid, and seed treatment insecticides for springtail control, Prosper, ND, 2017

Treatment/form.	Placement ^a	Rate (product/ac)	Rate (lb a.i./ac)	Sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose (%)	Gross return (\$/ac)
Counter 20G	B	5.9 lb	1.2	11,139 a	34.9 a	17.65 a	1,377
Counter 20G	B	4.5 lb	0.9	9,927 ab	31.9 ab	17.18 a	1,192
Poncho Beta	Seed		68 g a.i./ unit seed	9,725 abc	28.1 bc	18.55 a	1,321
Mantikor LFR (bifenthrin + pyraclostrobin)	3" T-band	19 fl oz	0.2 lb bifenthrin + 0.1 lb pyraclostrobin	8,979 bcd	26.6 bc	18.28 a	1,189
Cruiser 5FS	Seed		60 g a.i./ unit seed	8,278 cd	23.8 c	18.68 a	1,130
Check	---	----	---	8,266 d	23.9 c	18.63 a	1,122
LSD (0.05)				1,452.8	6.65	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aB = banded at planting; T-band = 3" swath over open seed furrow at planting; Seed = insecticidal seed treatment

Gross economic return results followed similar patterns to those for recoverable sucrose and root yields; however, percent sucrose influenced these patterns. Plots treated with the moderate rate of Counter 20G (5.9 lb product/ac) generated \$1,377/ac in gross revenue, which was \$185/ac greater than that from plots treated with the low (4.5 lb) rate of Counter. Similarly, plots treated with the 5.9-lb rate of Counter generated \$56/ac more gross revenue than Poncho Beta plots, and \$247/ac more revenue than plots planted with Cruiser-treated seed. An additional positive finding from this trial was that plots protected with the experimental material, Mantikor LFR, generated an average revenue increase of \$67/ac when compared to the untreated check plots. The increases in yield and revenue generated by insecticide treatments tested in this experiment show that effective tools are available for

managing subterranean springtails in sugarbeet. These findings also demonstrate the significance of subterranean springtails as serious economic pests of sugarbeet and demonstrate the importance of effectively managing them.

References Cited:

Boetel, M. A., R. J. Dregseth, and M. F. R. Khan. 2001. Springtails in sugarbeet: identification, biology, and management. Extension Circular #E-1205, North Dakota State University Coop. Ext. Svc.

Boetel, M. A., R. J. Dregseth, A. J. Schroeder, and C. D. Doetkott. 2006. Conventional and alternative placement of soil insecticides to control sugarbeet root maggot (Diptera: Ulidiidae) larvae. *J. Sugar Beet Res.* 43: 47–63.

SAS Institute. 2008. *The SAS System for Windows. Version 9.2.* SAS Institute Inc., 2002-2008. Cary, NC.

IMPACTS OF SEED LUBRICANTS ON SEEDLING ESTABLISHMENT AND YIELD: ON-FARM AND SMALL-PLOT TRIALS

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Introduction:

Neonicotinoid seed treatment insecticides (e.g., Cruiser, Poncho, etc.) have been implicated in honey bee kills near corn production fields in Indiana (Krupke et al. 2012). Those authors observed that planter hopper box seed-flow lubricants (e.g., talc) abrade seed-applied insecticides from corn seed coatings, and suggested that the resulting insecticide-laden dust is released into the air in exhaust plumes emitted from vacuum-based planters. As a result, they concluded that this subsequently can either directly or indirectly expose bees and potentially other pollinators. The findings from that research have precipitated public demands ranging from additional use restrictions to a complete ban on all uses of neonicotinoid seed treatment insecticides. In response to public concerns and perceived risk to pollinators from these insecticides, the Environmental Protection Agency (EPA) issued a moratorium on any new uses of currently labeled neonicotinoid products in April of 2015.

Concerns surrounding this phenomenon have also raised questions as to whether talcum or other seed-flow lubricants are necessary during row crop planting. If lubricants are not needed in sugarbeet planting, or if a less-abrasive alternative than talcum could perform at least as well without negatively impacting seed delivery and seedling establishment, it may provide evidence to support continued federal registration of neonicotinoid seed treatment insecticides used in sugarbeet production.

This experiment was carried out to determine if seed-flow lubricants (i.e., talc, graphite, talc/graphite mixture, Fluency AgentTM, or Fluency AdvancedTM [referred to in previous reports as “Fluency II”]) impact seed delivery, seedling establishment, or resulting sugarbeet yield parameters and revenue. This research could provide critical information to argue for maintaining neonicotinoid seed treatment registrations for use in sugarbeet if the EPA proposes a ban on using these materials in row crop production.

Materials and Methods:

This research involved two experiments that were carried out in grower-owned fields during the 2017 growing season. Study I involved a small-plot, replicated trial that was conducted near Hillsboro, ND. Study II was a large on-farm trial that was carried out by using conventional grower-owned equipment for planting and harvest. All seed-flow lubricant materials were applied at manufacturer-recommended rates.

Study I (small-plot trial): Plots were planted on 15 May, 2017 by using a 6-row John Deere MaxEmerge IITM planter. The planter was adjusted to deliver seed at a depth of 1¼ inch and a rate of one seed every 4½ inches of row length. Treatments in Study I included the following: 1) John Deere Premium Seed TalcTM (Deere & Co., Moline, IL); 2) John Deere Powdered GraphiteTM; 3) John Deere Talc/GraphiteTM combination seed lubricant (80% talc and 20% graphite); 4) Fluency AgentTM (Bayer Crop Science, Durham, NC); 5) Fluency AdvancedTM (a reformulated version from Bayer; NOTE: this was referred to as “Fluency II in previous reports); and a no-lubricant control. Betaseed 83CN, a glyphosate-resistant sugarbeet seed variety in two sizes (miniature pellet ~9/64-inch diam.) and Pro200, an extra-large pellet (~12.5/64-inch diam.) was used for the experiment. All seed included Poncho Beta (i.e., clothianidin + betacyfluthrin at 60:8 g a.i./100,000 seeds, respectively) insecticidal seed treatment to minimize the risk of soil insect feeding injury introducing unwanted variability to the experiment. Each plot was six rows (22-inch spacing) wide with the four centermost rows treated. The outer “guard” rows, one on the outer side of each plot, served as untreated buffer rows. Each plot was 35 feet long, and 25-foot tilled alleys were maintained between replicates throughout the growing season. The experiment was arranged in a split-plot design

with three replications of the treatments. Seed size was the whole-plot factor, and seed flow lubricant served as the sub-plot factor.

Treatment performance was compared using plant stand counts and yield parameters. Stand counts were made on 1, 15, and 29 June, 2017, which were 17, 31, and 45 days after planting (DAP), respectively. Those assessments involved counting all living plants in all four 35-ft long rows of each plot. Raw stand count observations were converted to plants per 100 linear row ft for the analyses.

Harvest: Treatment performance was also compared on the basis of sugarbeet yield parameters. The small-plot trial was harvested on 19 September, 2017. Immediately before harvest, the foliage was removed from all treatment plots by using a commercial-grade mechanical defoliator. After defoliation, all beets from the center two rows of each plot were extracted from the soil using a mechanical harvester and weighed in the field using a digital scale. A representative subsample of 12-18 beets was collected from each plot and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analysis.

Data analysis: All stand count and harvest data were initially subjected to analysis of variance (ANOVA) (SAS Institute, 2008) to determine whole- and sub-plot factor effects. All mean comparisons were carried out by using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance. The initial ANOVAs for the small-plot study at Hillsboro indicated no significant seed size \times lubricant ($P > 0.05$) interactions for any of the three stand count dates or any of the yield parameters. As such, combined analyses was conducted to compare seed flow lubricants on the basis of stand count and yield data averaged across both seed sizes.

Study II (on-farm trial): The on-farm trial was planted on 1 May, 2017 by using a 12-row John Deere 1730 MaxEmerge Plus™ planter. The planter was operated at 4.5 mph, and adjusted to deliver seed 1¼ inch deep at a rate of one seed every 4¾ inches of row length. Betaseed 8572 seed was used for planting all treatments. All seed was formulated as miniature pellets, and was prepared with the following seed-applied protectants: 1) Poncho Beta insecticide (68 g a.i./100,000-seed unit); 2) Tachigaren fungicide (45 g a.i./ unit); and 3) Kabina ST fungicide (14 g a.i./unit). All plots were also protected against seedling insect pests by applying Counter 20G at 5.9 lb product/ac in a modified (i.e., restricted to prevent granule deposition into seed furrow) band. The entire field also received a planting-time application of 10-34-0 (respective percentage of N, P, and K) starter fertilizer.

Study II included all treatments used in Study I, except the original formulation of Fluency Agent™. Each individual treatment plot was 12 rows (22-inch spacing) wide by 600 ft in length. The experiment was arranged in a randomized complete block design with three replications of the treatments.

Treatment performance was compared according to plant stand counts and yield parameters. Stand counts involved counting all living plants within 1/1000th ac long subsamples, of which four were taken at equally spaced intervals within the length of each treatment plot. Counts were taken on 24 May, and 1 and 22 June, which were 23, 31, and 52 days after planting (DAP), respectively. All plant stand count observations were converted to plants per 100 linear row ft before being subjected to statistical analysis.

Harvest: Sampling for harvest data was conducted twice in this experiment. Hand-harvested yield samples were collected on 12 October, 2017. Conventional drain spades were used to manually dig the samples, and each was comprised of all roots from within an 11.9-ft length of the same center-most row of each plot. One sample was collected at each of four locations within each treatment plot. Pre-harvest samples were collected at the same locations within each plot that stand counts were taken throughout the growing season. Samples were bagged and labeled, and sent to the American Crystal Sugar Company Tare Laboratory (East Grand Forks, MN) for sucrose content and quality analyses.

Machine harvesting procedures, which were carried out on 16 October, 2017, first involved removal of foliage from all treatment plots by using the grower's commercial-grade mechanical defoliator. Harvesting consisted of collecting all roots from each treatment plot with a conventional six-row Art's Way™ 690 sugarbeet harvester. Site-specific root tonnage data was collected from the on-board yield monitoring system in 50-ft increments from within each plot. Quality analysis parameters from pre-harvest samples were used in combination with tonnage data from the harvester yield monitor to calculate recoverable sucrose yield from each plot.

Data analysis: All machine-harvest yield data were subjected to analysis of variance (ANOVA) using the general linear models (GLM) procedure (SAS Institute, 2008), and treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

Results and Discussion:

Study I (small-plot trial): Plant stand count results from the initial whole-plot (i.e., seed size) treatment comparisons appear in Table 1. On the first date (17 DAP), plant populations in plots planted with Pro200 (i.e., extra-large) seed were significantly greater than those in plots seeded with miniature pellets. That was the only date on which there was a significant difference in plant population between the two seed sizes tested. The stand counts taken at 31 and 45 DAP indicated that plant populations for the two seed sizes were nearly identical, with only numerical differences between treatments of only three plants per 100 row ft. The relatively small difference between seed sizes during the first stand count, combined with the fact that no significant differences were detected on subsequent dates, suggested that seed size did not play a major role in the results of this trial.

Table 1. Whole-plot effect of seed size on plant population in a comparison of sugarbeet seed-flow lubricants in a small-plot field trial (Study I), Hillsboro, ND, 2017

Treatment/ form.	Stand counts ^a (plants / 100 row ft)		
	17 DAP ^b	31 DAP	45 DAP
Pro200	237 a	270 a	270 a
Mini	205 b	263 a	267 a
LSD (0.05)	20.8	NS	NS

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aSurviving plant stands were counted on June 1, 15, and 29, 2017, which were 17, 31, and 45 days after planting (DAP), respectively.

^bDAP = days after planting

Yield results from the whole-plot factor (i.e., seed size) treatments in the small-plot trial appear in Table 2. There were no significant differences between seed sizes with regard to recoverable sucrose yield, root yield, or percent sucrose content. The relative lack of differences in plant populations (Table 1), coupled with these findings of no significant effects of seed size on yield parameters, further suggested that the main-level factor of seed size had no impact on the overall results of this trial.

Table 2. Whole-plot effect of seed size on yield parameters in a comparison of sugarbeet seed-flow lubricants in a small-plot field trial (Study I), Hillsboro, ND, 2017

Treatment/ form.	Recoverable sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose content (%)	Gross return (\$/ac)
Pro200	9,218 a	30.4 a	16.29 a	1,068
Mini	8,846 a	29.4 a	16.19 a	1,013
LSD (0.05)	NS	NS	NS	---

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

Results from the combined analysis of plant stand counts for the small-plot trial are presented in Table 3. There were no significant differences in plant stands among any of the hopper-box lubricant treatments or between any lubricant and the no-lubricant control at any of the three stand count dates. Slight numerical differences in stand counts among treatments were somewhat apparent at the first stand count date (i.e., 17 DAP); however, by the last count (45 DAP), the average plant population in the treatment with the lowest plant stands (the no-lubricant control) had only 1.8% fewer plants per 100 ft of row than the treatment with the highest stands (talc/graphite mixture).

Table 3. Effects of seed-flow lubricants on sugarbeet *plant population* in a small-plot field trial (Study I), Hillsboro, ND, 2017

Treatment/ form.	Rate ^a	Stand count ^b (plants / 100 row ft)		
		17 DAP ^c	31 DAP	45 DAP
Talc/graphite mixture (80:20)	10.4 ml	236 a	273 a	272 a
Talc	20.4 ml	225 a	273 a	271 a
Fluency Advanced	29.6 ml	208 a	271 a	271 a
Graphite	4 ml	207 a	269 a	270 a
Fluency Agent	29.6 ml	223 a	258 a	261 a
None	---	228 a	258 a	267 a
LSD (0.05)		NS	NS	NS

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aLubricants were applied to seed at rates recommended by respective manufacturers, and are expressed in volume per unit (100,000-ct) of sugarbeet seed.

^bSurviving plant stands were counted on June 1, 15, and 29, 2017, which were 17, 31, and 45 days after planting (DAP), respectively.

^cDAP = days after planting

Yield results from the small-plot experiment appear in Table 4. As observed in the stand count analyses, there were no statistical differences among lubricants or between any single lubricant and the no-lubricant control with regard to recoverable sucrose yield, root tonnage, or percent sucrose content. Accordingly, there were only negligible differences in gross economic return among the entries tested. The highest overall gross economic returns in this study were achieved with the following treatments: talc/graphite mixture, talc, and graphite.

Table 4. Effects of seed-flow lubricants on sugarbeet *yield parameters* in a small-plot field trial (Study I), Hillsboro, ND, 2017

Treatment/ form.	Rate ^a	Recoverable sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose content (%)	Gross return (\$/ac)
Graphite	4 ml	9,254 a	30.5 a	16.30 a	1,072
Talc	20.4 ml	9,235 a	30.1 a	16.40 a	1,087
Talc/graphite mixture (80:20)	10.4 ml	9,209 a	29.8 a	16.55 a	1,092
None	---	8,965 a	30.4 a	15.97 a	997
Fluency Advanced	29.6 ml	8,870 a	29.2 a	16.28 a	1028
Fluency Agent	29.6 ml	8,657 a	29.2 a	15.97 a	967
LSD (0.05)		NS	NS	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aLubricants were applied to seed at rates recommended by respective manufacturers, and are expressed in volume per unit (100,000-ct) of sugarbeet seed.

Study II (on-farm trial): Plant stand count data from the on-farm trial appear in Table 5. There were no significant differences in stands were observed among seed lubricants or between any lubricant and the no-lubricant control for any of the stand count dates.

Table 5. Effects of seed-flow lubricants on sugarbeet plant populations in an on-farm trial (Study II), Glyndon, MN, 2017

Treatment/ form.	Rate ^a	Stand Count ^b (plants / 100 row ft)		
		23 DAP ^c	31 DAP	52 DAP
Talc	20.4 ml	220 a	213 a	206 a
Talc/Graphite Mix (80:20)	10.4 ml	216 a	212 a	207 a
None	---	215 a	209 a	207 a
Fluency Advanced	29.6 ml	210 a	205 a	202 a
Graphite	4 ml	206 a	206 a	201 a
LSD (0.05)		NS	NS	NS

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aLubricants were applied to seed at rates recommended by respective manufacturers, and are expressed in volume per unit (100,000-ct) of sugarbeet seed.

^bSurviving plant stands were counted on May 24, and on June 1 and 22, which were 23, 31, and 52 days after planting (DAP), respectively.

^cDAP = days after planting

Yield results from hand-harvesting the plots in Study II are presented in Table 6. Excellent recoverable sucrose and root yields were recorded for all entries in this study, including the no-lubricant control. Yield trends closely corresponded to those from the plant stand assessments. There were no statistical differences with regard to recoverable sucrose, root yield, or percent sucrose content among the seed lubricants, or between any lubricant and the no-lubricant control. Another aspect of these results that corresponded with the plant stand data was that the top-yielding entry, with regard to both recoverable sucrose and root yield, was the talc/graphite combination lubricant.

Table 6. Hand-harvested yield in an on-farm trial of sugarbeet seed lubricants (Study II), Glyndon, MN, 2017

Treatment/ form.	Rate ^a	Recoverable sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose content (%)	Gross return (\$/ac)
Talc/Graphite Mix (80:20)	10.4 ml	15,012 a	39.2 a	19.96 a	2,251
Talc	20.4 ml	14,800 a	39.0 a	19.82 a	2,198
None	---	14,649 a	38.1 a	20.04 a	2,201
Graphite	4 ml	14,323 a	37.1 a	20.08 a	2,162
Fluency Advanced	29.6 ml	13,919 a	35.9 a	20.13 a	2,110
LSD (0.05)		NS	NS	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aLubricants were applied to seed at rates recommended by respective manufacturers, and are expressed in volume per unit (100,000-ct) of sugarbeet seed.

Yield results from machine-harvesting the plots in Study II appear in Table 7. Treatment performance patterns were very similar to those from hand-harvesting subsamples. The highest average recoverable sucrose yields occurred in plots planted using either Fluency Advanced or the talc/graphite mixture lubricant. Although no statistically significant, the no-lubricant control plots had the lowest recoverable sucrose and root yields. With regard to gross revenue, the top three entries included Fluency Advanced, the no-lubricant control, and the talc/graphite mixture.

Table 7. Machine-harvested yield in an on-farm trial of sugarbeet seed lubricants (Study II), Glyndon, MN, 2017

Treatment/ form.	Rate ^a	Recoverable sucrose yield (lb/ac)	Root yield (T/ac)	Sucrose content (%)	Gross return (\$/ac)
Fluency Advanced	29.6 ml	15,126 a	39.0 a	20.13 a	2,291
Talc/Graphite Mix (80:20)	10.4 ml	15,034 a	39.2 a	19.97 a	2,255
Talc	20.4 ml	14,958 a	39.4 a	19.83 a	2,226
Graphite	4 ml	14,932 a	38.7 a	20.07 a	2,252
None	---	14,546 a	37.9 a	20.04 a	2,290
LSD (0.05)		NS	NS	NS	

Means within a column sharing a letter are not significantly ($P = 0.05$) different from each other (Fisher's Protected LSD test).

^aLubricants were applied to seed at rates recommended by respective manufacturers, and are expressed in volume per unit (100,000-ct) of sugarbeet seed.

Given the highly consistent results between repeated plant population assessments and all yield parameters measured in the two experiments conducted for this project, it appears that the hopper-box seed flow lubricants tested do not impose a statistically significant positive or negative impact on sugarbeet seedling establishment, yield, or gross economic return. Trends across these two studies could suggest that using a talc/graphite mixture, such as the 80:20 product used in these trials, can occasionally optimize plant stands and yield; however, it cannot be concluded that using any seed lubricant during sugarbeet planting is absolutely necessary to achieve acceptable results with planters similar to those used in this experiment (i.e., John Deere MaxEmerge II or John Deere MaxEmerge Plus).

It should be noted that, while planting the Fluency Advanced treatment plots in the on-farm trial, the onboard seed monitor reported the following error message: "DISABLED DUE TO ERRATIC SPACING". Therefore, we make the following recommendations: 1) the exclusion of a seed flow lubricant for use in sugarbeet planting is not recommended at this time; 2) growers interested in or deciding to use Fluency Advanced should test this material on a small acreage with their own planters to determine its utility and safety; 3) use rates of the seed lubricants tested in these experiments should be made according to lubricant and/or planter manufacturer guidelines; and 4) growers that use planter makes and models other than those used in these experiments should review their owner's manual to determine if a seed lubricant is recommended for their planter, as well as carefully and extensively test the seed lubricant(s) they select to determine if they are safe for use with their planters.

Irrespective of the error message, the results of these experiments collectively and strongly suggest that reducing or eliminating talc from use in these planters does not appear to impose deleterious effects on sugarbeet stand establishment that translate to statistically significant yield loss. Therefore, growers could likely deploy insecticidal seed treatments in a safe and effective manner by using any hopper-box lubricant tested in this study (or by excluding a lubricant) without negatively impacting sugarbeet seedling establishment, yield, or gross economic return.

References Cited:

- Krupke, C. H., G. J. Hunt, B. D. Eitzer, G. Andino, and K. Given. 2012.** Multiple routes of pesticide exposure for honey bees living near agricultural fields. *PLoS ONE* 7(1): e29268.
- SAS Institute. 2008.** The SAS System for Windows. Version 9.2. SAS Institute Inc., 2002-2008. Cary, NC.

Entomology Appendix A.: Agronomic, Rainfall, and Plot Maintenance Information

Location: St. Thomas (Pembina County), ND – Wayne Lessard Farm – *Sugarbeet Root Maggot Trials*

Plot size: Six 35-ft long rows, 4 center rows treated

Design: Randomized complete block, 4 replications

Soil name: Glyndon silt loam

Soil test: Organic matter = 3.1% pH = 7.8

Soil texture: 25.2% sand 53.3% silt 21.6% clay

Previous crop: Potatoes (2016)

Soil preparation: Field cultivator (1x)

Planting depth: 1.25"

Herbicides applied: June 12 Roundup PowerMAX (32 fl oz/ac) + Veracity (3 qt/100 gal)
 June 29 Roundup PowerMAX (22 fl oz/ac) + Veracity (3 qt/100 gal) +
 Outlook (18 fl oz/ac)

Rainfall May 16 0.15"
 (after seedbed May 20 0.15"
 preparation): May 21 0.38"
 May 22 0.08"
 May 27 0.17"
 May 29 0.03"
 Total/May **0.96"**
 June 2 0.46"
 June 9 0.85"
 June 10 0.04"
 June 13 0.56"
 June 16 0.21"
 June 19 0.11"
 June 21 0.10"
 June 28 0.29"
 Total/June **2.62"**
 July 1 0.08"
 July 9 0.06"
 July 11 0.29"
 July 12 0.04"
 July 17 0.02"
 July 19 0.15"
 July 22 0.02"
 July 25 0.02"
 Total/July **0.68"**
 Total/August **1.55"**
 Total/September **2.99"**

Damage ratings: July 31 & August 1

Harvest date: October 2 & 3

Yield sample size: 2 center rows x 35 ft length (70 row-ft total)

Location: Manvel (Grand Forks County), ND – Stuart Ferry Farm – *Wireworm Management Trial*

Seed variety: Betaseed 80RR52

Plot size: Two 25-ft long rows

Design: Randomized complete block, 4 replicates

Soil name: Bearden silty clay loam

Soil test: Organic matter = 8.0% pH = 7.5

Soil texture: 9.7% sand 58.6 % silt 31.8% clay

Previous crop: Wheat (2016)

Soil preparation: Heavy harrow with vibra-shank and packer (1x immediately before planting)

Planting depth: 1.25"

Planting date: June 20

Herbicides applied: June 26 Roundup PowerMAX (32 fl oz/ac) + Stinger (2 fl oz/ac) + AMS (1 lb)

Fungicides applied: July 18 Penncozeb (2 lbs) + Inspire XT (7 fl oz/ac) + Surfactant (3.2 fl oz/ac)
 Aug. 10 Topsin (16 fl oz/ac) + Supertin (8 fl oz/ac)
 Sept. 7 Priaxor (6.7 fl oz/ac) + Surfactant (3.2 fl oz/ac)

Rainfall: June 21 0.06"
 (after seedbed June 24 0.02"
 preparation): June 28 3.69"
Total/June **3.77"**
 July 1 0.06"
 July 4 0.19"
 July 9 0.03"
 July 11 0.52"
 July 17 0.13"
 July 20 0.02"
 July 22 0.03"
Total/July **0.98"**
Total/August **0.69"**
Total/September **4.21"**

Stand counts: June 30; July 7, 13, and 27

Harvest date: October 9

Yield sample size: 2 rows x 25 ft length (50 row-ft total)

Location: Prosper (Cass County), ND – NDSU Experiment Farm – *Springtail Management Trial*

Seed variety: Betaseed 80RR52

Plot size: Two 25-ft long rows

Design: Randomized complete block, 4 replications

Soil name: Bearden-Lindaas silty clay loam

Soil test: Organic matter = 3.4% pH = 7.1

Soil texture: 27.0% sand 46.5% silt 26.6% clay

Previous crop: Wheat (2016)

Soil preparation: Field cultivator (2x)

Planting depth: 1.25"

Planting date: May 19

Herbicides applied:

June 1	Roundup PowerMAX (32 fl oz/ac) + Class Act (1% v/v) + Interlock (12 fl oz/ac)
June 19	Roundup PowerMAX (32 fl oz/ac) + Class Act (1% v/v) + Interlock (4 fl oz/ac)
July 10	Roundup PowerMAX (32 fl oz/ac) + Class Act (1% v/v) + Interlock (4 fl oz/ac)

Fungicides applied:

June 6	Quadris (14.3 fl oz ac)
June 19	Quadris (14.3 fl oz ac)
July 21	Inspire XT (5.3 fl oz/ac) + Topsin (7.6 fl oz/ac)
Aug. 2	Super Tin (6 fl oz/ac) + Manzate (1.2 qt/ac)

Rainfall: (after seedbed preparation):

May 20	0.38"
May 23	0.03"
May 27	0.07"
May 28	0.03"
May 29	0.03"
Total/May	0.54"
June 6	0.33"
June 7	0.09"
June 9	0.29"
June 11	0.17"
June 13	1.85"
June 17	0.45"
June 27	0.08"
June 28	0.20"
Total/June	3.46"
July 4	0.63"
July 6	0.23"
July 9	0.04"
July 18	0.47"
July 21	0.16"
July 22	0.15"
July 31	0.29"
Total/July	1.97"
Total/August	2.07"

Total/September 0.22"

Stand counts: June 1, 15, and 29; July 7
Harvest date: September 18
Yield sample size: 2 rows x 25 ft length (50 row-ft total)

Location: Hillsboro (Traill County), ND – Glen Hultin Farm – *Small-plot Seed Lubricants Test*

Seed variety: Betaseed 83CN Mini Pellet & 83CN Pro200 Pellet

Plot size: Six 35-ft long rows, 4 center rows treated

Design: Randomized complete block, 4 replications

Soil name: Bearden–Perella silty clay loam

Soil test: Organic matter = 4.5% pH = 7.8

Soil texture: 9.1% sand 54.8% silt 36.2% clay

Previous crop: Wheat (2016)

Soil preparation: Harrow packer (1x)

Planting depth: 1.25"

Planting date: May 15

Herbicides applied: June 5 Roundup PowerMAX (32 fl oz/ac) + Veracity (3 qt/100 gal)
 June 29 Roundup PowerMAX (22 fl oz/ac) + Veracity (3 qt/100 gal)

Fungicides applied: Aug. 10 Topsin (7.5 fl oz/ac) + Inspire XT (7 fl oz/ac)

Rainfall May 20 0.30"
 (after seedbed May 21 0.06"
 preparation): May 28 0.15"
 Total/May **0.51"**
 June 2 0.03"
 June 7 0.14"
 June 11 0.18"
 June 13 0.84"
 June 17 0.35"
 June 19 0.04"
 June 28 1.30"
 Total/June **2.88"**
 July 4 0.22"
 July 5 0.10"
 July 6 0.19"
 July 11 0.34"
 July 18 0.10"
 July 21 0.12"
 July 22 0.28"
 July 31 0.23"
 Total/July **1.58"**
 Total/August **0.61"**
 Total/September **0.66"**

Stand counts: June 1, 15, and 29

Harvest date: September 19

Yield sample size: 2 center rows x 35 ft length (70 row-ft total)

Location: Glyndon (Clay County), MN – David Watt Farm – *On-farm Seed Lubricants Test*

Seed variety: Betaseed 8572

Plot size: Twelve 600-ft long rows

Design: Randomized complete block, 4 replicates

Soil name: Glyndon loam

Soil test: Organic matter = 4.0% pH = 7.8

Soil texture: 11.1% sand 62.1% silt 26.9% clay

Previous crop: Wheat (2016)

Soil preparation: Field cultivator with packer (1x)

Planting depth: 1.25"

Planting date: May 1

Herbicide applied: May 19 Roundup PowerMAX (28 fl oz/ac) + Brawl (1 pt/ac)
 June 12 Roundup PowerMAX (28 fl oz/ac) + Brawl (1 pt/ac)

Fungicides applied: July 7 Inspire (7 fl oz/ac) + Manzate (1 qt/ac)
 July 29 Tin (8 fl oz/ac) + Topsin (10 fl oz/ac)
 Aug. 18 Proline (7 fl oz/ac)

Rainfall
 (after seedbed
 preparation):

May 5	0.07"
May 16	0.28"
May 20	0.50"
May 28	0.19"
Total/May	1.04"
June 7	0.04"
June 11	0.19"
June 13	1.36"
June 17	0.11"
June 27	0.19"
June 28	0.30"
Total/June	2.26"
July 4	0.20"
July 6	0.03"
July 9	0.05"
July 18	0.22"
July 22	0.19"
July 31	0.19"
Total/July	0.89"
Total/August	2.29"
Total/September	2.75"

Stand counts: May 24; June 1 and 22

Harvest date: Hand harvest – October 12
 Machine harvest – October 16

Yield sample size: Hand harvest – four 0.00025-ac samples per plot
 Machine harvest – twelve 50-ft long rows per plot

Entomology Appendix B. 0 to 9 Scale for Rating Sugarbeet Root Maggot Feeding Injury

Treatment performance in preventing sugarbeet root maggot feeding injury was quantified for all root maggot control trials by rating beets on the 0 to 9 root injury rating scale of Campbell et al. (2000). Criteria for respective points on the scale are as follows:

- 0 = no scars
- 1 = 1 to 4 small (pin head size) scars
- 2 = 5 to 10 small scars
- 3 = 3 large scars or scattered small scars
- 4 = few large scars and /of numerous small scars
- 5 = several large scars and/or heavy feeding on laterals
- 6 = up to 1/4 root scarred
- 7 = 1/4 to 1/2 of root blackened by scars
- 8 = 1/2 to 3/4 root blackened by scars
- 9 = more than 3/4 of root area blackened

Reference Cited:

Campbell, L. G., J. D. Eide, L. J. Smith, and G. A. Smith. 2000. Control of the sugarbeet root maggot with the fungus *Metarhizium anisopliae*. J. Sugar Beet Res. 37: 57–69.

PLANT PATHOLOGY

NOTES

TURNING POINT SURVEY OF FUNGICIDE USE IN SUGARBEET IN MINNESOTA AND EASTERN NORTH DAKOTA IN 2016

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The second annual fungicide practices live polling questionnaire was conducted using Turning Point Technology at the 2017 Winter Sugarbeet Growers' Seminars. Responses are based on production practices from the 2016 growing season. The survey focuses on responses from growers in attendance at the Fargo, Grafton, Wahpeton, ND and Willmar, MN Grower Seminars. Respondents from each seminar indicated the county in which the majority of their sugarbeets were produced (Tables 1- 4). Survey results represent approximately 158,272 acres reported by 235 participants (Table 5) compared to 183,350 acres represented in 2016. The average sugarbeet acreage per respondent grown in 2016 was calculated from Table 5 at 673 acres, compared to 674 acres in 2015.

Survey participants were asked a series of questions regarding their fungicide practices used on sugarbeet in 2016. Twenty-eight percent of respondents reported that they used five sprays to control *Cercospora* Leaf Spot (Table 6) while 22% said they used three sprays, 17% used four sprays, 11% used seven sprays, 10% used six sprays, 7% used two sprays, 3% used one spray and 1% both used no sprays and more than seven sprays. Thirty-five percent of respondents both reported a fair amount of effectiveness and a poor amount of effectiveness (Table 7). Twenty-seven percent said they had a good amount of control from CLS spray, 3% had an excellent amount of effectiveness and 1% said they did not use any fungicide for control of CLS. Respondents were then asked when they experienced failure of fungicides to control CLS (Table 8). Twenty-seven percent of respondents reported failure between August 1 and August 15, 17% said field failure occurred between August 16 and August 31, 11% said that failure occurred between September 1 and September 15, 10% said it occurred before July 31, 5% said CLS field failure happened between September 16 and September 30 and 4% said after September 30. Meanwhile, 26% of respondents said they did not experience field failure (Table 9). Participants in the survey were then asked what fungicide was sprayed right before the field experienced failure. Thirty-three percentage of respondents said that Headline was sprayed right before failure, 20% reported Tin, 15% said Priaxor, 9% reported some kind of fungicide mixture, 5% said Topsin while the same percentage also reported Proline and Gem was sprayed right before field failure due to CLS. Four percent said Minerva or Eminent and 3% said the Inspire XT was sprayed right before failure.

Respondents were then asked about soil-borne diseases. Forty percent said their fields were affected by both *Rhizoctonia* and *Aphanomyces*, 36% said just *Rhizoctonia*, 15% had neither disease in their fields and 10% had only *Aphanomyces* (Table 10). Eighty seven percent of respondents used a *Rhizoctonia* resistant variety in 2016 (Table 11) while 88% used an *Aphanomyces* resistant variety (Table 17).

Participants were asked what methods were used to control *Rhizoctonia* and 55% said they used a seed treatment only, 35% used a seed treatment and a POST fungicide, 4% used a seed treatment plus an in-furrow fungicide while 4% also said they used a seed treatment, in-furrow fungicide and a POST fungicide. Two percent only used a POST fungicide (Table 12). Eighty-five percent of respondents used a Kabina seed treatment while 11% used a Rizolex + Metlock + Kabina mixture, 3% used a different seed treatment and 15 reported not using a seed treatment to control *Rhizoctonia* (Table 13). Eighty-seven percent of respondents did not use an in-furrow fungicide but 8% of respondents used Quadris in-furrow, 4% used Headline in-furrow to control *Rhizoctonia* and 1% used a different fungicide (Table 14).

Respondents were asked what POST fungicides were used to control *Rhizoctonia* and 45% did not use a POST fungicide to control *Rhizoctonia*. Of the remaining 55%, 44% used Quadris, 5% used Priaxor, 3% used Proline, 1% used Headline while 2% used a different fungicide (Table 15). Participants were then asked to grade the effectiveness of the POST fungicides that were used. Forty-one percent were unsure of the effectiveness, 32% said

they performed good, 17% reported fair results, 6% said they performed poorly and 4% said they were excellent (Table 16).

Participants were also asked about use of waste lime to control *Aphanomyces*. 56% of participants did not use waste lime in their fields while 23% used 5 tons/acre or less. Nineteen percent used between 6 and 10 tons/acre while 2% used more than 10 tons/acre (Table 18). Respondents were also asked about their soil pH. Thirty-six percent said it was between 8.0 and 8.5, 29% said that it was between 7.5 and 8.0, 22% said it was between 7.0 and 7.5, 6% said between 6.5 and 7.0, 5% said between 6.0 and 6.5 and 1% said between 8.5 and 9.0 (Table 19). As a follow-up question, growers were asked whether or not they were concerned about using waste lime on soils above 8.0 pH. Seventy-four percent said no while the remaining 26% said they were concerned (Table 20). Finally, the growers were asked how effective their waste lime was. Fifty percent of respondents did not apply lime, 19% said they had good results, 15% were unsure, 9% reported excellent results, 5% said fair and 1% said poor (Table 21).

Table 1. 2017 Fargo Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Barnes	3	9
Cass	7	21
Clay	11	32
Norman ¹	8	24
Richland	1	3
Trail	3	9
Wilkin ²	1	3
Total	34	100

¹Includes Mahnomon County

²Includes Otter Tail County

Table 2. 2017 Grafton Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Grand Forks	1	2
Kittson	4	7
Marshall	5	9
Pembina	19	35
Polk	1	2
Walsh	23	43
Other	1	2
Total	54	100

Table 3. 2017 Wahpeton Grower Seminar – Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Cass	2	4
Clay	3	7
Grant	5	11
Otter Tail	1	2
Richland	7	16
Stevens	1	2
Traverse	5	11
Wilkin	21	47
Total	45	100

Table 4. 2017 Willmar Grower Seminar - Number of survey respondents by county growing sugarbeet in 2016.

County	Number of Responses	Percent of Responses
Chippewa	36	33
Kandiyohi	17	16
Pope	0	0
Redwood	5	5
Renville	31	28
Stearns	3	3
Stevens	1	1
Swift	9	8
Other	7	6
Total	109	100

Table 5. Total sugarbeet acreage operated by respondents in 2016.

Location	Responses	Acres of sugarbeet									
		<99	100-199	200-299	300-399	400-599	600-799	800-999	1000-1499	1500-1999	2000+
		-----% of responses-----									
Grafton	54	6	15	11	9	17	9	11	9	2	9
Fargo	33	3	0	15	18	18	6	9	12	6	12
Wahpeton	42	2	7	2	10	33	17	12	10	5	2
Willmar	107	7	15	15	6	22	10	3	14	2	7
Total	235	6	11	12	9	22	11	7	12	3	7

Table 6. How many fungicide application did you make to control CLS in 2016?

Location	Respondents	Number of applications									
		0	1	2	3	4	5	6	7	>7	
		-----% of respondents-----									
Fargo	37	-	-	16	35	27	22	-	-	-	
Grafton	50	2	16	22	56	4	-	-	-		
Wahpeton	46	-	-	-	20	30	48	2	-		
Willmar	105	1	-	-	3	14	35	22	24	1	
Total	238	1	3	7	22	17	28	10	11	1	

Table 7. How effective were your fungicide applications on CLS in 2016?

Location	Respondents	Effectiveness of CLS sprays					
		Excellent	Good	Fair	Poor	Unsure	No applications
		-----% of respondents-----					
Fargo	36	3	47	39	11	-	-
Grafton	50	6	58	34	2	-	-
Wahpeton	45	-	11	29	60	-	-
Willmar	107	2	12	36	48	-	2
Total	238	3	27	35	35	-	1

Table 8. When did you experience failure of fungicides to control CLS?

Location	Respondents	Date of fungicide failure						
		No failure	July 31	August 15	August 31	September 15	September 30	After September 30
		-----% of respondents-----						
Fargo	32	25	9	13	31	9	9	3
Grafton	49	55	-	12	8	14	6	4
Wahpeton	44	2	18	55	18	5	2	-
Willmar	15	7	20	27	13	20	-	13
Total	140	26	10	27	17	11	5	4

Table 9. If you had failure with fungicides for CLS control, which fungicide did you apply prior to observing field failure?

Location	Respondents	Fungicide failure									
		Minerva, Eminent	Inspire XT	Proline	Headline	Priaxor	Gem	Tin Topsin	EBDC	Mixtures	
		-----% of respondents-----									
Fargo	21	-	10	10	38	10	-	19	-	-	14
Grafton	34	-	3	3	6	26	-	26	12	-	24
Wahpeton	40	8	-	3	78	8	-	-	3	-	3
Willmar	88	6	2	7	22	16	10	27	6	-	5
Total	183	4	3	5	33	15	5	20	5	-	9

Table 10. What soil-borne diseases affected your sugarbeet production in 2016?

Location	Respondents	Root disease			
		Rhizoctonia	Aphanomyces	Both	Neither
		-----% of respondents-----			
Fargo	34	35	15	35	15
Grafton	49	27	14	57	2
Wahpeton	43	47	2	21	30
Total	126	36	10	40	15

Table 11. Did you use a *Rhizoctonia solani* resistant variety in 2016?

Location	Respondents	Variety type	
		Yes	No
		-----% respondents-----	
Fargo	35	97	3
Grafton	47	94	6
Wahpeton	40	90	10
Willmar	98	80	20
Total	220	87	13

Table 12. What methods were used to control *Rhizoctonia solani* in 2016?

Location	Respondents	Treatment methods					
		Seed treatment only	In-Furrow only	Postemergence only	Seed treatment + In-Furrow	Seed treatment + Postemergence	All three treatments used
		-----% of respondents-----					
Fargo	34	47	-	-	-	53	-
Grafton	48	42	-	-	4	54	-
Wahpeton	42	86	-	-	2	10	2
Willmar	99	52	-	4	6	30	8
Total	223	55	-	2	4	35	4

Table 13. Which seed treatment did you use to control *Rhizoctonia solani* in 2016?

Location	Respondents	Seed treatment			
		Kabina	Rizolex + Metlock + Kabina	Other	None
		-----% of respondents-----			
Fargo	35	86	14	-	-
Grafton	49	82	10	4	4
Wahpeton	39	87	10	3	-
Willmar	101	85	10	4	1
Total	224	85	11	3	1

Table 14. Which fungicide did you apply in-furrow to control *R. solani* in 2016?

Location	Respondents	In-furrow fungicide use			
		Headline	Quadris	Other	None
		-----% of respondents-----			
Fargo	32	6	9	-	84
Grafton	49	2	8	-	90
Wahpeton	41	-	10	-	90
Willmar	104	6	7	3	85
Total	226	4	8	1	87

Table 15. Which POST fungicide did you use to control *R. solani* in 2016?

Location	Respondents	POST fungicide					None
		Headline	Quadris	Proline	Priaxor	Other	
		-----% of respondents-----					
Fargo	34	-	59	-	3	-	38
Grafton	51	4	63	2	14	-	18
Wahpeton	40	-	10	-	-	5	85
Willmar	102	1	44	5	3	2	45
Total	227	1	44	3	5	2	45

Table 16. How effective were your POST fungicides at controlling *Rhizoctonia solani* in 2016?

Location	Respondents	Effectiveness of fungicides				
		Excellent	Good	Fair	Poor	Unsure
		-----% of respondents-----				
Fargo	30	3	47	13	-	37
Grafton	46	2	41	35	7	15
Wahpeton	33	-	9	3	3	85
Willmar	89	6	30	16	8	40
Total	198	4	32	17	6	41

Table 17. Did you use an *Aphanomyces* resistant variety in 2016?

Location	Respondents	Variety type	
		Yes	No
		-----% respondents-----	
Fargo	25	96	4
Grafton	47	87	13
Wahpeton	38	84	16
Total	110	88	12

Table 18. What rate of precipitated calcium carbonate (waste lime) did you use?

Location	Respondents	Lime use rate			
		None	>5 T/A	6-10 T/A	10+ T/A
		-----% of respondents-----			
Fargo	33	61	3	27	9
Grafton	52	77	-	21	2
Wahpeton	41	39	15	44	2
Willmar	101	51	46	4	-
Total	227	56	23	19	2

Table 19. What is your soil pH?

Location	Respondents	Soil pH					
		6.0-6.5	6.5-7.0	7.0-7.5	7.5-8.0	8.0-8.5	8.5-9.0
		-----% of respondents-----					
Fargo	32	-	6	13	31	50	-
Grafton	45	9	7	29	27	27	2
Total	77	5	6	22	29	36	1

Table 20. Are you concerned about using waste lime on pH soils above 8.0?

Location	Respondents	Safety concerns	
		Yes	No
		-----% respondents-----	
Fargo	32	28	72
Grafton	48	25	75
Total	80	26	74

Table 21. How effective was waste lime at controlling Aphanomyces?

Location	Respondents	Waste lime effectiveness					
		Excellent	Good	Fair	Poor	Unsure	No Lime
		-----% of respondents-----					
Fargo	36	8	19	-	-	22	50
Grafton	49	6	10	8	-	6	69
Wahpeton	42	26	19	5	-	17	33
Willmar	100	3	24	5	3	16	49
Total	227	9	19	5	1	15	50

INTEGRATED MANAGEMENT OF RHIZOCTONIA ON SUGARBEET WITH RESISTANT VARIETIES, AT-PLANTING TREATMENTS, AND POSTEMERGENCE FUNGICIDES

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Rhizoctonia damping-off and crown and root rot (RCRR) caused by *Rhizoctonia solani* AG 2-2 have been the most common root diseases on sugarbeet in Minnesota and North Dakota for several years (1, 3-4,6). Disease can occur throughout the growing season and reduces plant stand, root yield, and quality. Warm and wet soil conditions favor infection. Disease management options include rotating with non-host crops (cereals), planting partially resistant varieties, planting early when soil temperatures are cool, improving soil drainage, and applying fungicides as seed treatments, in-furrow (IF), or postemergence. An integrated management strategy should take advantage of multiple control options to reduce Rhizoctonia crown and root rot.

OBJECTIVES

A field trial was established to evaluate an integrated management strategy consisting of a resistant (R) and a moderately susceptible (MS) variety with new available seed treatments alone and in combination with two postemergence azoxystrobin application timings for 1) control of early-season damping-off and RCRR and 2) effect on yield and quality of sugarbeet.

MATERIALS AND METHODS

The trial was established at three locations, one at the University of Minnesota, Northwest Research and Outreach Center, Crookston, one at Wahpeton (MDFC), ND and one at Renville (SMBSC), MN. All locations were fertilized for optimal yield and quality. At each location, a combination of a R and MS variety treated with penthiopyrad (Kabina ST), flupyroxad (Systiva), sedaxane (Vibrance), or untreated was planted in four replicate plots. Plots were set up in a split-split plot design at all 3 locations. Main plots were varieties, the first split was seed treatments, and the last split was postemergence azoxystrobin timings. Seed treatments and rates are summarized in Table 1 and were applied by Germains Seed Technology, Fargo, ND. Each variety by seed treatment combination was planted in triplicate, so that at the 4- or 8-leaf stage, one plot of each variety by seed treatment combination received a postemergence 7-inch band application of azoxystrobin (14.3 fl oz product A⁻¹) while one was left as a stand-alone treatment. Controls for each variety included no seed treatment at planting with each postemergence azoxystrobin timing and without postemergence azoxystrobin. Two-year average Rhizoctonia ratings in American Crystal Sugar Company tests for the R and MS varieties were 4.0 and 4.7, respectively (7).

NWROC site. Prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley broadcast at 35 kg ha⁻¹ and incorporated with a Rau seedbed finisher. The trial was sown in six-row plots (22-inch row spacing, 30-ft rows) on May 10 at 4.5-inch seed spacing. Counter 20G (8 lb A⁻¹) was applied at planting for control of sugarbeet root maggot. Glyphosate (4.5 lb product ae/gallon) was applied on May 15 (22 oz A⁻¹), June 1, 7, and 12 (28 oz A⁻¹), and July 5 (32 oz A⁻¹) for control of weeds. The June 1 application also included S-metolachlor (0.94 lb a.i. A⁻¹). Postemergence azoxystrobin timings were applied in a 7-inch band in 10 gallon/A using 4002 nozzles and 34 psi on June 12 (4-leaf stage, ~4.5 weeks after planting) or June 20 (8-leaf stage, 6 weeks after planting). Cercospora leaf spot was controlled by Supertin + Topsin M (6 + 10 oz product in 19 gallons of water A⁻¹) applied with 8002 flat fan nozzles at 100 psi on July 24 and Inspire (7 oz product in 19 gallons of water A⁻¹) on August 8.

MDFC site. Prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley (35 kg ha⁻¹). The trial was sown in six-row plots (22-inch row spacing, 30-ft rows) on May 26 at 4.5-inch seed spacing. Glyphosate (4.5 lb product ae/gallon) tank-mixed with N-tense (9.6 oz A⁻¹) was applied on May 31. This weed control application was repeated again on June 20 and July 03 (plus Outlook 12 oz A⁻¹). Postemergence azoxystrobin was applied in a 7-inch band on June 16 (4-leaf stage, 3 weeks after planting) or June 29 (8-leaf stage, 5 weeks after planting). Cercospora

leafspot was controlled by separate applications of TPTH+Topsin (8 & 10 oz A⁻¹, respectively) on July 13, Inspire XT+Badge SC (7 & 32 oz A⁻¹, respectively) on July 25, TPTH + Manzate (8 & 38.4 oz A⁻¹, respectively) on August 04, Minerva Duo (16 oz A⁻¹) on Aug 15 and TPTH+ Badge SC (8 & 32 oz A⁻¹, respectively) as last application on Aug 29. All fungicides for CLS control were applied utilizing a 3pt-mounted sprayer dispersing the products in broadcast pattern at a water volume of 15 GPA with TeeJet 8002 flat fan nozzles at 80 psi.

Table 1. Application type, product names, active ingredients, and rates of fungicides used at planting in a field trial for control of *Rhizoctonia solani* AG 2-2 on sugarbeet. Each at-plant treatment was used in combination with a *Rhizoctonia* resistant (2-year average rating = 4.0) and moderately susceptible (2-year average rating = 4.8) variety, and all treatment combinations in triplicate, with one set receiving a postemergence 7-inch band application of azoxystrobin (14.3 fl oz A⁻¹) at 4- or 8-leaf stage. Standard rates of Apron + Thiram and 45 g/unit Tachigaren were on all seed.

Application	Product	Active ingredient	Rate
None	-	-	-
Seed	Kabina ST	Penthiopyrad	14 g a.i./unit seed
Seed	Systiva	Fluxapyroxad	5 g a.i./unit seed
Seed	Vibrance	Sedaxane	1.5 g a.i./unit seed

Table 2. Monthly precipitation in inches at three sites during 2017 crop season based on weather stations.

Month	Precipitation in inches		
	NWROC	MDFC	SMBSC
May	0.94	1.33	2.42
June	3.41	3.64	1.18
July	1.42	2.62	1.97
August	0.77	5.00	6.92
September	4.01	4.31	1.34
Total	10.55	16.91	13.83

SMBSC site. Prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley (35 kg ha⁻¹). The trial was sown in six-row plots (22-inch row spacing, 30-ft rows) on May 12 at 4.6-inch seed spacing. Weeds were controlled by application of Powermax (28 oz A⁻¹) + Dual magnum (16 oz A⁻¹) on June 5 and Powermax (22 oz A⁻¹) on July 06. Postemergence azoxystrobin timings were applied on June 09 (4-leaf, ~4 weeks after planting), or June 20 (8-leaf, ~5 weeks after planting) as 7 inch bands using 80002E nozzles at 40 psi. Fungicides were applied for controlling *Cercospora* leaf spot on July 10 (TPTH + Topsin, 8 & 20 oz A⁻¹, respectively), July 21 (Inspire XT + Badge SC, 7 & 32 oz A⁻¹, respectively), July 31 (TPTH + Dithane F-45, 8 & 51.2 oz A⁻¹, respectively), Aug 12 (Minerva + Dithane F-45, 13 & 51.2 oz A⁻¹, respectively), Aug 23 (TPTH + Badge SC, 8 & 32 oz A⁻¹, respectively) and Sept 06 (Proline + Dithane F-45, 5.7 & 51.2 oz A⁻¹, respectively). All fungicides for CLS control were applied in a water volume of 19.3 GPA with 11002 nozzles at 70 psi.

At NWROC stand counts were done beginning 2 weeks after planting through 8 weeks after planting. At MDFC stand counts were done 2 through 6 weeks after planting. At SMBSC stand counts were done 3, 5, and 8 weeks after planting. The trial was harvested on September 20 at the NWROC, Sept 19 at Renville and October 09 at Wahpeton. Data were collected for number of harvested roots (NWROC only), yield, and quality. Twenty roots per plot also were arbitrarily selected and rated for severity of RCRR using a 0 to 7 scale (0 = healthy root, 7 = root completely rotted and foliage dead). Disease incidence was reported as the percent of rated roots with a root rot rating of > 2.

Data were subjected to analysis of variance using SAS Proc GLM (SAS Institute, Cary, NC) for main effects of variety, at-plant treatment, postemergence azoxystrobin application, and all possible interactions. Means were separated by Fisher's Protected Least Significant Difference ($P = 0.05$).

RESULTS AND DISCUSSION

NWROC site: 2017 growing season was drier and cooler at the NWROC during the period of May - August. Rainfall at the NWROC was just 0.94 inch during the month of May compared to a 30-year average of 3.04 inches for May.

Average four-inch bare soil temperatures at the NWROC were 52.4 °F and 61.9 °F for the months of May and June, respectively. Average four-inch soil temperature did not cross 65 °F until July 04. There were no significant ($P > 0.05$) two way or three way interactions for stand data. For harvest data there was a significant seed treatment x postemergence application interaction ($P < 0.05$) for root rot rating and incidence (Tab. 3). Resistant and moderately susceptible variety had similar stands from 2 to 8 weeks after planting (WAP). At-planting (seed) treatments and untreated control had similar stands at 2 WAP and by 3 WAP all the seed treatments had higher stands compared to untreated control. At 5 WAP, Vibrance had highest stands, Kabina and Systiva had intermediate, and untreated control had lowest stands (Fig. 1). Total rainfall for the months of May - August was 6.54 inches in 2017 compared to a 30-year average of 12.88 for the same time period. Soil moisture remained low throughout the growing season, resulting in low *Rhizoctonia* disease pressure in this trial. As a result, there were no significant differences among treatments for *Rhizoctonia* root rot or yield and quality parameters between varieties and also untreated control and seed treatments. There were no significant differences between two varieties for harvest data (Tab. 3). Yield, percent sucrose, recoverable sucrose A⁻¹ (RSA), percent sucrose and recoverable sucrose T⁻¹ (RST) were not significantly different for the seed treatments and untreated control (Tab. 3). Yield, percent sucrose, RSA and RST were not significantly different between Quadris (4- or 8-leaf) and no Quadris application. Some rainfall in September created slight disease pressure in the plots leading to minor differences in disease severity between no Quadris and 4-8 leaf Quadris applications. Root rot severity and percent incidence (percent of roots with a disease rating of > 2.0) was slightly higher in the no Quadris treatments for control and all seed treatments, intermediate in 4-leaf Quadris treatments for control and all seed treatments, and lowest in 8-leaf Quadris for control, Kabina, and Vibrance treatments (Figs. 2A and 2B). Similar benefit from postemergence Quadris application was also evident in 2016 (5).

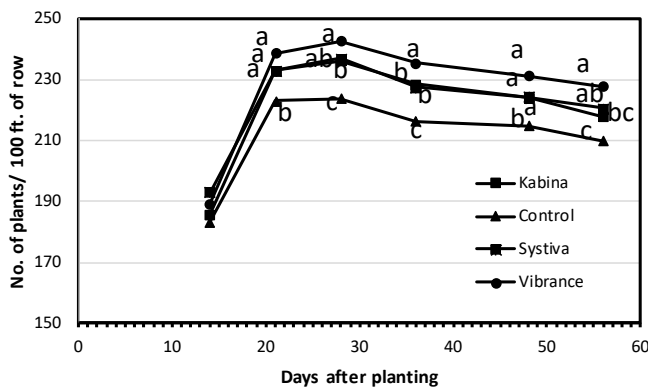


Fig. 1. NWROC site: Emergence and stand establishment for fungicide treatments on seed or untreated control. For each stand count date, values sharing the same letter are not significantly different ($P = 0.05$); NS = not significantly different. Data shown represents mean of 24 plots averaged across varieties and postemergence treatments.

Table 3. NWROC site: Main effects of variety, at-planting (seed), and postemergence fungicide treatments on *Rhizoctonia* crown and root rot and sugarbeet yield and quality in a field trial sown May 10, 2017.

Main effect (Apron + Maxim on all seed)	No. harv. roots/100 ft ^T	RCRR (0-7) ^{TU}	RCRR % incidence ^{TV}	Yield ton A ^{-1T}	Sucrose ^T		
					%	lb ton ⁻¹	lb A ⁻¹
Variety ^W Resistant	184	0.6	4.6	19.3	18.3	345	6630

Moderately Susceptible	196	0.7	11.4	20.8	17.8	333	6890
ANOVA p-value	0.1026	0.5862	0.3881	0.4668	0.2156	0.1528	0.642
LSD ($P = 0.05$)	NS	NS	NS	NS	NS	NS	NS
At-planting treatments ^x							
Untreated control	185	0.8	9	20.6	18.0	339	6969
Kabina ST @ 14 g a.i./unit	189	0.7	10	20.0	18.0	338	6724
Systiva @ 5 g a.i./unit	190	0.7	9	19.7	18.0	338	6665
Vibrance @ 1.5 g a.i./unit	196	0.4	4	19.7	18.1	340	6681
ANOVA p-value	0.3296	0.2454	0.2666	0.5700	0.8385	0.9038	0.4313
LSD ($P = 0.05$)	NS	NS	NS	NS	NS	NS	NS
Postemergence fungicide ^y							
None	185	1.0 a	14 a	19.5	18.0	338	6581
4-leaf Quadris @ 14.3 fl. oz./A	192	0.5 b	7 b	20.2	18.2	341	6874
8-leaf Quadris @ 14.3 fl. oz./A	192	0.4 b	4 b	20.3	18.0	338	6825
ANOVA p-value	0.0539	<0.0001	<0.0001	0.2045	0.2846	0.3927	0.1113
LSD ($P = 0.05$)	NS	0.19	3.2	NS	NS	NS	NS
Vty x Seed	NS	NS	NS	NS	NS	NS	NS
Vty x Post	NS	NS	NS	NS	NS	NS	NS
Seed x Post	NS	0.0206	0.0086	NS	NS	NS	NS
Vty x Seed x Post	NS	NS	NS	NS	NS	NS	NS

^T Numbers followed by the same letter are not significantly different; LSD = Least Significant Difference, $P = 0.05$; NS = not significantly different

^U RCRR = Rhizoctonia crown and root rot; 0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead

^V RCRR = Rhizoctonia crown and root rot; percent of roots with rating greater than two

^W Values represent mean of 48 plots (4 replicate plots across 4 at-planting treatments and 3 postemergence treatments)

^X Values represent mean of 24 plots (4 replicate plots across 2 varieties and 3 postemergence treatments)

^Y Values represent mean of 32 plots (4 replicate plots across 2 varieties and 4 at-planting treatments)

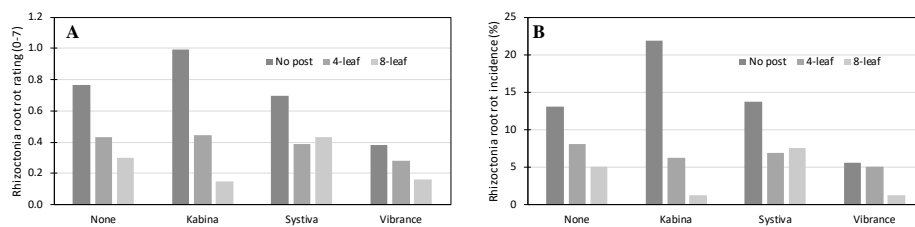


Fig. 2. NWROC site: Effect of seed and postemergence treatments on **A**) Rhizoctonia root rot severity (0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead) and **B**) Rhizoctonia root rot incidence (percent of roots with rating greater than two).

MDFC site: This site received below normal rainfall during May – July and above normal rainfall during August-September. Average 4-inch bare soil temperatures for May (59 °F) and June (68 °F) were lower compared to 2016 (64 °F and 74 °F for May and June, respectively). Average four-inch soil temperature was over 65 °F on June 02, reached ~ 70 °F for a week followed by a ~65 °F from June 17 until July 03. Low early season soil moisture coupled with lower soil temperatures did not create heavy disease pressure at this site. There were significant ($P < 0.05$) variety x seed treatment interactions and variety x seed treatment x postemergence three way interactions for percent sugar, purity, and RST; variety x postemergence interactions for root rot rating (Tab. 4). Both varieties had similar stands until 6 WAP and had similar yield, percent sucrose, RST, and RSA (Tab. 4). There were no significant differences for stands between seed treatments and untreated control until 6 WAP. Yield was not significantly different between untreated control and seed treatments. Some rainfall in August and September created slight disease pressure in the plots leading to minor differences in disease severity and some harvest parameters between no Quadris and 4-8

leaf Quadris applications. Yield was not significantly different between no Quadris and 4- or 8-leaf application. RSA was higher in 8-leaf Quadris application compared to 4-leaf or no Quadris application. Root rot incidence was lower in 4- or 8-leaf application compared to no Quadris. For resistant variety, percent sucrose and RST were highest for untreated control and lowest for Systiva, whereas for moderately susceptible variety Systiva had highest percent sucrose and RST with lowest for Kabina (Figs. 3A and 3B). For resistant variety root rot severity was lowest for 4-leaf Quadris application, intermediate for 8-leaf and highest for no Quadris application (Fig. 3C). For moderately susceptible variety root rot severity was lower for 4- or 8-leaf Quadris application compared to no Quadris application (Fig. 3C). Similar benefit from postemergence Quadris application was also evident in 2016 (5).

Table 4. MDFC site: Main effects of variety, at-planting (seed), and postemergence fungicide treatments on Rhizoctonia crown and root rot and sugarbeet yield and quality in a field trial sown May 26, 2017.

Main effect (Apron + Maxim on all seed)	RCRR (0-7) ^{TU}	RCRR % incidence ^{TV}	Yield ton A ^{-1T}	Sucrose ^T		
				%	lb ton ⁻¹	lb A ⁻¹
Variety ^W						
Resistant	0.3	6.1	27.0	16.2	266	7195
Moderately Susceptible	0.6	11.3	27.0	15.3	248	6698
ANOVA p-value	0.1203	0.1754	0.9775	0.0587	0.0756	0.2039
LSD (<i>P</i> = 0.05)	NS	NS	NS	NS	NS	NS
At-planting treatments ^X						
Untreated control	0.5	10.8	27.2	15.9	262	7116
Kabina ST @ 14 g a.i./unit	0.4	7.9	26.5	15.5	252	6690
Systiva @ 5 g a.i./unit	0.4	7.9	27.3	15.8	257	7016
Vibrance @ 1.5 g a.i./unit	0.4	8.1	26.9	15.9	259	6963
ANOVA p-value	0.6365	0.5959	0.6152	0.4018	0.3529	0.2540
LSD (<i>P</i> = 0.05)	NS	NS	NS	NS	NS	NS
Postemergence fungicide ^Y						
None	0.8 a	15.2 a	26.4	15.6 b	254 b	6720 b
4-leaf Quadris @ 14.3 fl. oz./A	0.3 b	5.0 b	27.1	15.7 b	255 b	6916 b
8-leaf Quadris @ 14.3 fl. oz./A	0.3 b	5.9 b	27.4	16.0 a	263 a	7203 a
ANOVA p-value	<0.0001	<0.0001	0.0612	0.0008	0.0002	0.0008
LSD (<i>P</i> = 0.05)	0.18	4.0	NS	0.18	4.28	240
Vty x Seed	NS	NS	NS	0.0491	0.0485	NS
Vty x Post	0.0454	NS	NS	NS	NS	NS
Seed x Post	NS	NS	NS	NS	NS	NS
Vty x Seed x Post	NS	NS	NS	0.0209	0.0067	NS

^T Numbers followed by the same letter are not significantly different; LSD = Least Significant Difference, *P* = 0.05; NS = not significantly different

^U RCRR = Rhizoctonia crown and root rot; 0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead

^V RCRR = Rhizoctonia crown and root rot; percent of roots with rating greater than two

^W Values represent mean of 48 plots (4 replicate plots across 4 at-planting treatments and 3 postemergence treatments)

^X Values represent mean of 24 plots (4 replicate plots across 2 varieties and 3 postemergence treatments)

^Y Values represent mean of 32 plots (4 replicate plots across 2 varieties and 4 at-planting treatments)

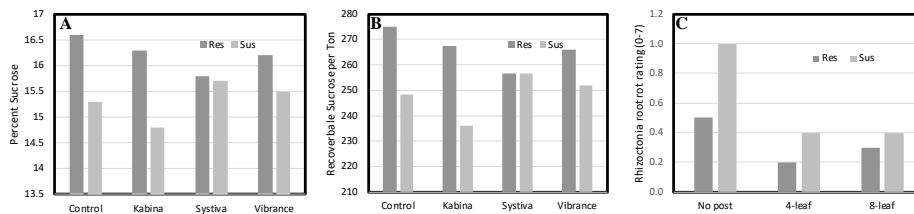


Fig. 3. MDFC site: Effect of variety and seed treatments on **A**) percent sucrose and **B**) recoverable sucrose per ton. Effect of variety and postemergence treatments on **C**) Rhizoctonia root rot severity (0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead).

Table 5. SMBSC site: Main effects of variety, at-planting (seed), and postemergence fungicide treatments on Rhizoctonia crown and root rot and sugarbeet yield and quality in a field trial sown May 12, 2017.

Main effect (Apron + Maxim on all seed)	RCRR (0-7) ^{TU}	RCRR % incidence ^{TV}	Yield ton A ^{-1T}	Sucrose [†]		
				%	lb ton ⁻¹	lb A ⁻¹
Variety ^W						
Resistant	0.2	4	27.9	15.4	255	7117
Moderately Susceptible	0.7	14	29.8	14.4	232	6875
ANOVA p-value	0.5720	0.5290	0.0167	0.0256	0.0301	0.2042
LSD (<i>P</i> = 0.05)	NS	NS	1.2	0.78	19	NS
At-planting treatments ^X						
Untreated control	0.4	9	29.2	15.1	248	7211
Kabina ST @ 14 g a.i./unit	0.5	9	29.5	14.8	239	7014
Systiva @ 5 g a.i./unit	0.4	8	28.3	15.1	246	6974
Vibrance @ 1.5 g a.i./unit	0.5	9	28.3	14.8	241	6785
ANOVA p-value	0.7040	0.9277	0.8082	0.2471	0.2165	0.7068
LSD (<i>P</i> = 0.05)	NS	NS	NS	NS	NS	NS
Postemergence fungicide ^Y						
None	0.6 a	11 a	29.1	14.9	242	7024
4-leaf Quadris @ 14.3 fl. oz./A	0.5 a	10 a	28.5	15.0	245	6950
8-leaf Quadris @ 14.3 fl. oz./A	0.3 b	5 b	29.0	14.9	244	7014
ANOVA p-value	0.0086	0.0043	0.4935	0.7539	0.6955	0.8390
LSD (<i>P</i> = 0.05)	0.19	3.7	NS	NS	NS	NS
Vty x Seed	NS	NS	NS	NS	NS	NS
Vty x Post	NS	NS	NS	NS	NS	NS
Seed x Post	0.0138	0.0222	NS	NS	NS	NS
Vty x Seed x Post	NS	NS	NS	NS	NS	NS

^T Numbers followed by the same letter are not significantly different; LSD = Least Significant Difference, *P* = 0.05; NS = not significantly different

^U RCRR = Rhizoctonia crown and root rot; 0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead

^V RCRR = Rhizoctonia crown and root rot; percent of roots with rating greater than two

^W Values represent mean of 48 plots (4 replicate plots across 4 at-planting treatments and 3 postemergence treatments)

^X Values represent mean of 24 plots (4 replicate plots across 2 varieties and 3 postemergence treatments)

^Y Values represent mean of 32 plots (4 replicate plots across 2 varieties and 4 at-planting treatments)

SMBSC site: This site received only 5.52 inches rainfall May-July in 2017 compared to 13.63 inches in 2016 making the early part of growing season on the drier side. The month of August received 6.92 inches rainfall followed by very dry September (1.34 inches and 4.84 inches in 2017 and 2016, respectively). Average four-inch bare soil temperatures at SMBSC were 57.7 °F and 70.2 °F for the months of May and June, respectively. Average four-inch bare soil temperature crossed 65 °F on June 01 which is typical for southern Minnesota. Low soil moisture during the growing season resulted in very low disease pressure at this site. There were significant (*P* < 0.05) seed treatment x postemergence application interactions for root rot rating and incidence and no three way interactions (Tab. 5). From 2 to 9 WAP there were no differences in stand between two varieties. However, by harvest, moderately susceptible variety had higher yield. Resistant variety had higher percent sugar and RST compared to moderately susceptible variety (Tab. 5). Stand data and harvest data were not different between seed treatments and untreated control (Tab.

5). Heavy rainfall in August created slight disease pressure in the plots leading to minor differences in disease severity and incidence between no Quadris and 4-8 leaf Quadris applications. Yield, percent sugar, RSA and RST were not significantly different between Quadris (4- or 8-leaf) and no Quadris application. Root rot severity and percent incidence (percent of roots with a disease rating of > 2.0) was higher for no Quadris and 4-leaf Quadris compared to 8-leaf Quadris for untreated control; highest for no Quadris, intermediate for 8-leaf and lowest for 4-leaf Quadris application for Kabina; higher for 4-leaf Quadris application compared to no or 8-leaf Quadris for Systiva; highest for no Quadris, intermediate for 4-leaf and lowest for 8-leaf Quadris application for Vibrance seed treatment (Figs. 4A and 4B).

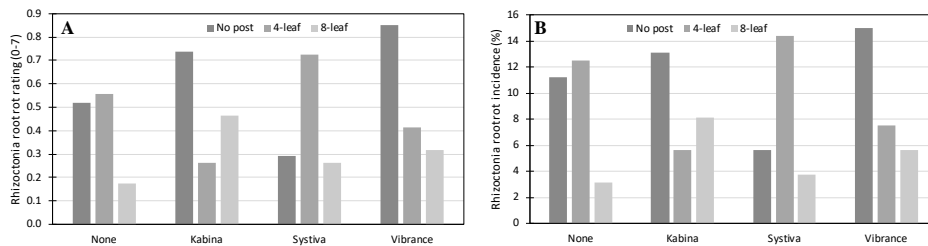


Fig. 4. SMBSC site: Effect of seed and postemergence treatments on **A)** *Rhizoctonia* root rot severity (0-7 scale (adjusted rating), 0 = root clean, no disease, 7 = root completely rotted and plant dead) and **B)** *Rhizoctonia* root rot incidence (percent of roots with rating greater than two).

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LITERATURE CITED

1. Brantner, J.R. 2015. Plant pathology laboratory: summary of 2013-2014 field samples. 2014 Sugarbeet Res. Ext. Rept. 44:138-139.
2. Brantner, J.R., H.R. Mickelson, and E.A. Crane. 2014. Effect of *Rhizoctonia solani* inoculum density and sugarbeet variety susceptibility on disease onset and development. 2013 Sugarbeet Res. Ext. Rept. 44:203-208.
3. Brantner, J.R. and C.E. Windels. 2011. Plant pathology laboratory: summary of 2009-2010 field samples. 2010 Sugarbeet Res. Ext. Rept. 41:260-261.
4. Brantner, J.R. and C.E. Windels. 2009. Plant pathology laboratory: summary of 2007-2008 field samples. 2008 Sugarbeet Res. Ext. Rept. 39:250-251.
5. Chanda, A. K., Brantner, J. R., Metzger, M., Bloomquist, M., and Groen, C. 2017. Integrated Management of *Rhizoctonia* on Sugarbeet with Varietal Resistance, At-Planting Treatments and Postemergence Fungicides. 2016 Sugarbeet Res. Ext. Rept. 47:174-179.

6. Crane, E., Brantner, J.R., and Windels, C.E. 2013. Plant pathology laboratory: summary of 2011-2012 field samples. 2012 Sugarbeet Res. Ext. Rept. 43:169-170.
7. Niehaus, W.S. 2017. Results of American Crystal's 2016 Official Coded Variety Trials. 2016 Sugarbeet Res. Ext. Rept. 47:207-259.

REAL-TIME PCR-BASED DETECTION OF *RHIZOCTONIA* LEVELS IN SOIL

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Rhizoctonia damping-off and crown and root rot (RCRR) caused by *Rhizoctonia solani* AG 2-2 have been the most common root diseases on sugarbeet in Minnesota and North Dakota for several years (1-3,6). Disease can occur throughout the growing season and reduces plant stand, root yield, and quality. Warm and wet soil conditions favor infection. Control options include rotating with non-host crops (cereals), planting partially resistant varieties, planting early when soil temperatures are cool, improving soil drainage, and applying fungicides as seed treatments, in-furrow (IF), or postemergence. An integrated management strategy should take advantage of multiple control options to reduce Rhizoctonia crown and root rot (5).

OBJECTIVE

To develop a real-time PCR assay for detection and quantification of DNA of *R. solani* AG 2-2 directly from soil samples for use in predicting inoculum potential.

MATERIALS AND METHODS

Soil sample collection. In 2016 we located 16 fields with a history of Rhizoctonia root rot based on the best knowledge of the agriculturists from ACSC (8 fields), MDFC (4 fields), and SMBSC (4 fields). From each field, 5 soil cores were taken at a depth of 6 inches representing approximately 1 acre area. Each soil core was divided into 0-2 inch, 2-4 inch and 4-6 inch sub-samples. In total, we collected 240 soil samples from all 16 fields (16 x 15). Total soil DNA was isolated from all 240 samples. At each sampling point (16 fields x 5 sites per field = 80 samples) where we collected soil cores, we also collected approximately 1 gallon of soil to determine Rhizoctonia root rot index (RRI) values using a growth chamber assay.

Growth chamber assay. For each of the 80 samples, soil was added to four 10 x 10 x 10 cm pots (~350 cc soil/pot). Seed was sown (25 seed/pot, 4 replicate pots/infested soil treatment) and then another 250 cc of soil was added over the seed to each pot. Pots were arranged in a randomized block design and incubated in a growth chamber at 77°F with a 14-hour photoperiod for 4 weeks. Pots were watered once or twice daily to keep soil moisture high to favor infection by *R. solani*. Seedlings were counted three times per week. Dying seedlings were removed and assayed in the laboratory to verify presence of *R. solani*. Four weeks after planting, remaining seedlings were removed from soil, washed, and rated on a 0-3 scale (0 = no disease, 3 = hypocotyl completely necrotic/plant dead). The number of seedlings that died during the 4-week assay along with the ratings after 4 weeks were used to calculate a root rot index (RRI, 0 = no disease, 100 = all plants died during the 4-week assay).

Root rot rating. In each of the 16 fields, 10 sugarbeet roots adjacent to the soil sampling site were rated for root rot severity using a 0-7 scale (10 roots x 5 spots = 50 roots per field).

Soil DNA isolation. PowerMax[®] soil DNA isolation kits from MO BIO Laboratories Inc. (Carlsbad, CA) were used for DNA isolation. Manufacturer's protocols were followed, using 5 g (dry wt.) of soil as starting material. Final DNA was eluted in 5 mL of Solution C6, concentrated and stored at -20 °C until downstream PCR application.

Real-time PCR. Primers and probe specific for internal transcribed spacer (ITS) region of *R. solani* anastomosis group (AG) 2-2 used in this study were developed by Budge et al. (4). All real-time PCR assays were set up as duplicate 20 µL reactions using LightCycler[®] 480 Probes Master (Roche Life Science) following manufacturer's protocols. 20x Custom TaqMan[®] Gene Expression Assay (contains 18 µM each primer and 5 µM 6-FAM[™] dye-labeled TaqMan[®] MGB probe) was obtained from Life Technologies (Carlsbad, CA) and 1 µL of DNA template was used in the assay. Thermal cycling parameters were 50 °C for 2 min, 95 °C for 10 min, followed by 40 cycles of 95

°C for 15 sec and 60 °C for 1 min. *R. solani* AG 2-2 IIIB and IV DNA (10ng/ μL) as positive control and no template control was included in each run. Real-time PCR assays were performed using Roche LightCycler® 480 System.

RESULTS AND DISCUSSION

The mean root rot ratings ranged from 1.12 to 3.72 and root rot incidence values ranged from 18 to 68 % for 16 fields. The mean root rot index (RRI) ranged from 0 to 100% for 16 fields. The lowest Ct value of 27.02 (highest Rhizoctonia DNA) was found in one field in MDFC area (MDFC4). There was a significant correlation at the field level between RRI and root rot ratings ($r = 0.59$ and $r^2 = 0.34$), and RRI and root rot incidence values ($r = 0.56$ and $r^2 = 0.32$). There was a significant correlation between RRI and DNA of *R. solani* ($r = 0.24$; $r^2 = 0.06$) (Fig. 1). However, some soil samples with higher RRI had lower DNA of *R. solani* Ag 2-2. This could be explained by the presence of other AG groups such as AG 4 in soil which can cause seedling damping off.

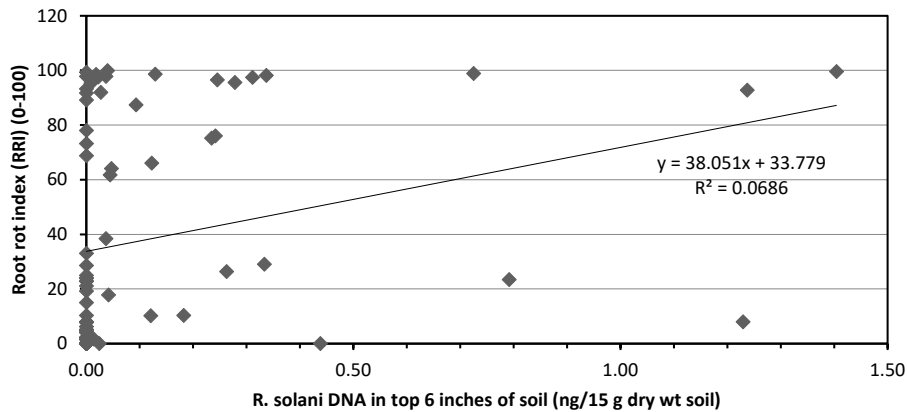


Figure 1. Relationship between Rhizoctonia root rot index and amount of DNA of *Rhizoctonia solani* AG 2-2 in soil

There was also a significant correlation between root rot rating and DNA of *R. solani* ($r = 0.31$; $r^2 = 0.11$) (Fig. 2). However, soil samples from some fields with higher root rot ratings had lower amounts of DNA of AG 2-2. This could be explained by the non-uniform distribution of Rhizoctonia inoculum in the soil. We also observed Aphanomyces in some of the soil samples corroborating the evidence that mixed infestation of soil with Rhizoctonia and Aphanomyces is gradually increasing in our growing region. Out of 80 samples, DNA of Rhizoctonia was detected in twenty nine 0-2 inch samples, fifteen 2-4 inch samples, and eleven 4-6 inch samples. In only 6 out of 80 samples, DNA of Rhizoctonia was detected in all 0-2, 2-4 and 4-6 inches depths.

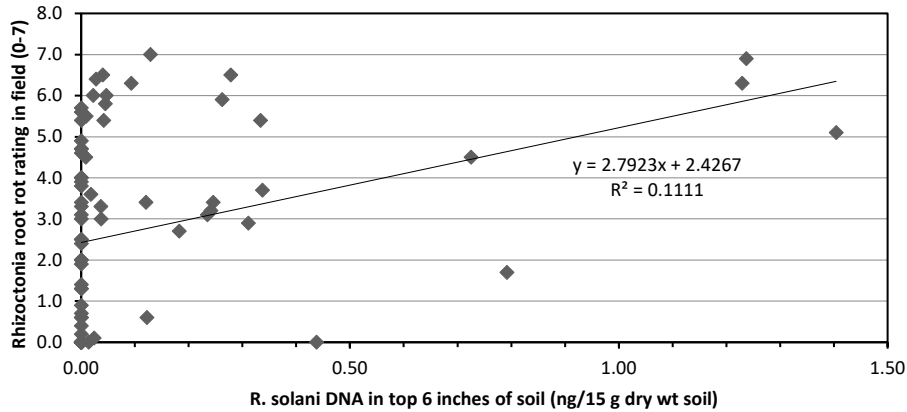


Figure 2. Relationship between *Rhizoctonia* root rot rating and amount of DNA of *Rhizoctonia solani* AG 2-2 in soil.

ACKNOWLEDGEMENTS

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LITERATURE CITED

8. Brantner, J.R. 2015. Plant pathology laboratory: summary of 2013-2014 field samples. 2014 Sugarbeet Res. Ext. Rept. 44:138-139.
9. Brantner, J.R. and C.E. Windels. 2011. Plant pathology laboratory: summary of 2009-2010 field samples. 2010 Sugarbeet Res. Ext. Rept. 41:260-261.
10. Brantner, J.R. and C.E. Windels. 2009. Plant pathology laboratory: summary of 2007-2008 field samples. 2008 Sugarbeet Res. Ext. Rept. 39:250-251.
11. Budge, G.E., Shaw, M.W., Colyer, A., Pietravalle, S., and N. Boonham. 2009. Molecular tools to investigate *Rhizoctonia solani* distribution in soil. *Plant Pathol.* 58:1071-1080.
12. Chanda, A. K., Brantner, J. R., Metzger, M., Bloomquist, M., and Groen, C. 2017. Integrated Management of *Rhizoctonia* on Sugarbeet with Varietal Resistance, At-Planting Treatments and Postemergence Fungicides. 2016 Sugarbeet Res. Ext. Rept. 47:174-179.
13. Crane, E., Brantner, J.R., and C.E. Windels. 2013. Plant pathology laboratory: summary of 2011-2012 field samples. 2012 Sugarbeet Res. Ext. Rept. 43:169-170.

REMOTE SENSING FOR DETECTION OF RHIZOCTONIA CROWN AND ROOT ROT OF SUGARBEET

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Rhizoctonia crown and root rot (RCRR), caused by *Rhizoctonia solani* AG 2-2, is becoming more frequent and widespread in the sugarbeet-growing regions of Minnesota and North Dakota. In this region, symptoms of RCRR typically begin at about 8 weeks after planting and continue to develop until harvest. Infected plants occur sporadically or in large portions of the field. Advances in remote sensors and vehicle platforms have regenerated interest in within-season aerial mapping/detection of RCRR.

Remotely assessing plant health by measuring the reflectance of incident electromagnetic radiation is well established (Nilsson, 1995). Evaluating the relative canopy reflectance of specific wavelengths can provide insight into the impacts of insects (e.g. Alves et al. 2015), nutrient state (e.g. Felderhof & Gillieson 2012) and disease (well-reviewed in Oerke et al. 2014). Specifically, spectral reflectance has been demonstrated to show the presence of *Rhizoctonia solani* AG 2-2 prior to the development of visible symptoms (Reynolds et al. 2009 and 2012).

Detection of plant diseases is based on pathogen-induced degradation of chlorophyll and subsequent deterioration of palisade parenchyma cells in the leaf (Inoue, 2003). Chlorophyll absorbs red wavelengths of energy, so its degradation results in increased reflectance in the Red wavelengths (~620nm-680nm). Parenchyma cells reflect Near-Infrared (NIR) energy (~700nm-1100nm) and their deterioration results in lowered reflectance of NIR. The comparative ratio of these wavelengths can, therefore, provide insight into the level of stress being endured by a plant. Remotely sensed plant reflectance has been used for detecting sugarbeet diseases including Rhizomania (Steddom et al., 2005), Cercospora leaf spot (Steddom et al., 2005), sugarbeet cyst nematode (Schmitz et al., 2003), and RCRR (Laudien et al., 2003, 2004, 2006, Reynolds et al. 2009 and 2012). Most early research detected RCRR at the end of the growing season but did not address earlier-season detection of the disease or the relationship of reflectance to severity of root rot symptoms. There have been indications, however, that earlier identification of infection may be possible. Reynolds et al. (2009 and 2012) found that specific wavelengths and vegetative indices were more closely associated with plants showing lower root ratings of the disease. While several indices were correlated with disease severity, the narrowband *modified Spectral Ratio* (mSR) index, calculated as $[(R750-R445)/(R705-R445)]$ where R is the measure of reflection in that wavelength, appeared to allow for earlier detection of RCRR than did wideband indices such as *Normalized Difference Vegetative Index* (NDVI) or *Optimized Soil Adjusted Vegetative Index* (OSAVI) that were correlated with RCRR infection severity (Reynolds 2010). Unfortunately, this research effort was not able to further pursue and refine the further diagnostic capabilities of reflectance measurements.

The objectives of this study were 1) to establish baseline hyperspectral reflectance data associated with disease severity for RCRR of partially resistant and susceptible sugarbeet varieties 2) to identify the most sensitive and strongest diagnostic reflectance data (wavelengths) for detection of disease.

Materials and Methods:

Field experiments and design - In 2016/ and 2017 sugarbeet varieties partially resistant (4.0 rating) and susceptible (4.7 rating) to RCRR were planted in field plots (4 replications) at the University of Minnesota, NWROC. Plots were 35 ft long by 4 rows wide. In inoculated plots, *Rhizoctonia solani* AG 2-2 IIIB infested barley grain (at three different rates; 20 kg/ha, 40 kg/ha, and 60 kg/ha) was broadcast at planting to mimic naturally infested Rhizoctonia fields with low, medium, and high levels. For one treatment, near canopy closure, plants were inoculated (four center rows of six-row plots) with *R. solani* AG 2-2 IIIB infested barley grains by placing two whole kernels about 1-inch below the soil surface and adjacent to the root. One treatment included non-inoculated control. Plots were split into a 22-ft section (4-rows wide) for visual assessment of disease (0-7 scale, 0 = root clean, 7 = root completely rotted), with the remaining 13-ft length (4-rows wide) left for reflectance imagery. At each sample date, twelve plants were randomly selected and pulled from one end from the 22' section of each plot and their roots visually assessed for disease

symptoms. Simultaneously, one of the oldest leaves from that plant was assessed for reflectance using the spectroradiometer.

Reflectance data - Hyperspectral reflectance of individual leaves on plants was obtained using an Ocean Optics Flame hyperspectral radiometer. The Ocean Optics Flame is sensitive to the visual and near-infrared wavelengths (VIS/NIR) typically ranging from ~350nm to ~1100nm, however due to signal loss at the extreme edges of the sensor, only wavelength responses between 400nm – 950nm were used in data analyses. A self-illuminated, leaf-clip sensor was used with the Ocean Optics radiometer, the illuminator of which provided light over all wavelengths that the sensor can read (350nm-1100nm). Consequently, the absolute reflectance, within sensitivity limits, of the plant's leaf surface could be assessed as the percent of light energy being reflected (reflectance). Multispectral imagery was also obtained using multi-sensor arrays which included a standard visual RGB camera and 3 cameras sensitive to specified wavelengths in the Near-Infrared (NIR) (Sentra, Minneapolis MN). These sensors were mounted on small Unmanned Aerial Systems (i.e. drones). Measurements were obtained weekly starting at 8 weeks after planting and 2 weeks after whole barley inoculation (~11 am to 2 pm).

Disease data - Disease ratings were taken from sampled plants at the same time as reflectance data was measured. Tap roots were visually assessed for RCRR using a 0-to-7 scale (Ruppel et al. 1979), where 0 = no visible lesions; 1 = superficial, scattered inactive lesions; 2 = shallow, dry rot cankers or active lesions on ≤5% of root surface; 3 = deep dry-rot cankers at crown or extensive lateral lesions affecting 6 to 25% of the root; 4 = rot affecting 26 to 50% of tap root, with cracks and cankers up to 5 mm deep; 5 = 51 to 75% of tap root blackened, with rot extending into interior and roots usually misshapen with cracks and rifts; 6 = entire root blackened except extreme tip; and 7 = root 100% rotted and foliage dead.

Data analyses – Disease ratings were assigned the percentage of root covered with necrosis associated with that rating (i.e. rating of: 0=0%, 1=2.5%, 2=5%, 3=25%, 4= 50%, 5=75%, 6=95%, 7=100% root covered). The percent of root covered by necrosis values (% necrosis) were then transformed using ArcSine of the square root to normalize the data. To assess the wavelengths of reflected light most influenced by RCRR infection, individual wavelengths (from 400nm to 950nm) were correlated with %necrosis for all individual plants at all dates. The respective correlation coefficients from wavelengths and disease rating were used to calculate the Akaike Information Criteria (AIC) values which were used to rank each wavelength for its ability to predict RCRR disease rating (Akaike 1974). The AIC is a metric used to evaluate best fit, in this case, which wavelengths are most closely associated with RCRR disease rating. Lower AIC ratings indicate better fit (Burnham and Anderson, 2003). The AIC values were then compared using a Relative Likelihood calculation ($RL_i = \exp((\text{lowest AIC} - AIC_i)/2)$), where RL_i is the Relative Likelihood at a specific wavelength (i) of it being best fit of all AICs (i.e. most closely associated with RCRR infection).

Results and Discussion:

Plots were sampled 7/10, 7/13, 7/14, 7/18, 7/20, 7/26, 8/02, 8/08, and 8/15. Unfortunately, due to equipment failure, data from 7/26 had to be discarded from the analyses. All other dates had expressed symptoms in inoculated plots (for both resistant and susceptible varieties), resulting a gradient of infection throughout the season.

The leaf clip readings from the Ocean Optics Flame Spectroradiometer returned the percent reflectance of light in wavelengths from 350nm to 1100nm (fig 1). Unfortunately, the design of CCD-based sensors, such as those used in most VIS/NIR spectroradiometers, results limitations at the extreme edges of the sensor's range, resulting in increased 'noise' and loss of signal integrity. As can be seen in Fig 1, the signal to noise ratio of data below 450nm and above 950 nm deteriorates rapidly. Consequently, we decided to use reflection responses at wavelengths from 450nm-950nm only for analyses (fig 2).

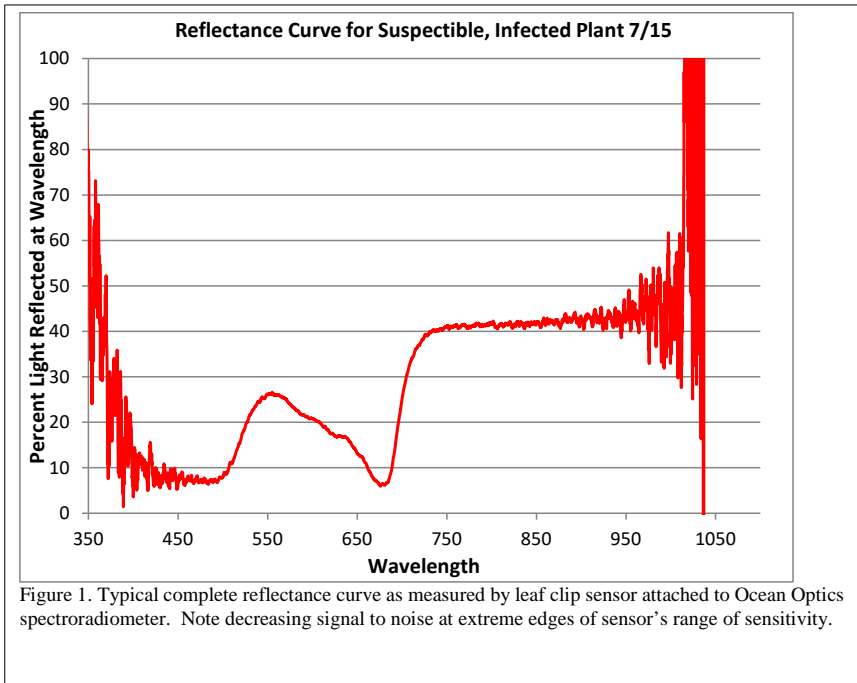


Figure 1. Typical complete reflectance curve as measured by leaf clip sensor attached to Ocean Optics spectroradiometer. Note decreasing signal to noise at extreme edges of sensor's range of sensitivity.

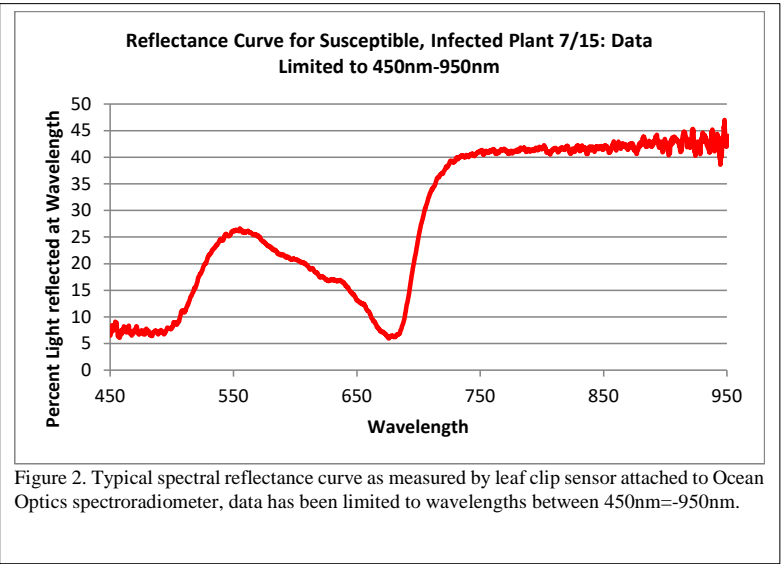


Figure 2. Typical spectral reflectance curve as measured by leaf clip sensor attached to Ocean Optics spectroradiometer, data has been limited to wavelengths between 450nm-950nm.

A multiple correlation incorporating date and variety was used to assess the disease ratings of 12 plants per plot at each date (40 plots X 12 plants = 480 plants per sample date) with all 500 analyzed wavelengths (450nm-950nm) (a total of 240,000 wavelengths associated with disease ratings were assessed at each sample date). The resulting

correlations were assessed for association to RCRR disease rating using AIC, AIC's were plotted against assessed wavelength for all dates (fig 3) (resulting in a total of 1,680,000 wavelengths associated with disease ratings – this multiple regression will stress computing limits!). The lowest AIC value on the AIC vs Wavelengths graph is considered to be the most important (i.e. in our case, the wavelengths most closely associated with RCRR infection).

It is important to note that the AIC is not a statistic and therefore cannot be directly compared, its units depend on the value of the coefficients in the association analysis and on the number of parameters in the model being assessed. To compare AIC values of wavelengths to assess which wavelengths was most closely associated with RCRR infection, a Relative Likelihood analysis was conducted (fig 4).

There is increasing variation in AIC values above 750 nm, this reflects the variation in reflectance values found in that range; there was no clear association of reflection of NIR above 750 and RCRR infection. The lowest AIC values appear just below 700nm (the range between red and NIR wavelengths). The relative Likelihood analysis indicates that, in our data, the wavelengths most closely associated with RCRR infection occur in a narrow band of approximately 20nm, centered around 689nm. The next highest peak occurred at 698 nm (although this may actually be a part of the 689 narrow bandwidth. Most sources consider these values to be technically red wavelengths, although some sources

$$AIC = -2*LN(\lambda) + (2*P)$$

Where:
 λ = metric of association (coefficient)
and
P = # parameters in the model

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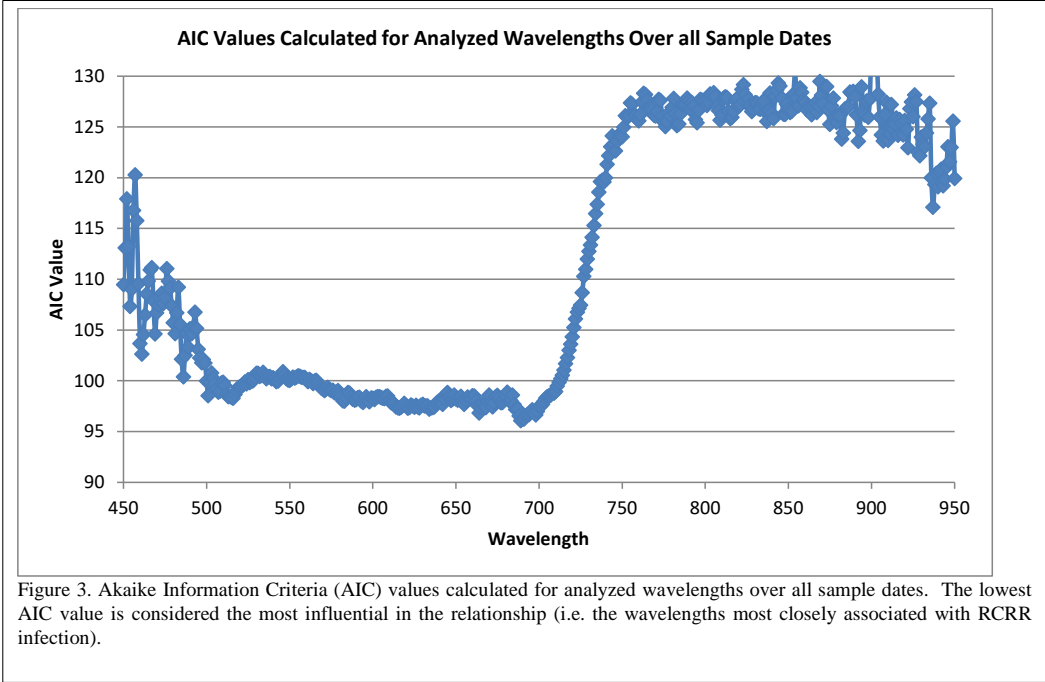


Figure 3. Akaike Information Criteria (AIC) values calculated for analyzed wavelengths over all sample dates. The lowest AIC value is considered the most influential in the relationship (i.e. the wavelengths most closely associated with RCRR infection).

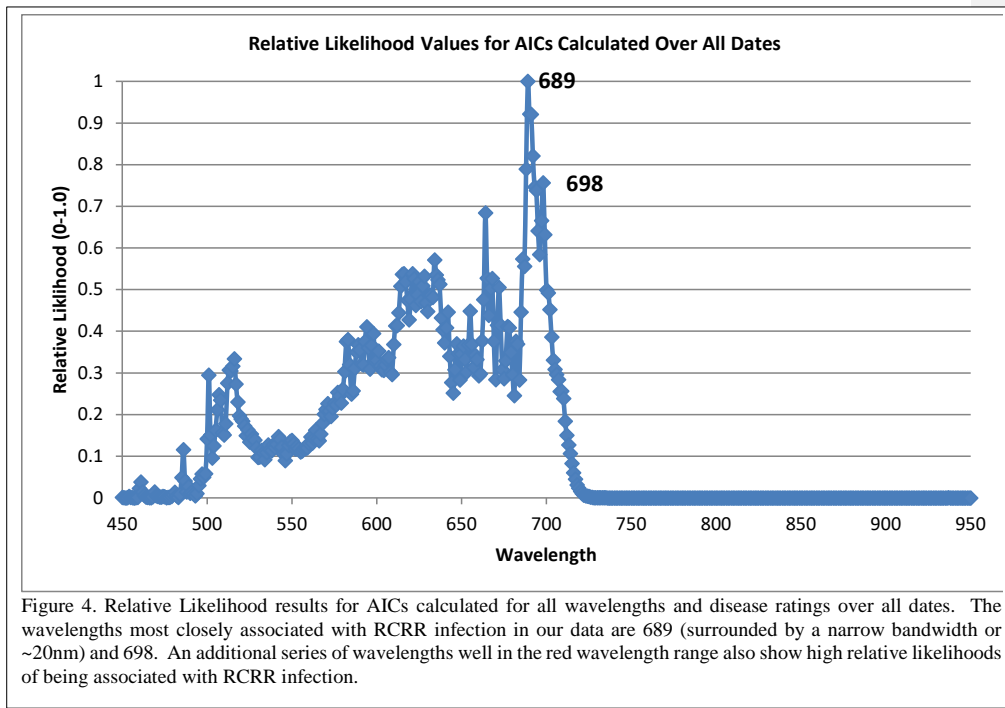


Figure 4. Relative Likelihood results for AICs calculated for all wavelengths and disease ratings over all dates. The wavelengths most closely associated with RCRR infection in our data are 689 (surrounded by a narrow bandwidth or ~20nm) and 698. An additional series of wavelengths well in the red wavelength range also show high relative likelihoods of being associated with RCRR infection.

place them in the low near-infrared. In either case, this range of wavelengths where red and NIR abut, sometimes inaccurately called the 'Red Edge', is an area where stress reactions in plants often affect the leaf's ability to reflect solar energy. So these results are not surprising. In addition to the narrow band around 689, there are several groups of wavelengths in the range of red wavelengths that show high relative likelihood of being associated with RCRR infection (582nm-602nm, 615nm-634nm, and a very narrow band between 663nm-665nm). Results from aerial imagery from VIS/NIR multi-sensor arrays and Forward Looking Infrared (FLIR) thermal cameras (e.g. fig 5) from all dates are still being analyzed. Results from these sensors will be combined with the reflection data to refine the ability to remotely sense *Rhizoctonia* Crown and Root Rot.

Conclusions:

In 2017, we found that RCRR does significantly affect the reflection characteristics of infected plants. The data in this study indicates that the most effective wavelengths to assess RCRR infection are a 20nm band centered around 689nm with additional narrow bands of wavelengths in other areas of the red wavelengths. These wavelengths may provide the best potential for the development of RCRR sensors, although additional data from aerial VIS/NIR and thermal imagery may provide significant improvement.

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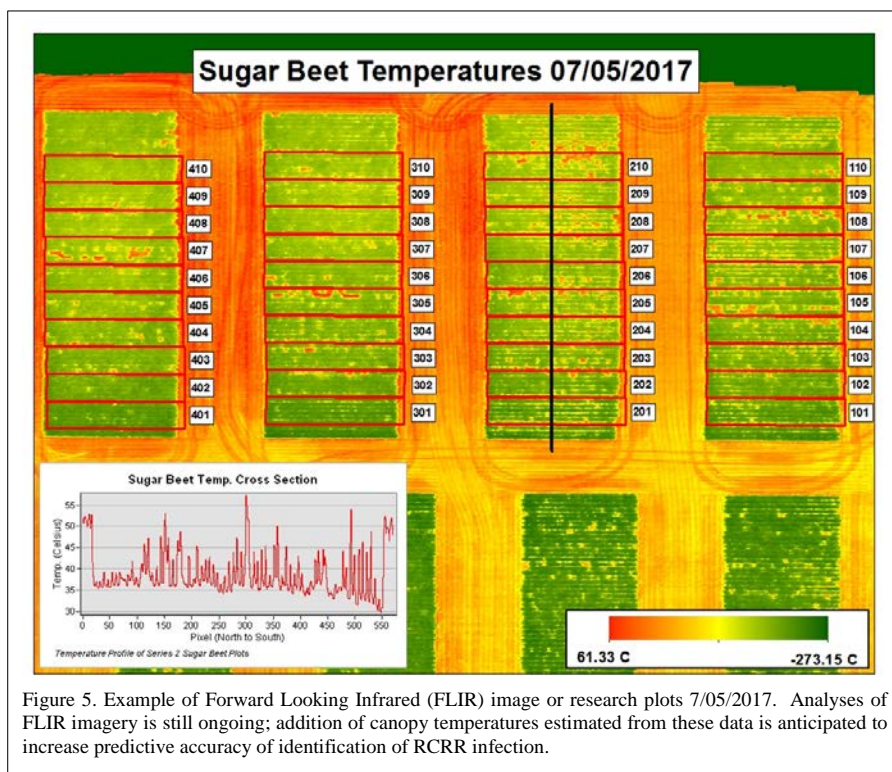


Figure 5. Example of Forward Looking Infrared (FLIR) image of research plots 7/05/2017. Analyses of FLIR imagery is still ongoing; addition of canopy temperatures estimated from these data is anticipated to increase predictive accuracy of identification of RCRR infection.

Literature Cited

- Alves, T. M., I. V. MacRae, and R. L. Koch. 2015. Soybean aphid (Hemiptera: Aphididae) affects soybean spectral reflectance. *Journal of Economic Entomology*, DOI: <http://dx.doi.org/10.1093/jee/fov250>.
- Akaike, H., 1974. A new look at the statistical model identification. *IEEE Trans. Automat. Contr.* 19, 716–723. doi:10.1109/TAC.1974.1100705
- Burnham, K.P. and Anderson, D.R., 2003. *Model selection and multimodel inference: a practical information-theoretic approach*. Springer Science & Business Media.
- Felderhof, L. and Gillieson, D. 2012. Near-infrared imagery from unmanned aerial systems and satellites can be used to specify fertilizer application rates in tree crops. *Canadian Journal of Remote Sensing*, 37(4): 376-386.
- Inoue, Y. 2003. Synergy of remote sensing and modeling for estimating ecophysiological processes in plant production. *Plant Production Sci.* 6:3-5.
- Laudien, R. G. Bareth, and R. Doluschitz. 2003. Analysis of hyperspectral field data for detection of sugar beet diseases. *Proc. EFITS Conf*, 2003.
- Laudien, R., G. Bareth, and R. Doluschitz. 2004. Comparison of remote sensing based analysis of crop diseases by using high resolution multispectral and hyperspectral data – case study: *Rhizoctonia solani* in sugar beet. Pages 670-676 in: *Proc. 12th Int. Conf Geoinformatics-Geospatial Information Research: Bridging the Pacific and Atlantic*, Univ. Gavle, Sweden, 7-9 June 2004.

- Laudien, R., K. Burcky, R. Doluschitz, and G. Bareth. 2006. Establishment of a web-based spectral database for the analysis of hyperspectral data – case study: *Rhizoctonia solani*-inoculated sugarbeets. *Zuckerindustrie* 131:164-170.
- Nilsson, H.E. 1995. Remote sensing and image analysis in plant pathology. *Annu. Rev. Phytopathology* 15:489-527.
- Oerke, E. C., Mahlein, A. K., & Steiner, U. (2014). Proximal Sensing of Plant Diseases. In *Detection and Diagnostics of Plant Pathogens* (pp. 55-68). Springer Netherlands.
- Reynolds, G. J. 2010. Remote sensing for detection of *Rhizoctonia* crown and root rot in Sugar Beet and the impact of the disease on chlorophyll Content. Dissertation submitted to the University of Minnesota.
- Reynolds, G.J., C.E. Windels, I.V. MacRae, and S. Laguette. 2009. Hyperspectral remote sensing for detection of *Rhizoctonia* crown and root rot in sugar beet. *Phytopathology* 99(6): s108-s108.
- Reynolds, G.J., C.E. Windels, I.V. MacRae, and S. Laguette. 2012. Remote sensing for assessing *Rhizoctonia* crown and root rot in sugar beet. *Phytopathology* 96(4): 497-505.
- Ruppel, E.G., C.L. Schneider, R.J. Hecker, and G.I. Hogaboam. 1979. Creating epiphytotics of *Rhizoctonia* root rot and evaluating for resistance to *Rhizoctonia* so/ani in sugarbeet field plots. *Plant Dis.* 63:518-522.
- Schmitz, A., S. Kiewnick, J. Schlang, K. Schmidt, and R.A. Sikora. 2003. Use of remote sensing to identify the spatial distribution of the sugar beet cyst nematode *Heterodera schachtii*. Pages 561-562 In: Program book of the Joint Conference of ECPA-ECPLF, A. Werner and A. Jarfe (Hrsg.)
- Steddom, K., M. W. Bredehoeft, M. Khan, and C. M. Rush. 2005. Comparison of visual and multispectral radiometric disease evaluations of *Cercospora* leaf spot of sugar beet. *Plant Disease.* 89(2):153-158.
- Xu, J., & B. Su. 2017. Significant remote sensing vegetative indices: a review of developments and applications. *J. of Sensors* 2017. <https://doi.org/10.1155/2017/1353691>.

COMPARISON OF POSTEMERGENCE FUNGICIDES FOR CONTROL OF RHIZOCTONIA CROWN AND ROOT ROT OF SUGARBEET

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Rhizoctonia crown and root rot (RCRR) caused by the soilborne fungus *Rhizoctonia solani* AG 2-2 is a common problem in the sugarbeet growing areas of Minnesota and North Dakota. The disease can cause damping-off on seedlings and infect older roots throughout the growing season. Warm (65°F+) soil combined with excess moisture conditions favor infection and disease development. Control methods include rotating with non-host (cereal) crops such as wheat, sowing partially resistant varieties, and the use of seed treatment, in-furrow, and/or postemergence fungicides. Several options are available to sugarbeet growers for control of Rhizoctonia, including some newly registered products. Data is needed to compare new fungicides alongside established postemergence fungicides.

OBJECTIVES

A field trial inoculated with *R. solani* AG 2-2 was established to evaluate postemergence application of several registered and other fungicides for control of Rhizoctonia crown and root rot and effect on sugarbeet yield and quality.

MATERIALS AND METHODS

The trial was established at the University of Minnesota, Northwest Research and Outreach Center, Crookston on a Hegne-Fargo complex soil. Prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley broadcast at 35 kg ha⁻¹ and incorporated with a Rau seedbed finisher. The trial was sown with the moderately susceptible cultivar 'Crystal 101RR' in six-row plots (22-inch row spacing) on May 8 at 4.5-inch seed spacing. Seed treatments included standard rates of Apron + Thiram, Tachigaren (45 g product/unit), and Kabina ST (14 g a.i./unit). Counter 20G (8 lb/A) was applied at planting for control of root maggot. Glyphosate (4.5 lb product ae/gallon) was applied on June 5 (22 oz A⁻¹) and 21 (28 oz A⁻¹), and July 5 (32 oz A⁻¹) for control of weeds. Treatments were assigned to plots (6 rows wide, 25 ft long) arranged in a randomized block design with four replicates. Postemergence fungicides were applied June 16 in a 7-inch band using 10 gallons of water/A with the exception of a broadcast application of the 14 fl oz A⁻¹ rate of Quadris. Fungicides, active ingredients, and rates are summarized in Table 1, including 17.6 fl oz A⁻¹ AZteroid (azoxystrobin), 10 and 14 fl oz A⁻¹ Quadris (azoxystrobin), 7 fl oz A⁻¹ Topguard EQ (azoxystrobin + flutriafol), 6.7 fl oz A⁻¹ Priaxor (fluxapyroxad + pyraclostrobin) plus 0.25% non-ionic surfactant (NIS), and 5.7 fl oz A⁻¹ Proline (prothioconazole) plus 0.125% NIS by volume. Plants were inoculated 3 days after fungicide treatments by applying *R. solani*-infested ground barley inoculum (23 g/25 ft of row) over each of the center four rows by hand. Following inoculation, plots were cultivated and loosened soil was hand-raked into crowns to create a favorable environment for infection with *R. solani*. A no-fungicide, inoculated control was also included. Stand counts were taken on June 9, 19, and 26.

Table 1. Product names, active ingredients, and rates used in a field trial evaluating postemergence fungicides for control of *Rhizoctonia solani* AG 2-2 on sugarbeet. Standard rates of Apron + Thiram, 45 g product/unit Tachigaren, and 14 g a.i./unit Kabina were on all seed. Fungicides were applied June 16 in a 7-inch band or broadcast using 10 gallons of water/A.

Product	Active ingredient	Product rate	Active ingredient rate	Application
Untreated control	-	-	-	-
AZteroid	Azoxystrobin	17.6 fl oz A ⁻¹	103 g A ⁻¹	Band
Quadris	Azoxystrobin	10 fl oz A ⁻¹	73 g A ⁻¹	Band
Quadris	Azoxystrobin	14 fl oz A ⁻¹	103 g A ⁻¹	Band
Quadris	Azoxystrobin	14 fl oz A ⁻¹	103 g A ⁻¹	Broadcast
Topguard EQ ^z	Azoxystrobin + Flutriafol	7 fl oz A ⁻¹	61 + 45 g A ⁻¹	Band
Priaxor + NIS (0.25%)	Pyraclostrobin + Fluxapyroxad	6.7 fl oz A ⁻¹	66 + 33 g A ⁻¹	Band
Proline + NIS (0.125%)	Prothioconazole	5.7 fl oz A ⁻¹	81 g A ⁻¹	Band

^z Topguard EQ is registered in sugar beet only for use on foliar diseases

Cercospora leaf spot was controlled by Supertin + Topsis M (6 + 10 oz product in 19 gallons of water A⁻¹) applied with 8002 flat fan nozzles at 100 psi on July 25 and Inspire (7 oz product in 19 gallons of water A⁻¹) on August 8. The

trial was harvested on October 4 and data were collected for number of harvested roots, yield, and quality. The number of harvested roots and baseline stand counts at the time of inoculation (June 19) were used to calculate percent stand loss. Twenty roots per plot also were arbitrarily selected and rated for severity of RCRR using a 0 to 7 scale (0 = healthy root, 7 = root completely rotted and foliage dead). Disease incidence was reported as the percent of rated roots with a root rot rating of > 2.

Data were subjected to analysis of variance using SAS Proc GLM (SAS Institute, Cary, NC). Means were separated by Fisher's Protected Least Significant Difference ($P = 0.05$).

RESULTS AND DISCUSSION

Harvest data is summarized in Table 2. Moderate rainfall in June (3.41 inches) resulted in adequate disease pressure for some infection, but low rainfall in July and August (1.42 and 0.77 inch, respectively) slowed development of disease. Percent stand loss, RCRR rating and incidence, and root and sucrose yields were significantly different ($P = 0.05$) among treatments (Table 2). Percent stand loss and RCRR ratings and incidence were lower for all postemergence fungicides compared to the untreated control (Table 2). Among fungicide treatments, root and sucrose yields were highest for Topguard EQ, lowest for Priaxor, and intermediate for AZteroid, Quadris treatments, and Proline (Table 2). Percent sucrose and sucrose per ton were highest for the band applications of Quadris and Proline, lowest for the untreated control and Priaxor, and intermediate for AZteroid, broadcast Quadris and Topguard EQ (Table 2).

Stand loss in the untreated control was 66%, while RCRR rating and incidence averaged 3.7 and 75%, respectively, indicating a fairly high level of disease, despite dry soil conditions throughout July and August. Yet all postemergence fungicides provided significant control of RCRR and increased root and sucrose yield compared to the untreated control. In this trial, the broadcast application of Quadris and band application of Proline performed similarly to the band applications of Quadris. In past trials, these treatments have given mixed results. This trial was inoculated both prior to planting and at the time of fungicide application. Properly timed postemergence fungicides have good potential for decreasing RCRR and increasing root and sucrose yield.

Table 2. Effect of postemergence fungicides on percent stand loss, RCRR ratings and incidence, and root and sucrose yields in a sugar beet field trial inoculated with *Rhizoctonia solani* AG 2-2.

Treatment ^v	Percent stand loss ^{wx}	RCRR ^w (0-7) ^y	RCRR ^w % Incidence ^z	Yield ^w T/A	Sucrose ^w		
					%	lb/ton	lb recov./A
Untreated control	66 a	3.7 a	75 a	23.2 c	16.9 c	314 b	7324 c
AZteroid @ 17.6 fl oz A ⁻¹	22 b	0.7 b	15 b	33.6 ab	17.6 ab	330 ab	11084 ab
Quadris @ 10 fl oz A ⁻¹	15 b	0.9 b	16 b	33.5 ab	17.9 a	336 a	11272 a
Quadris @ 14 fl oz A ⁻¹	27 b	1.2 b	25 b	31.9 ab	17.7 a	334 a	10659 ab
Quadris @ 14 fl oz A ⁻¹ broadcast	14 b	1.1 b	21 b	33.4 ab	17.4 abc	327 ab	10944 ab
Topguard EQ @ 7 fl oz A ⁻¹	23 b	1.1 b	20 b	35.5 a	17.5 abc	330 ab	11715 a
Priaxor @ 6.7 fl oz A ⁻¹	25 b	1.5 b	26 b	31.0 b	16.9 bc	316 b	9809 b
+ NIS (0.25%)							
Proline @ 5.7 fl oz A ⁻¹	25 b	1.6 b	33 b	32.7 ab	17.9 a	336 a	11013 ab
+ NIS (0.125%)							
ANOVA P-value	0.0001	<0.0001	<0.0001	<0.0001	0.0297	0.0460	0.0001
LSD ($P = 0.05$) ^w	17.6	0.92	18.4	3.86	0.68	15.4	1451

^v Postemergence fungicide applications were made on June 16 using 10 gallons of water/A in a 7-inch band except where noted as broadcast; prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley broadcast at 35 kg ha⁻¹ and incorporated with a Rau seedbed finisher; plots were inoculated again on June 16 (after fungicide applications) by applying *R. solani*-infested ground barley inoculum (23 g/25 ft of row) over each of the center four rows by hand, followed by cultivation and hand-raking to move some soil into the crowns.

^w For each column, numbers followed by the same letter are not significantly different according to Fisher's Protected Least Significant Difference (LSD); NS = not significantly different

^x Percent stand loss = percent of stand present at the time of inoculation that died by harvest;

^y RCRR = Rhizoctonia crown and root rot; 0-7 scale, 0 = root clean, no disease, 7 = root completely rotted and plant dead

^z RCRR = Rhizoctonia crown and root rot; percent of roots with rating > 2

ACKNOWLEDGEMENTS

We thank Jeff Nielsen, Hal Mickelson, and student workers Tim Cymbaluk, Brandon Kasprick, and Muira MacRae; the University of Minnesota, Northwest Research and Outreach Center, Crookston for providing land, equipment and

other facilities; Crystal Beet Seed for providing seed; BASF, Bayer, FMC Agricultural Solutions, Syngenta, and Vive Crop Protection for fungicide samples; and American Crystal Sugar Company Quality Laboratory, East Grand Forks, MN for quality analyses.

EVALUATION OF AT-PLANTING FUNGICIDE TREATMENTS FOR CONTROL OF *RHIZOCTONIA SOLANI* ON SUGARBEET

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Rhizoctonia damping-off and crown and root rot (RCRR) caused by *Rhizoctonia solani* AG 2-2 have been the most common root diseases on sugarbeet in Minnesota and North Dakota for several years (1-2, 4-5, 8). Disease can occur throughout the growing season and reduce plant stand, root yield, and quality. Warm and wet soil conditions favor infection. Disease management options include rotating with non-host crops (cereals), planting partially resistant varieties, planting early when soil temperatures are cool, improving soil drainage, and applying fungicides as seed treatments, in-furrow (IF), and/or postemergence. An integrated management strategy should take advantage of multiple control options to reduce Rhizoctonia crown and root rot.

OBJECTIVES

A field trial was established to evaluate various at-planting fungicide treatments (seed treatment and in-furrow) for 1) control of early-season damping-off and RCRR and 2) effect on yield and quality of sugarbeet.

MATERIALS AND METHODS

The trial was established at the University of Minnesota, Northwest Research and Outreach Center (NWROC), Crookston. Field plots were fertilized for optimal yield and quality. A moderately susceptible variety (Crystal 101RR) with a 2-year average Rhizoctonia rating of 4.7 was used (9). A randomized complete block design with four replications was used. Seed treatments and rates are summarized in Table 1 and were applied by Germain's Seed Technology, Fargo, ND. In-furrow fungicides (Table 1) were applied down the drip tube in 6 gallons total volume A⁻¹. The untreated control included no seed or in-furrow fungicide treatment at planting. Prior to planting, soil was infested with *R. solani* AG 2-2-infested whole barley broadcast at 35 kg ha⁻¹ and incorporated with a Rau seedbed finisher. The trial was sown in six-row plots (22-inch row spacing, 25-ft rows) on May 11 at 4.5-inch seed spacing. Counter 20G (8 lb A⁻¹) was applied at planting for control of sugarbeet root maggot and 3 gallons A⁻¹ starter fertilizer (10-34-0) was applied across all treatment combinations. Glyphosate (4.5 lb product ae/gallon) was applied on June 5 (22 oz A⁻¹) and 21 (28 oz A⁻¹), and July 5 (32 oz A⁻¹) for control of weeds. Cercospora leaf spot was controlled by Supertin + Topsin M (6 + 10 oz product in 19 gallons of water A⁻¹) applied with 8002 flat fan nozzles at 100 psi on July 25 and Inspire (7 oz product in 19 gallons of water A⁻¹) on August 8.

Table 1. Application type, product names, active ingredients, and rates of fungicides used at planting in a field trial for control of *Rhizoctonia solani* AG 2-2 on sugarbeet. Standard rates of Apron + Thiram and 45 g/unit Tachigaren were on all seed. In-furrow fungicides were applied down the drip tube in a total volume of 6 gal/A.

Application	Product	Active ingredient	Rate ^z
None	-	-	-
Seed	Kabina ST	Penthiopyrad	14 g a.i./unit seed
Seed	Metlock Suite + Kabina ST	Metcon + Rizo + Penthio	0.21 + 0.5 + 7 g a.i./unit seed
Seed	Systiva	Fluxapyroxad	5 g a.i./unit seed
Seed	Vibrance	Sedaxane	1.5 g a.i./unit seed
In-furrow	AZteroid	Azoxystrobin	11.9 fl oz product A ⁻¹
In-furrow	Quadris	Azoxystrobin	10.0 fl oz product A ⁻¹
In-furrow	Xanthion	Pyraclostrobin + Bacillus amyloliquefaciens	9.0 fl oz product A ⁻¹ 1.8 fl oz product A ⁻¹

^z 11.9 fl oz AZteroid and 10 fl oz Quadris contain 0.15 and 0.16 lb azoxystrobin, respectively

Stand counts were done beginning 2 weeks after planting through 9 weeks after planting. The trial was harvested on October 5. Data were collected for number of harvested roots, yield, and quality. Twenty roots per plot also were arbitrarily selected and rated for severity of RCRR using a 0 to 7 scale (0 = healthy root, 7 = root completely rotted)

and foliage dead). Disease incidence was reported as the percent of rated roots with a root rot rating of > 2. Data were subjected to analysis of variance using SAS Proc GLM (SAS Institute, Cary, NC). Treatment means were separated using Fisher's protected least significant difference (LSD) test at a 0.05 level of significance.

RESULTS AND DISCUSSION

There were significant differences among treatments for plant stands at 2 through 9 weeks after planting (Fig. 1). At 2 weeks after planting, Systiva seed treatment had higher stand than the untreated control (Fig. 1). From 3 to 7 weeks after planting, all seed treatments resulted in significantly higher plant stand than the untreated control (Fig. 1). At 9 weeks after planting, only Metlock Suite + Kabina and Systiva were significantly higher in plant stand than the untreated control (Fig. 1). In-furrow fungicides resulted in stands similar to the untreated control throughout the first 9 weeks after planting (Fig. 1). For all stand counts, mean plant stand for seed treatments was significantly higher than the mean plant stand for in-furrow fungicides according to orthogonal contrasts ($P = 0.05$). It is not unusual for stand establishment to be reduced for in-furrow fungicides compared to seed treatments. Soil moisture and temperature were lower than normal at the NWROC during the period of emergence. Rainfall at the NWROC was just 0.94 inch during the month of May compared to a 30-year average of 3.04 inches for May. Average four-inch bare soil temperatures at the NWROC were 52.4 °F and 61.9 °F for the months of May and June, respectively. Average four-inch soil temperature did not cross 65 °F until July 4.

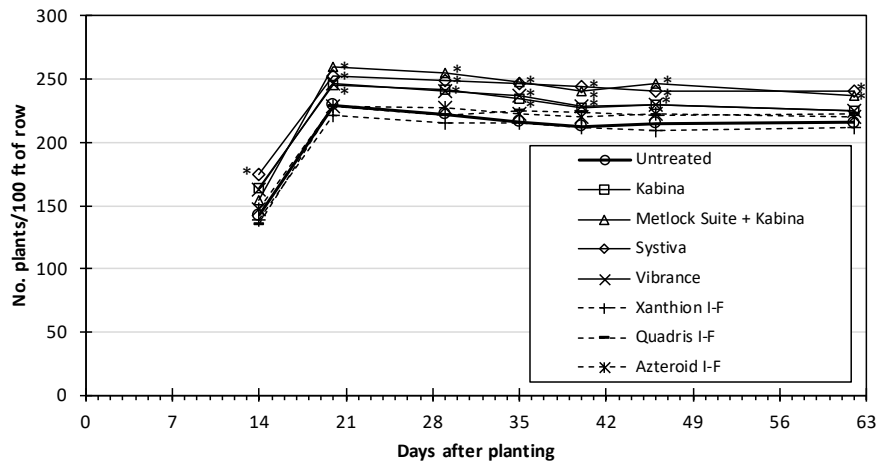


Fig. 1. Emergence and stand establishment for seed treatment (solid lines) and in-furrow (I-F, dotted lines) fungicides in a sugarbeet field trial infested with *Rhizoctonia solani* AG 2-2. For each stand count date, symbols marked with an asterisk represent stands significantly ($P = 0.05$) higher than the untreated control (bold solid line).

Table 2. Effects of at-planting (seed treatment or in-furrow) fungicide treatments on *Rhizoctonia* crown and root rot and sugarbeet yield and quality in a *Rhizoctonia*-infested field trial at the University of Minnesota, Northwest Research and Outreach Center, Crookston.

Sucrose^x

Treatment	No. harv. Roots/100 ft. ^x	RCRR (0-7) ^{xy}	RCRR % incidence ^{xz}	Yield ^x	%	lb ton ⁻¹	lb A ⁻¹
Untreated control	174	1.2	24	30.2	17.8	337	10170
Kabina ST	193	0.7	15	31.9	18.0	340	10844
Met. Suite + 7 g Kabina	200	1.3	25	31.3	17.7	333	10430
Systiva	205	1.1	21	33.9	18.0	339	11494
Vibrance	183	1.5	28	29.4	18.0	341	10063
AZteroid in-furrow	193	0.6	14	33.8	18.3	349	11767
Quadris in-furrow	191	0.9	15	31.7	17.8	337	10681
Xanthion in-furrow	189	0.8	15	31.9	18.1	342	10947
ANOVA P-value	0.2138	0.2437	0.3962	0.3233	0.8594	0.6769	0.2532
LSD (<i>P</i> = 0.05) ^x	NS	NS	NS	NS	NS	NS	NS
Contrast analysis							
Seed vs in-furrow							
Mean of Seed trts.	195	1.1	22	31.6	17.9	339	10708
Mean of In-furrow trts.	191	0.7	15	32.4	18.0	343	11132
P-value	0.4391	0.0706	0.0771	0.3635	0.5758	0.3726	0.2261

^x Values represent mean of 4 plots, NS = not significantly different

^y RCRR = Rhizoctonia crown and root rot; 0-7 scale, 0 = root clean, no disease, 7 = root completely rotted and plant dead

^z RCRR = Rhizoctonia crown and root rot; percent of roots with rating > 2

Soil moisture remained low throughout the growing season, resulting in low Rhizoctonia disease pressure in this trial. Total rainfall for the four months of May to August was 6.54 inches in 2017 compared to a 30-year average of 12.88 for the same four months. As a result, there were no significant differences among treatments for Rhizoctonia crown and root rot or yield and quality parameters (Table 2). Root rot ratings were low for all treatments with means ranging from 0.6 to 1.2 on the 0-7 scale (Table 2), reflecting the low disease pressure from *R. solani*. Disease incidence, reported as the percent of roots with a disease rating >2 ranged from 14 to 28% (Table 2). Root and sucrose yields were good for all treatments with root yields ranging from 29.4 to 33.8 ton A⁻¹ and sucrose ranging from 17.7 to 18.3%. Lack of significant differences at harvest in 2017 is in contrast with typical years with higher disease pressure, where in-furrow fungicides typically result in lower root rot ratings and higher yields at harvest compared to seed treatments (6-7).

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LITERATURE CITED

- Brantner, J.R. and A.K. Chanda. 2017. Plant pathology laboratory: summary of 2015-2016 field samples. 2016 Sugarbeet Res. Ext. Rept. 47:203-204.
- Brantner, J.R. 2015. Plant pathology laboratory: summary of 2013-2014 field samples. 2014 Sugarbeet Res. Ext. Rept. 45:138-139.
- Brantner, J.R. and A.K. Chanda. 2015. Integrated management of Rhizoctonia on sugarbeet with varietal resistance, seed treatment, and postemergence fungicides. 2014 Sugarbeet Res. Ext. Rept. 45: 142-146

17. Brantner, J.R. and C.E. Windels. 2011. Plant pathology laboratory: summary of 2009-2010 field samples. 2010 Sugarbeet Res. Ext. Rept. 41:260-261.
18. Brantner, J.R. and C.E. Windels. 2009. Plant pathology laboratory: summary of 2007-2008 field samples. 2008 Sugarbeet Res. Ext. Rept. 39:250-251.
19. Chanda, A.K. and J.R. Brantner. 2017. Evaluation of at-planting fungicide treatments for control of *Rhizoctonia solani* on sugarbeet. 2016 Sugarbeet Res. Ext. Rept. 47:166-168.
20. Chanda, A.K. and J.R. Brantner. 2016. Evaluation of at-planting fungicide treatments for control of *Rhizoctonia Solani*. 2015 Sugarbeet Res. Ext. Rept. 46:151-153.
21. Crane, E., Brantner, J.R., and Windels, C.E. 2013. Plant pathology laboratory: summary of 2011-2012 field samples. 2012 Sugarbeet Res. Ext. Rept. 43:169-170.
22. Niehaus, W.S. 2017. Results of American Crystal's 2016 official coded variety trials. 2016 Sugarbeet Res. Ext. Rept. 47:207-259.

USING POST FUNGICIDE APPLICATION AND SEED TREATMENTS FOR CONTROL OF RHIZOCTONIA SOLANI

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Rhizoctonia root and crown rot, caused by *Rhizoctonia solani* Kühn, is currently the most devastating soil borne disease of sugarbeet (*Beta vulgaris* L.) in North Dakota and Minnesota. In the bi-state area, *R. solani* anastomosis group (AG) 1, AG-2-2, AG-4 and AG-5 cause damping off and AG-2-2 causes root and crown rot of sugarbeet (Windels and Nabben 1989). *R. solani* survives as thickened hyphae and sclerotia in organic material and is endemic in soils where sugarbeet is grown. *R. solani* has a wide host range including broad leaf crops and weeds (Anderson 1982; Nelson et al. 2002). Crop rotations of three or more years with small grains planted before sugarbeet is recommended to reduce disease incidence (Windels and Lamey 1998). In fields with a history of high disease severity, growers may plant varieties that are more resistant but with significantly lower yield potential compared to more susceptible varieties (Panella and Ruppel 1996). Research showed that timely application of azoxystrobin provided effective disease control but not when applied after infection or after symptoms were observed (Brantner and Windels, 2002; Jacobsen et al. 2002). Fungicidal seed treatments were developed and commercialized starting in 2013 to provide early season protection from *R. solani*.

The objective of this research was to evaluate the fungicidal seed treatments with and without a post-application fungicide their effectiveness at controlling *R. solani* and impact on yield and quality in sugarbeet.

MATERIALS AND METHODS

A field trial was conducted at Hickson, ND in 2017. The site was inoculated on 28 April with *R. solani* AG 2-2 IIIB grown on barley. Inoculum was broadcast using a three-point mounted rotary/spinner type spreader calibrated to deliver 58 lbs/A of inoculum. The inoculum was incorporated with a Kongsilde field cultivator to about the two-inch depth before planting. The experimental design was a randomized complete block with four replicates. Field plots comprised of six 25-foot long rows spaced 22 inches apart. Plots were planted to stand on 3 May with a known susceptible variety. Seeds were treated with Tachigaren at 45 g/kg seed to provide early season protection against *Aphanomyces cochlioides*, and Poncho Beta to provide early season insect control. Counter 20G was also applied at 9 lb/A at planting to control insect pests. Weeds were controlled on 1 and 13 June 10 July. Fungicides were sprayed to control Cercospora leaf spot on 24 July and 2 August.

The fungicides treatments and rates of fungicide used are listed in Table 1. Different commercial seed treatments were used alone and with a post fungicide applied in a 7-inch band application. The band-applications were made on 12 June at the eight leaf stage using 17 gal of spray solution/A.

Stand counts were taken during the season and at harvest. The middle two-rows of plots were harvested on 11 September and weights were recorded. Samples (12-15 roots) from each plot, not including roots on the ends of plots, were analyzed for quality at American Crystal Sugar Company tare laboratory at East Grand Forks, MN. The data analysis was performed with the ANOVA procedure of the Agriculture Research Manager, version 8 software package (Gylling Data Management Inc., Brookings, South Dakota, 2010). The least significant difference (LSD) test was used to compare treatments when the F-test for treatments was significant.

RESULTS AND DISCUSSIONS

Dry conditions after planting resulted in delayed emergence. The first significant rainfall was 22 days after planting on May 25 and again on May 30. Plant stand was very variable in all treatments and counts taken on June 7 indicated variable stands but no significant differences among treatments. Seedling damping-off was not observed in June, probably because the dry conditions were not favorable for disease development. Rainfall on July 11 (2.84") and 19 (0.52") resulted in conditions more favorable for infection by *R. solani*. Typical symptoms of Rhizoctonia root rot including leaf wilting, yellowing, followed by death of leaves and then entire plants were observed starting in August. It should be noted that infection was not uniform in plots.

In the non-diseased conditions which prevailed early in the growing season, there were no significant differences in plant stand among seed treatments. At harvest, plant stand, although not statistically significantly, were lower in

treatments which had only fungicide seed treatment(s) or no seed treatment compared to treatments with a post fungicide application Quadris application at the 8-leaf stage. The environmental conditions and visual symptoms on infected plants indicated that there was some *R. solani* infection later in the growing season. It was likely that post application of Quadris provided some protection of plants from the later season Rhizoctonia root rot and the trend for higher plant stand, tonnage, sucrose concentration and recoverable sucrose. Overall dry conditions with favorable growing degree days along with adequate soil moisture resulted in relatively high tonnage, sucrose concentration and recoverable sucrose in all treatments, including the non-treated check. The benefits of using Quadris was best demonstrated where the fungicide was used only as a post application compared to the treatment using no fungicidal seed and post treatments (non-treated check). Since it is not known what environmental conditions will prevail during the growing season, and that none of the recommended fungicides are curative (that is, will not control *R. solani* after symptoms are observed), the prophylactic use of seed treatments and a post fungicide application when plants are at the 4- to 8-leaf stage should provide effective protection from *R. solani*.

References

- Anderson, N. A. 1982. The genetics and pathology of *Rhizoctonia solani*. Annu. Rev. Phytopathol. 20:329-347.
- Brantner, J. and Windels, C.E. 2002. Band and broadcast applications of quadris for control of Rhizoctonia root and crown rot on sugarbeet. In: 2001 Sugarbeet Res. Ext. Rep. Fargo, ND: NDSU Ext. Serv. 32:282-286.
- Jacobsen, B. J., Zidack, N. K., Mickelson, J. and Ansley, J. 2002. Integrated management strategies for Rhizoctonia crown and root rot. In: 2001 Sugarbeet Res. Ext. Rep. Fargo, ND: NDSU Ext. Serv. 32:293-295.
- Nelson, B., T. Helms, T. Christianson, and I. Kural. 1996. Characterization and pathogenicity of *Rhizoctonia solani* from soybean. Plant Dis. 80:74-80.
- Panella, L. and E. G. Ruppel. 1996. Availability of germplasm for resistance against *Rhizoctonia* spp. Pages 515-527. In: *Rhizoctonia* Species: Taxonomy, molecular biology, ecology, pathology and disease control. B. Sneh, S. Jabaji-Hare, S. Neate, and G. Dijat, eds. Kluwer Academic Publishers, Dordrecht, Netherlands.
- Windels, E. W. and H. A. Lamey. 1998. Identification and control of seedling diseases, root rot, and rhizomania on sugarbeet. Univ. Minnesota and North Dakota State Univ. Ext. Serv. Bull. PP-1142 , BU-7192-S.
- Windels, C. E., and D. J. Nabben. 1989. Characterization and pathogenicity of anastomosis groups of *Rhizoctonia solani* isolated from *Beta vulgaris*. Phytopathol. 79:83-88.

Table 1. Effect of seed treatments and post fungicide application for control of Rhizoctonia root rot on sugarbeet at Hickson, ND in 2017

Product and Rate in fl oz/A	19 June Stand Count	9 August Stand Count	11 September Stand Count	11 September Yield Ton/A	11 September Sucrose concentration %	11 September Recoverable sucrose lb/A
Untreated	207	188	144	30.7	17.7	9,799
Kabina 14g	207	193	149	32.3	17.2	9,832
Vibrance	214	199	148	32.0	17.5	10,041
Metlock + Rizolex + Kabina 7g	207	199	149	32.5	17.0	9,809
Quadris 9.2 fl oz	193	204	162	35.7	17.8	11,426
Kabina 14g fb	202	211	167	32.1	17.7	10,223
Quadris 9.2 fl oz						
Vibrance fb	209	205	164	32.3	17.7	10,332
Quadris 9.2 fl oz						
Metlock + Rizolex + Kabina 7g fb	215	229	171	34.9	17.4	10,922
Quadris 9.2 fl oz						
LSD (P=0.10)	12	20	NS	3.6	0.67	1095

*Treatment applied POST on 2 June.

SEED TREATMENT AND INFURROW FUNGICIDES FOR RHIZOCTONIA CONTROL

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Rhizoctonia root and crown rot, caused by *Rhizoctonia solani* Kühn, is currently the most devastating soil borne disease of sugarbeet (*Beta vulgaris* L.) in North Dakota and Minnesota. In the bi-state area, *R. solani* anastomosis group (AG) 1, AG-2-2, AG-4 and AG-5 cause damping off and AG-2-2 causes root and crown rot of sugarbeet (Windels and Nabben 1989). *R. solani* survives as thickened hyphae and sclerotia in organic material and is endemic in soils where sugarbeet is grown. *R. solani* has a wide host range including broad leaf crops and weeds (Anderson 1982; Nelson et al. 2002). Crop rotations of three or more years with small grains planted before sugarbeet is recommended to reduce disease incidence (Windels and Lamey 1998). In fields with a history of high disease severity, growers may plant varieties that are more resistant but with significantly lower yield potential compared to more susceptible varieties (Panella and Ruppel 1996). Research showed that timely application of azoxystrobin provided effective disease control but not when applied after infection or after symptoms were observed (Brantner and Windels, 2002; Jacobsen et al. 2002). Fungicidal seed treatments were developed and commercialized starting in 2013 to provide early season protection from *R. solani* and to facilitate the practice of using a liquid starter fertilizer at planting and speed-up the rate of planting. It will be useful to know whether seed treatments are compatible with in-furrow fungicides when needed for areas with high disease pressure, whether seed treatments provide season long disease protection, and whether multiple post-fungicide applications provide better disease control compared to one post-application at the 4-leaf stage.

The objective of this research was to determine whether seed treatments are compatible with in-furrow fungicides when needed for areas with high disease pressure, whether seed treatments provide season long disease protection, and whether multiple post-fungicide applications provide better disease control compared to one post-application at the 4-leaf stage.

MATERIALS AND METHODS

A field trial was conducted at Hickson, ND in 2017. The site was inoculated on 28 April with *R. solani* AG 2-2 IIIB grown on barley. Inoculum was broadcast using a three-point mounted rotary/spinner type spreader calibrated to deliver 58 lbs/A of inoculum. The inoculum was incorporated with a Kongsilde field cultivator to about the two-inch depth before planting. The experimental design was a randomized complete block with four replicates. Field plots comprised of six 25-foot long rows spaced 22 inches apart. Plots were planted to stand on 3 May with a Rhizoctonia susceptible variety. Seeds were treated with Tachigaren at 45 g/kg seed to provide early season protection against *Aphanomyces cochlioides*, and Poncho Beta. Counter 20G was also applied at 9 lb/A at planting to control insect pests. Weeds were controlled on 1 and 13 June and 10 July. Fungicides were sprayed to control Cercospora Leaf Spot on 24 July and 2 August.

The fungicides and rates used are listed in Table 1. Treatments were applied as an in-furrow application. The in-furrow applications were made on 3 May (at planting) using 7.1 gal of spray solution/A.

Stand counts were taken during the season and at harvest. The middle two-rows of plots were harvested on 11 September and weights were recorded. Samples (12-15 roots) from each plot, not including roots on the ends of plots, were analyzed for quality at American Crystal Sugar Company tare laboratory at East Grand Forks, MN. The data analysis was performed with the ANOVA procedure of the Agriculture Research Manager, version 8 software package (Gylling Data Management Inc., Brookings, South Dakota, 2010). The least significant difference (LSD) test was used to compare treatments when the F-test for treatments was significant.

RESULTS AND DISCUSSIONS

The first significant rainfall was 20 days after planting on May 25 and again on May 30. Emergence was non-uniform and occurred over a wide range of dates resulting in plant stand ranging from 158 to 182 on June 7 and 165 to 193 on June 23; however, there were no significant differences in plant stand among treatments on June 23 nor at harvest. It should be noted that dry conditions at and after planting were not favorable for infection and disease development by *R. solani* and Rhizoctonia damping-off was not observed. Later in the season, after mid-July, Rhizoctonia root rot

symptoms and death of plants in some treatments were observed. There were no significant differences in tonnage nor in sucrose concentration among treatments. There were significant differences in sugar loss to molasses which resulted in significant differences in recoverable sucrose among treatments. The seed treatments which had no post-fungicide applications all had lower tonnage compared to the same seed treatments with post-fungicide applications. Likewise, the check with no seed treatment also had lower tonnage than the non-treated seed with a post-fungicide application. Since *Rhizoctonia* root rot was observed later in the season, it is likely that the post fungicide applications provided better disease protection in those treatments leading to higher recoverable sucrose. In this trial, the seed treatments used alone did not result in as high recoverable sucrose per acre as seed treatments with post-application fungicides, or treatments with post-application fungicides. It was safe to use seed treatments with in-furrow fungicides. Based on the field data, it will be useful for growers to continue to use fungicide seed treatments to provide protection in years when conditions are favorable for *Rhizoctonia* damping-off. However, seed treatments do not provide season long protection against *R. solani*, so post-fungicide applications will still be necessary. In this trial, two post-fungicide applications (at the 4-6 and at the 8-10 leaf stages) resulted in the highest recoverable sucrose per acre. Research will continue to determine the best time and number of post fungicide applications for effective control of *R. solani* and highest recoverable sucrose

References

- Anderson, N. A. 1982. The genetics and pathology of *Rhizoctonia solani*. *Annu. Rev. Phytopathol.* 20:329-347.
- Brantner, J. and Windels, C.E. 2002. Band and broadcast applications of quadris for control of *Rhizoctonia* root and crown rot on sugarbeet. In: 2001 Sugarbeet Res. Ext. Rep. Fargo, ND: NDSU Ext. Serv. 32:282-286.
- Jacobsen, B. J., Zidack, N. K., Mickelson, J. and Ansley, J. 2002. Integrated management strategies for *Rhizoctonia* crown and root rot. In: 2001 Sugarbeet Res. Ext. Rep. Fargo, ND: NDSU Ext. Serv. 32:293-295.
- Nelson, B., T. Helms, T. Christianson, and I. Kural. 1996. Characterization and pathogenicity of *Rhizoctonia solani* from soybean. *Plant Dis.* 80:74-80.
- Panella, L. and E. G. Ruppel. 1996. Availability of germplasm for resistance against *Rhizoctonia* spp. Pages 515-527, In: *Rhizoctonia* Species: Taxonomy, molecular biology, ecology, pathology and disease control. B. Sneh, S. Jabaji-Hare, S. Neate, and G. Dijat, eds. Kluwer Academic Publishers, Dordrecht, Netherlands.
- Windels, E. W. and H. A. Lamey. 1998. Identification and control of seedling diseases, root rot, and rhizomania on sugarbeet. *Univ. Minnesota and North Dakota State Univ. Ext. Serv. Bull.* PP-1142 , BU-7192-S.
- Windels, C. E., and D. J. Nabben. 1989. Characterization and pathogenicity of anastomosis groups of *Rhizoctonia solani* isolated from *Beta vulgaris*. *Phytopathol.* 79:83-88.

Table 1. Effect of fungicides from in-furrow applications and seed treatments on Rhizoctonia root rot at Hickson, ND in 2017

Product and Rate in fl oz/A	Application dates	12 June Stand Count	11 Sept Stand Count	11 Sept Yield	11 Sept Sucrose concentration	11 Sept SLM	11 Sept Recoverable sucrose
		beets/100'	beets/100'	Ton/A	%	%	lb/A
Untreated	-	205	179	31.0	17.7	1.66	9,871
Kabina	Seed trt	200	162	32.6	17.6	1.70	10,385
Vibrance	Seed trt	210	164	31.3	17.9	1.61	10,205
Metlock + Rizolex + Kabina	Seed trt	214	174	33.5	17.9	1.57	10,920
Systiva	Seed trt	202	175	30.5	18.0	1.65	9,947
Kabina/ Quadris 9.2 fl oz	Seed trt/ 12 June	197	195	31.1	18.3	1.61	10,357
Vibrance/ Quadris 9.2 fl oz	Seed trt/ 12 June	212	166	32.2	17.7	1.59	10,333
Metlock + Rizolex + Kabina/ Quadris 9.2 fl oz	Seed trt/ 12 June	206	190	32.0	17.9	1.65	10,349
Systiva/ Quadris 9.2 fl oz	Seed trt/ 12 June	211	165	33.5	17.9	1.60	10,947
Kabina/ Quadris 9.2 fl oz/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 12 June/ 20 June	212	189	33.1	17.6	1.58	10,614
Vibrance/ Quadris 9.2 fl oz/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 12 June/ 20 June	216	193	31.8	18.1	1.62	10,476
Metlock + Rizolex + Kabina/ Quadris 9.2 fl oz/ Proline 5.6 fl oz + NIS 0.125 % v/v	Seed trt/ 12 June/ 20 June	216	189	34.5	17.7	1.67	11,020
Systiva/ Quadris 9.2 fl oz/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 12 June/ 20 June	216	192	32.2	18.1	1.59	10,578
Quadris 9.2 fl oz	12 June	207	173	31.5	17.9	1.66	10,192
Quadris 9.2 fl oz/ Proline 5.6 fl oz + NIS 0.125% v/v	12 June/ 20 June	212	164	31.7	18.2	1.55	10,538
Quadris 9.2 fl oz IF	3 May	218	193	33.4	17.9	1.73	10,749
Quadris 9.2 fl oz IF/ Proline 5.7 fl oz + 0.125% v/v	3 May/ 12 June	217	184	32.3	17.9	1.59	10,524
Quadris 9.2 fl oz IF/ Proline 5.7 fl oz + 0.125% v/v/ Priaxor 6.7 fl oz	3 May/ 12 June/ 20 June	204	168	35.3	17.6	1.69	11,215
Kabina + Quadris 9.2 fl oz IF	Seed trt/ 3 May	209	161	30.4	17.6	1.59	9,778
Vibrance + Quadris 9.2 fl oz IF	Seed trt/ 3 May	195	179	31.9	17.8	1.66	10,223
Metlock + Rizolex + Kabina + Quadris 9.2 fl oz IF	Seed trt/ 3 May	199	167	27.8	18.2	1.64	9,172
Systiva + Quadris 9.2 fl oz	Seed trt/ 3 May	213	175	32.1	18.3	1.66	10,679
Kabina + Quadris 9.2 fl oz IF/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 3 May/ 12 June	205	187	30.0	18.5	1.58	10,126
Vibrance + Quadris 9.2 fl oz IF/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 3 May/ 12 June	181	170	33.0	18.2	1.55	10,993

Metlock + Rizolex + Kabina + Quadris 9.2 fl oz IF/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 3 May/ 12 June	198	163	32.2	18.3	1.63	10,682
Systiva + Quadris 9.2 fl oz/ Proline 5.7 fl oz + NIS 0.125% v/v	Seed trt/ 3 May/ 12 June	207	169	32.4	17.9	1.68	10,419
LSD (P=0.10)	-	15	NS	NS	NS	NS	NS

SENSITIVITY OF *CERCOSPORA BETICOLA* TO FOLIAR FUNGICIDES IN 2017

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Leaf spot, caused by the fungus *Cercospora beticola*, is an endemic disease of sugarbeet produced in the Northern Great Plains area of North Dakota and Minnesota that reduces both yield and sucrose content. The disease is controlled by crop rotation, resistant varieties and timely fungicide applications. *Cercospora* leaf spot usually appears in the last half of the growing season, and multiple fungicide applications are necessary for disease management. Fungicides are used at high label rates and are alternated for best efficacy, but in recent years, mixtures are becoming more common. The most frequently used fungicides are Tin (triphenyl tin hydroxide), Topsin (thiophanate methyl), Eminent (tetraconazole), Proline (prothioconazole), Inspire (difenoconazole) and Headline (pyraclostrobin). In 2017, most of the DMI and QoI fungicides were applied as mixtures with either mancozeb or copper and Topsin is usually applied as a tank mix with Tin.

Like many other fungi, *C. beticola* has the ability to become less sensitive (resistant) to the fungicides used to control them after repeated exposure, and increased disease losses can result. Because both *C. beticola* and the fungicides used for management have histories of fungicide resistance in our production areas and other production areas in the US, Europe and Chile, it is important to monitor our *C. beticola* population for changes in sensitivity to the fungicides in order to achieve maximum disease control. We have monitored fungicide sensitivity of field isolates of *C. beticola* collected from fields representing the sugarbeet production area of the Red River Valley region to the commonly used fungicides in our area annually since 2003. In 2017, extensive sensitivity monitoring was conducted for Tin, Eminent, Inspire, Proline and Headline.

OBJECTIVES

- 1) Monitor changes in sensitivity of *Cercospora beticola* isolates to Tin (fentin hydroxide)
- 2) Monitor changes in sensitivity of *Cercospora beticola* isolates to Topsin (thiophanate methyl)
- 3) Monitor changes in sensitivity of *Cercospora beticola* to three triazole (DMI) fungicides: Eminent (tetraconazole) and Inspire (difenoconazole) and Proline (prothioconazole)
- 4) Test *Cercospora beticola* isolates for the presence of the G143A mutation that confers resistance to Headline (pyraclostrobin) fungicide
- 5) Distribute results of sensitivity monitoring in a timely manner to the sugarbeet industry in order to make fungicide recommendations for disease management and fungicide resistance management for *Cercospora* leaf spot disease in our region.

METHODS AND MATERIALS

In 2017, with financial support of the Sugarbeet Research and Extension Board of MN and ND, we tested 1105 *C. beticola* field isolates collected from throughout the sugarbeet production regions of ND and MN for sensitivity testing to Tin, Eminent, Inspire, Proline and Headline. For this report we use the commercial name of the fungicides, but all testing was conducted using the technical grade active ingredient of each fungicide, not the formulated commercial fungicide. The term µg/ml is equivalent to ppm.

Sugarbeet leaves with *Cercospora* leaf spot (CLS) were collected from commercial sugarbeet fields by agronomists from American Crystal Sugar Company, Minn-Dak Farmers Cooperative and Southern Minnesota Beet Sugar Cooperative representing all production areas in ND and MN. Leaves were delivered to our lab, and processed quickly to insure viability of spores. From each field sample, *C. beticola* spores were collected from a minimum of five spots per leaf from five leaves and mixed to make a composite of approximately 2500 spores. A subsample of the spore composite was transferred to a Petri plates containing water agar amended with Tin at 1 µg/ml. Germination of 100 spores on Tin amended water agar plates were counted 16 hours later and percent germination calculated. Germinated spores are considered resistant. Sensitivity to Topsin is tested alternate years and was not tested in 2017.

For triazole fungicide sensitivity testing, a radial growth procedure is used. A single spore subculture from the spore composite was grown on water agar medium amended with serial ten-fold dilutions of each technical grade triazole fungicide from 0.01 – 10.0 ppm. A separate test was conducted for each triazole fungicide. After 15 days, inhibition of radial growth was measured, and compared to the growth of *C. beticola* on non-amended water agar medium. This data was used to calculate an EC₅₀ value for each isolate; EC₅₀ is a standardized method of measuring fungicide resistance and is calculated by comparing the concentration of fungicide that reduces radial growth of *C. beticola* by 50% compared to the growth on non-amended media. Higher EC₅₀ values mean reduced sensitivity to the fungicide. An RF (resistance factor) was calculated by dividing the EC₅₀ value by the baseline value so fungicides can be directly compared.

For Headline resistance testing we use a PCR based molecular procedure to test for the presence of a specific mutation in *C. beticola* that imparts resistance to Headline. This procedure detects a specific mutation, G143A, which results in complete resistance to Headline. DNA is extracted from the remaining spore composite and tested by real-time PCR using primers specific for the G143A mutation. The test enables us to estimate the percentage of spores with the G143A mutation in each sample. Each sample tested contains approximately 2500-5000 spores and the DNA from this spore pool will test for the G143A mutation from each spore. The spore germination test we previously used only tested one spore per five spot/five leaf sample. The PCR test is also more sensitive and requires less interpretation than the previously used spore germination test. The PCR test will estimate the incidence of resistance in the population of spores tested, and give a better indication of Headline resistance in a field.

RESULTS AND DISCUSSION

CLS in 2017 was common but much less severe than in 2016, and CLS in general was well managed by the fungicide programs that were used. The majority of the CLS samples were delivered to our lab at the end of the season in late September and early October. Almost all samples arrived in excellent condition and delivered as fresh samples. Field samples (n=1105) representing all production areas and factory districts were tested for sensitivity to five fungicides: tin, tetraconazole (Eminent), difenoconazole (the most active part of Inspire), prothioconazole (Proline) and pyraclostrobin (Headline). Three additional DMI, one SDHI and one QoI fungicides not registered in the US for CLS were tested for activity against *C. beticola* in lab trials. One new DMI and one new QoI appeared to have good activity against *C. beticola*.

TIN. Tolerance (resistance) to Tin was first reported in 1994 at concentrations of 1-2 µg/ml. At these levels, disease control in the field is reduced. The incidence of fields with isolates resistant to Tin at 1.0 µg/ml increased between 1997 and 1999, but the incidence of fields with resistant isolates has been declining since the introduction of additional fungicides for resistance management, including Eminent in 1999, Gem in 2002 and Headline in 2003. In 1998, the percentage of fields with isolates resistant to Tin at 1.0 µg/ml was 64.6%, and declined to less than 10% from 2002 to 2010. From 2011 to 2014 there was an increase in the number of fields with resistance (**Figure 1**), and from 2015 to 2017, the incidence of fields with isolates resistant to Tin increased from 38.5% to 97% (**Figure 1**). The severity of resistance (as expressed as germination rate of spores from fields with resistant isolates) ranged from 1 to 100%, with the average germination rate ranging from 16 to 28% during the five year period of 2012 to 2017 (**Figure 2**). The incidence of fields with tin resistance increased in all factory districts, with the lowest incidence in the Drayton, East Grand Forks and Hillsboro factory districts (**Figure 3**). The low severity of resistance (<30%) may be the reason that tin is still an effective fungicide for managing CLS despite widespread incidence of resistance to tin.

TOPSIN. Resistance to Topsin has been present in our area since 1999, and is also common and widespread in European Union production areas. Resistance has historically been >70% but has declined below that level in six of the past twelve years. Topsin resistance, in sugarbeet and other crops, tends to decline when it is not used, but reappears quickly when it is again used in the field. In 2014, the percentage of fields with isolates resistant to Topsin at 5 µg/ml was 73.5% and in 2016 increased to 86.0% (data not shown). The incidence of resistance as measured by germination rate of spores from fields with resistant isolates ranged from 1 to 100%, with the average germination rate of 25%. Most applications of Topsin are as tank mixtures with Tin, which seems to be an effective management practice. Sensitivity to Topsin is measured in alternate years and was not tested in 2017.

DMI (triazoles). Sensitivity of *C. beticola* isolates to the DMI fungicides Eminent and Inspire, as measured by Resistance Factor values (RF), only doubled from 2007 to 2010, with average RF values <3 (RF values are the calculated EC₅₀ values divided by the baseline values). From 2011 to 2014, RF values of both Eminent and Inspire

increased to 54.5 and 68.3 respectively (**Figure 4**). Surprisingly, in 2015 there was a 29% and 69% decline in RF values to Eminent and Inspire respectively across all factory districts to average RF values of 39.0 and 21 (**Figure 4**). In 2016, the RF value of Eminent declined slightly and increased slightly for Inspire across all factory districts (**Figure 4**). In 2017, RF values for both Eminent and Inspire increased (**Figure 4**), ranging from 27.1 in the Moorhead district to 57.0 in the Hillsboro district (**Figure 5**).

The RF values of *C. beticola* isolates to Proline from 2016 to 2017 were 6.5 and 9.1 respectively, much lower than either Eminent or Inspire RF values (**Figure 4**), and was observed in every factory district (**Figure 5**). Proline has become more widely used for managing CLS in recent years.

The resistance to the triazole fungicides we see in US isolates of *C. beticola* is due to overexpression of Cyp51 enzyme, and not due to a specific genetic mutation, so it will be difficult to develop a PCR assay for this group of fungicides. In companion studies we have conducted, higher levels of resistance to triazole fungicides are present in *C. beticola* isolates collected from Italy and France than found in the RRV production area. We do not know if the reduction in RF values indicates a fitness penalty or not, but it will continue to be important to monitor resistance to triazole fungicides in the RRV region due to their widespread use. We are testing other DMI fungicides in our lab for their activity against *C. beticola*, but unfortunately, most of them are not registered for CLS management.

HEADLINE. Beginning in 2012, a PCR based molecular procedure was used to test for the presence of the G143A mutation in *C. beticola* using the remainder of the composite spore sample containing approximately 2500-5000 spores. The presence of this mutation indicates absolute resistance to Headline. The results are placed in five categories based on an estimate of the percentage of spores with the G143A mutation: S = no spores with G143A; S/r = <50 of the spores with G143A; S/R = equal number of spores with G143A; R/s >50% of the spores with G143A; and R = all spores with G143A. The G143A mutation was first detected in the RRV production area in 2012 and incidence of this mutation has increased in the population of isolates we test every year since then. Resistance to Headline in 2017 was similar to 2016. Across all factory districts in 2017, 10.9% of the isolates collected had all spores without the G143A mutation; the G143A mutation was found in 89.1% of the samples, and 64.2% of the samples had >50 of the spores with the G143A mutation (**Figure 6**). Samples with an R rating (all spores resistant) increased from 40.0% to 55.8% (**Figure 6**). Resistance (R) was detected in all factory districts ranging 45.6% in the East Grand Forks district to 70.3% in the Moorhead district (**Figure 7**). Samples with S (all spores sensitive) ranged from 3.0% in the SMBSC district to 10.9% in the Moorhead district. Based on this data, the QoI fungicides Headline and Gem will likely not control CLS and will not be widely used in the near future. Although this is a stable mutation, we will continue to partially monitor for resistance to Headline in the RRV production area, particularly because Headline is often the only fungicide used, and is used annually even in the absence of disease. We do not know if there is a fitness penalty associated with the G143A mutation, but based on observation in MI and Italy, Austria and Serbia, where QoI resistance due to the G143A mutation is widespread, it appears that isolates with the G143A mutation are stable and can survive and increase in the population.

An increasing concern is the development of *C. beticola* isolates with resistance (reduced sensitivity) to more than one fungicide. Of the isolates tested in 2017:

- 25.9% were resistant to Eminent > 1ppm
- 47.1% were resistant to Inspire > 1ppm
- 97% were resistant to tin >1 ppm
- 89.1% were resistant to Headline
- 27.7% were resistant to tin plus a DMI
- 14.0% were resistant to tin plus Headline plus a DMI

In 2016, 14.4% of the isolates tested were resistant to tin plus a DMI plus Headline.

Previously we conducted a greenhouse trial to determine if isolates of *C. beticola* with high levels of resistance results in decreased disease control by field application rates of Eminent compared with isolates sensitive to Eminent. Results of this work showed that the break point for causing more disease was the EC₅₀ value of >1 µg/ml. At this value, there was significantly more disease when the field rate of Eminent was used. This trial was conducted using a CLS susceptible variety. We repeated this study using a CLS resistant variety to see if the break point results were the same or not. The break point for disease loss for a CLS resistant variety increased to the EC₅₀ value of 10

µg/ml. After this level of resistance, there was a significant loss in disease control. This study suggests that variety resistance increases the level of *C. beticola* isolated resistance necessary for disease loss five-fold. A solid recommendation for CLS management will be to use varieties with good CLS resistance, and to find higher levels of resistance in future years. The use of varieties with increased levels of resistance will be important to manage CLs in future years and breeding for CLS resistance should be encouraged. Differences in aggressiveness among isolates may account for inconsistency of data and should be considered during resistance breeding. Measuring disease loss due to fungicide resistance is difficult, and additional work is necessary to confirm and document the results of these preliminary trials with CLS and Eminent resistant to *C. beticola*.

SUMMARY

1. Resistance to Tin at 1.0 µg/ml almost disappeared in our region from 2003-2010, but has increased since 2011, probably due to increased use. In 2017, isolates from 97% of the fields samples had some resistance to tin (incidence), with a mean germination rate of 28% (severity). Tin resistance was found in all factory districts.
2. Topsin was not tested in 2017.
3. Resistance to both Eminent and Inspire, as measured by RF values, increase slightly in 2017 in all factory districts. The RF values for Proline were much lower than either Eminent or Inspire.
4. The incidence of isolates with the G143A mutation that results in resistance to Headline remained about the same in 2017 as it was in 2016 across all factory districts. Approximately 90% of the fields sampled have some level of resistance to Headline, and approximately 50% of the fields sampled have >50% of the spores resistant to Headline. These findings may limit the effective use of Headline for CLS management in future years.
5. The incidence of *C. beticola* isolates with resistance to multiple fungicides is a concern. About 14 % of our isolates have resistance to five fungicides.
6. *C. beticola* isolates with resistance caused more disease (leaf spots) than sensitive plants treated with Eminent at the field rate in greenhouse trials, and isolates with resistance can cause as much or disease than the sensitive isolates in plants not treated with Eminent. There is a difference between CLS susceptible and resistant varieties disease loss based on isolate resistance to Eminent. The EC₅₀ value break point for significant disease loss for a susceptible variety is 1.0 µg/ml for the susceptible varieties compared to a break point of 10.0 µg/ml for a resistant variety
7. We recommend continuing disease control recommendations currently in place including fungicide rotation, using high label rate of fungicides, mixtures with mancozeb or copper, scouting at end of the season to decide the necessity of a late application, using fungicide resistance maps for fungicide selection, using a resistant variety, spray intervals of 14 days, and applying fungicides to insure maximum coverage. It appears that early fungicide applications in 2017 helped manage CLS and early applications should continue in 2018. Improved disease control may be possible with improvements in fungicide coverage using proper spray nozzles and spray parameters such as ground speed, timing and gallonage.

Figure 1. Incidence of fields with *C. beticola* resistant to Tin at 1.0 µg/ml as measured by spore germination collected in ND and MN from 1998 to 2017

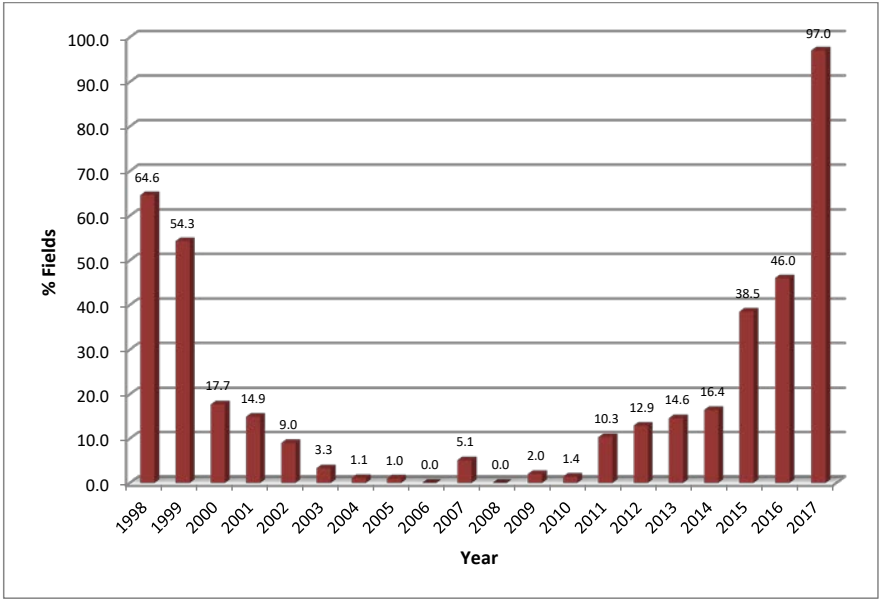


Figure 2. Incidence and severity of Tin resistance in *C. beticola* isolates collected from sugarbeet fields in ND and MN from 2003 to 2017

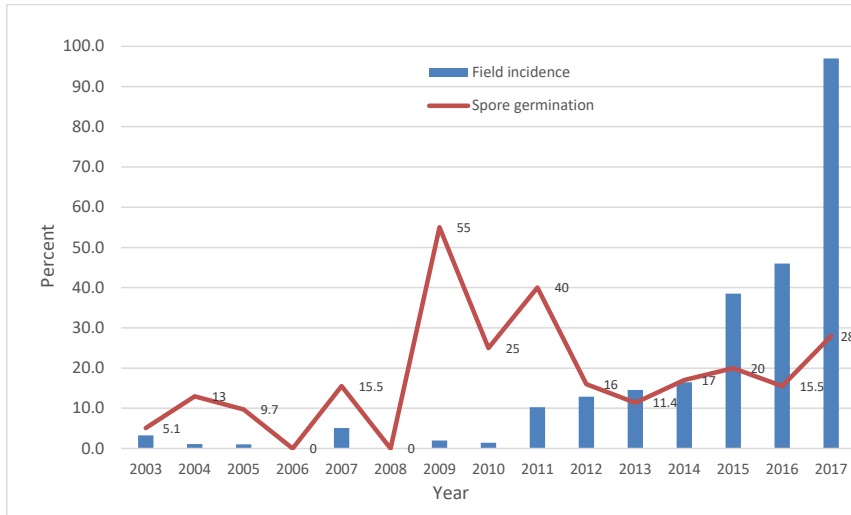


Figure 3. Incidence of fields with *C. beticola* isolates collected in ND and MN resistant to Tin from 2013 to 2017 by factory district

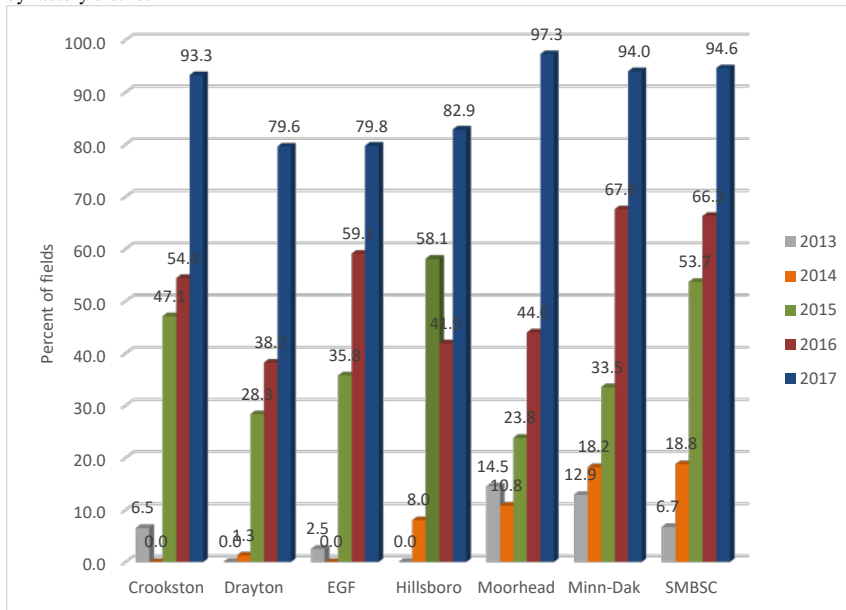


Figure 4. Resistance Factor values of *C. beticola* isolates collected in ND and MN from 2007-2017 to Eminent (tetraconazole), Inspire (difenoconazole) and Proline (prothioconazole)

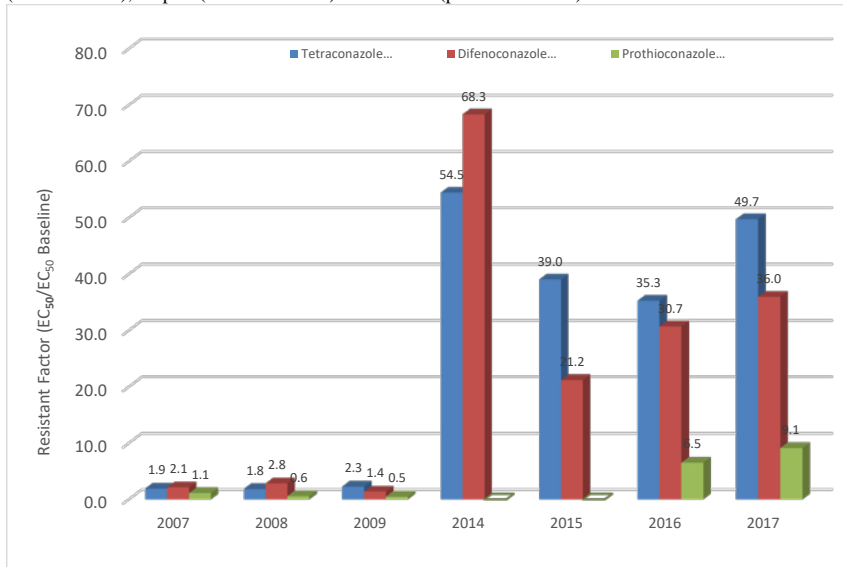


Figure 5. Sensitivity of *C. beticola* isolates collected in 2017 to Eminent, Inspire and Proline by factory district as expressed by Resistance Factor values

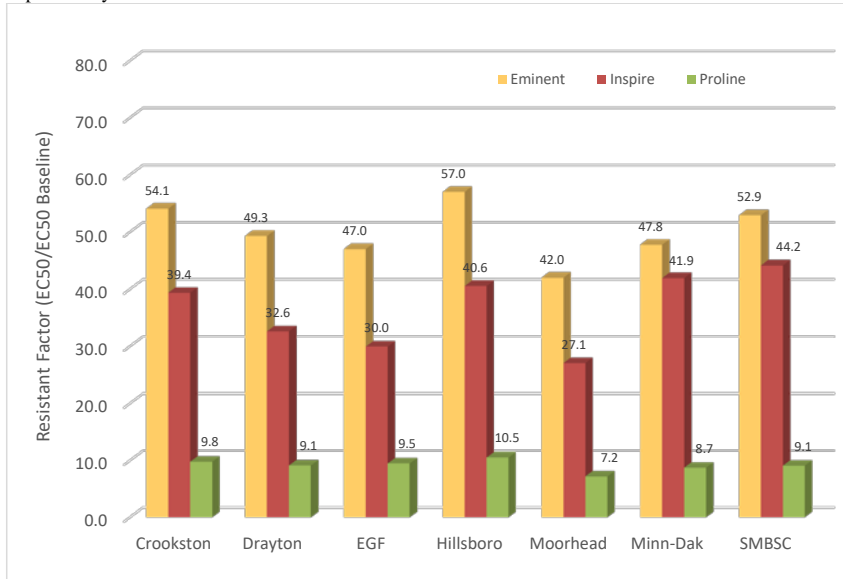


Figure 6. Sensitivity of *C. beticola* isolates collected in ND and MN to Headline from 2012 to 2017 as expressed by the percentage of spores with G143A mutation

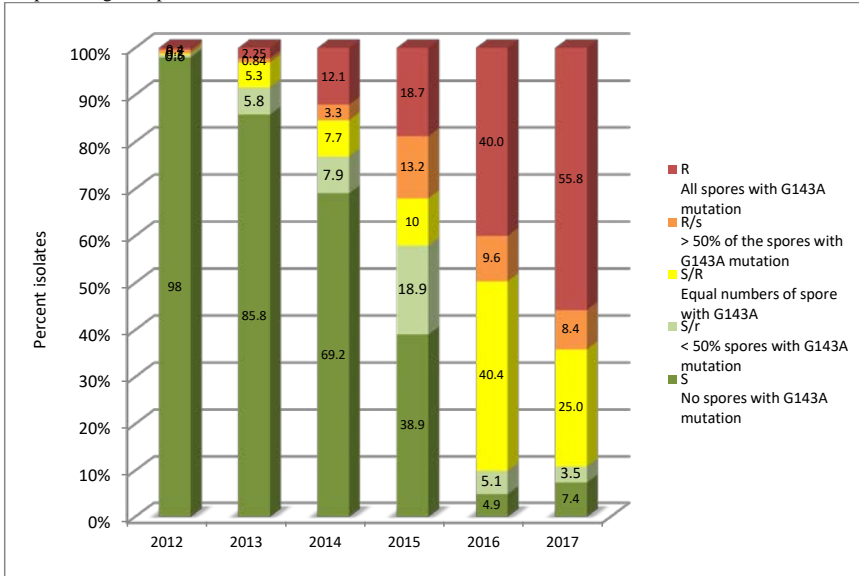
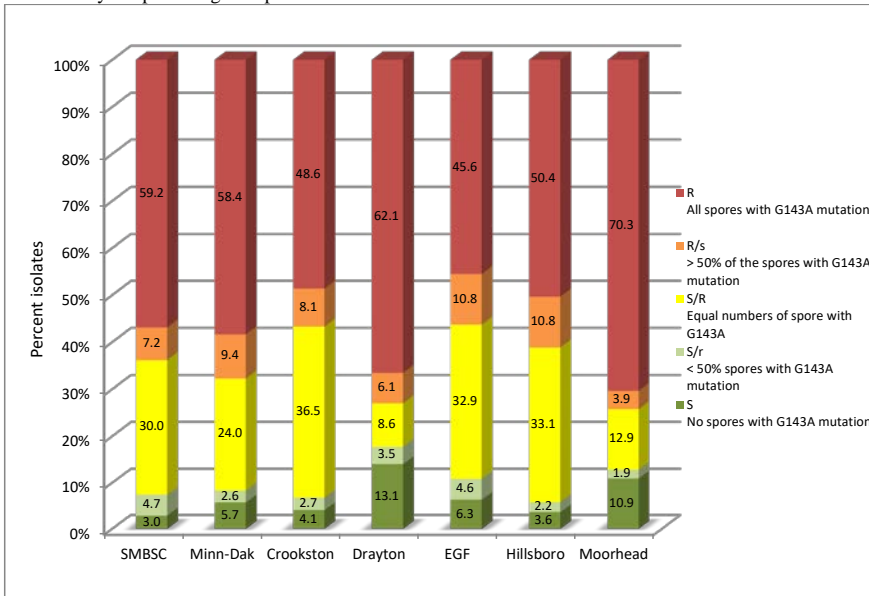


Figure 7. Sensitivity of *C. beticola* isolates collected in ND and MN in 2016 to Headline by factory district as measured by the percentage of spores with G143A mutation



EFFICACY OF FUNGICIDES FOR CONTROLLING CERCOSPORA LEAF SPOT ON SUGARBEET

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Cercospora leaf spot (CLS), caused by the fungus *Cercospora beticola* Sacc., is the most economically damaging foliar disease of sugarbeet in Minnesota and North Dakota. The disease reduces root yield and sucrose concentration and increases impurity concentrations resulting in reduced extractable sucrose and higher processing losses (Smith and Ruppel, 1973; Khan and Smith, 2005). Roots of diseased plants do not store well in storage piles that are processed in a 7 to 9 month period in North Dakota and Minnesota (Smith and Ruppel, 1973). Cercospora leaf spot is managed by integrating the use of tolerant varieties, reducing inoculum by crop rotation and tillage, and fungicide applications (Khan et al; 2007). It is difficult to combine high levels of Cercospora leaf spot resistance with high recoverable sucrose in sugarbeet (Smith and Campbell, 1996). Consequently, commercial varieties generally have only moderate levels of resistance and require fungicide applications to obtain acceptable levels of protection against Cercospora leaf spot (Miller et al., 1994) under moderate and high disease severity.

The objective of this research was to evaluate the efficacy of fungicides used in rotation to control Cercospora leaf spot on sugarbeet.

MATERIALS AND METHODS

A field trial was conducted at Foxhome, MN in 2017. The experimental design was a randomized complete block with four replicates. Field plots comprised of six 30-foot long rows spaced 22 inches apart. Plots were planted on 5 May with a variety susceptible to Cercospora Leaf Spot. Seeds were treated with Tachigaren (45 g/kg seed), Kabina 14g and Nipsit Inside. Seed spacing within the row was 4.7 inches. Weeds were controlled with two herbicide applications on 1 June and 19 June. Quadris was applied on 24 May and 6 June to control *Rhizoctonia solani*. Plots were inoculated on 29 June with *C. beticola* inoculum.

Fungicide spray treatments were applied with a CO₂ pressurized 4-nozzle boom sprayer with 11002 TT TwinJet nozzles calibrated to deliver 17 gpa of solution at 60 p.s.i pressure to the middle four rows of plots. All fungicide treatments were initiated on 19 July. Most treatments included four fungicide applications on 19 July, 31 July, 21 August and 6 September. One treatment received applications on a shorter interval and had application dates of 19 July, 31 July, 7 August, 21 August and 6 September. Treatments were applied at rates indicated in Table 1.

Cercospora leaf spot severity was rated on the leaf spot assessment scale of 1 to 10 (Jones and Windels, 1991). A rating of 1 indicated the presence of 1- 5 spots/leaf or 0.1% disease severity and a rating of 10 indicated 50% or higher disease severity. Cercospora leaf spot severity was assessed five times during the season. The rating performed on 16 September is reported.

Plots were defoliated mechanically and harvested using a mechanical harvester on 20 September. The middle two rows of each plot were harvested and weighed for root yield. Twelve to 15 representative roots from each plot, not including roots on the ends of the plot, were analyzed for quality at the American Crystal Sugar Company Quality Tare Laboratory, Moorhead, MN. The data analysis was performed with the ANOVA procedure of the Agriculture Research Manager, version 8 software package (Gylling Data Management Inc., Brookings, South Dakota, 2010). The least significant difference (LSD) test was used to compare treatments when the F-test for treatments was significant.

RESULTS AND DISCUSSIONS

Environmental conditions (especially moisture in the form of rainfall) were not favorable for rapid development of *C. beticola* after inoculation on 29 June and first symptoms at very low incidence were not visible until mid-July. On 11 August, CLS rating for the non-treated check was 4.2, still below the CLS rating (6.0) at which economic losses typically occur. Rainfall events during the week of 13 through 19 of August resulted in favorable conditions for rapid disease development as indicated by a CLS rating of 9.3 for the non-treated check on 24 August, followed by loss of mature leaves and re-growth of new leaves in the first week of September.

The CLS population was resistant to QoI fungicides and had the G143A mutation. CLS was effectively controlled when mixtures with different modes of action used individually at full or ¾ the recommended rates were used, and

when applications were made at 14 day and 10 to 12 day intervals. It was not possible to apply treatments scheduled for 14 July because of wet field conditions, resulting to a later application date on 21 August. The non-treated check had significantly higher CLS ratings compared to the fungicide treatments (Table 1). The fungicide treatments provided effective control of CLS which resulted in significantly higher sugar concentration, recoverable sucrose per acre, and recoverable sucrose per ton of sugarbeet compared to the non-treated check.

This research indicated that fungicides should be applied starting promptly at first symptoms of CLS and continued during the season once environmental conditions are favorable for disease development since our field have a high pathogen population. Each application should comprise of at least two modes of action, and when necessary such as during periods of regular rainfall, spray interval should be reduced from 14 to 10 to 12 days. In this trial, fungicide application was discontinued in early September to facilitate harvesting in mid- to late-September.

General comments for Cercospora leaf spot control in growers' fields in North Dakota and Minnesota where inoculum levels will probably be high in 2018 and CLS tolerant (KWS ratings of 5.2 and less) varieties are grown:

1. The first fungicide application should be made when disease symptoms are first observed (which entails scouting after row closure). If the first application is late, control will be difficult all season.
2. Subsequent applications should be made when symptoms are present and environmental conditions (2 consecutive days DIV obtained at <http://ndawn.ndsu.nodak.edu>) are favorable ($DIV \geq 7$) for disease development.
3. Use mixtures of fungicides that are effective at controlling Cercospora leaf spot in an alternation program.
4. Use the recommended rates of fungicides to control Cercospora leaf spot.
5. During periods of regular rainfall, shorten application interval from 14 days to 10 to 12 days; use aerial applicators during periods when wet field conditions prevent the use of ground rigs.
6. Limit or avoid using fungicides to which the pathogen population has become resistant or less sensitive.
7. Only one application of a benzimidazole fungicide (such as Topsin M 4.5F) in combination with a protectant fungicide (such as SuperTin). The use of TPTH mixed with a QoI or DMI fungicides will increase the effectiveness of the QoIs and DMIs.
8. Limiting the use of Qoi's (strobilurins) to one application for control of QoI sensitive populations of *C. beticola* will prolong the effectiveness of these fungicides. Limit the total number of DMI fungicides to 50% or less of the total number of fungicide applications in a season for CLS.
9. Use high volumes of water (15 to 20 gpa for ground-rigs and 3 to 5 gpa for aerial application) with fungicides for effective disease control.
10. Mix, mix, Mix! Try to alternate mixtures with different modes of action for controlling CLS and managing fungicide resistance.

The following fungicides in several classes of chemistry are registered for use in sugarbeet:

<u>Strobilurins</u>	<u>Sterol Inhibitors</u>	<u>Ethylenebisdithiocarbamate (EBDC)</u>
Headline/Pyrac	Eminent/Minerva	Penncozeb
Gem	Inspire XT	Manzate
Quadris	Proline	Mancozeb
Priaxor	Minerva Duo	Maneb
	Enable	
	Topguard	
<u>Benzimidazole</u>	<u>TriphenylTin Hydroxide (TPTH)</u>	<u>Copper</u>
Topsin	SuperTin	Kocide
	AgriTin	Badge
		Champion

Table 1. Effect of fungicides on Cercospora leaf spot control and sugarbeet yield and quality at Foxhome, MN in 2017.

Treatment and rate/A	CLS*	Root yield Ton/A	Sucrose concentration %	Recoverable sucrose lb/Ton lb/A		Returns** \$/A
Inspire XT 5.3 fl oz + Topsin 7.6 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt/ Minerva Duo 16 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt/ Proline 3.8 fl oz + NIS 0.125 %v/v + Manzate 1.2 qt***	4.3	36.48	17.63	331.6	12,085	1,449.47
Minerva Duo 16 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt/ Priaxor 6 fl oz + Manzate 1.2 qt/ Inspire XT 5.3 fl oz + Manzate 1.2 qt	4.8	34.93	18.05	338.2	11,774	1,448.65
Inspire XT 7 fl oz + Topsin 10 fl oz/ Super Tin 8 fl oz + Manzate 1.6 qt/ Minerva Duo 16 fl oz/ Super Tin 8 fl oz + Manzate 1.6 qt	4.8	34.38	17.53	329.2	11,309	1,349.46
Inspire XT 5.3 fl oz + Manzate 1.2 qt/ Super Tin 6 fl oz + Manzate 1.2 qt/ Minerva Duo 16 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt	5.0	34.68	17.38	325.5	11,271	1,338.25
Inspire XT 5.3 fl oz + Super Tin 6 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt/ Priaxor 6 fl oz + Manzate 1.2 qt/ Minerva Duo 12 fl oz + Manzate 1.2 qt	4.8	33.00	17.65	331.5	10,923	1,305.95
Manzate 1.2 qt + Topsin 7.6 fl oz/ Inspire XT 5.3 fl oz + Super Tin 6 fl oz/ Priaxor 6 fl oz + Super Tin 6 fl oz/ Minerva Duo 16 fl oz	5.5	33.85	17.45	325.5	11,015	1,298.22
Inspire XT 5.3 fl oz + Topsin 7.6 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt/ Minerva Duo 16 fl oz/ Super Tin 6 fl oz + Manzate 1.2 qt	5.5	32.60	17.43	328.6	10,704	1,288.07
Super Tin 6 fl oz + Topsin 7.6 fl oz/ Inspire XT 5.3 fl oz + Badge 3 pt/ Super Tin 6 fl oz + Manzate 1.2 qt/ Minerva Duo 12 fl oz + Badge 3 pt	5.3	34.28	17.43	327.7	11,218	1,278.67
Super Tin 6 fl oz + Manzate 1.2 qt + Topsin 7.6 fl oz/ Inspire XT 5.3 fl oz + Manzate 1.2 qt/ Super Tin 6 fl oz + Manzate 1.2 qt	5.3	35.23	16.83	312.2	10,957	1,245.46
Inspire XT 5.3 fl oz + Manzate 1.2 qt/ Super Tin 6 fl oz + Manzate 1.2 qt/ Priaxor 6 fl oz + Super Tin 6 fl oz/ Proline 3.8 fl oz + NIS 0.125 % v/v + Manzate 1.2 qt	5.3	33.15	17.33	323.6	10,716	1,238.33
Super Tin 6 fl oz + Manzate 1.2 qt + Topsin 7.6 fl oz/ Inspire XT 5.3 fl oz + Manzate 1.2 qt/ Super Tin 6 fl oz + Manzate 1.2 qt/ Priaxor 6 fl oz + Super Tin 6 fl oz	5.0	34.18	17.18	318.6	10,853	1,235.36
Super Tin 6 fl oz + Topsin 7.6 fl oz/ Inspire XT 7 fl oz/ Priaxor 8 fl oz/ Super Tin 8 fl oz	5.8	33.95	16.95	315.1	10,692	1,220.30
Inspire XT 7 fl oz + Manzate 1.6 qt/ Manzate 1.6 qt/ Proline 5 fl oz + NIS 0.125 %v/v + Topsin 10 fl oz/ Manzate 1.6 qt	5.5	34.58	16.95	315.0	10,900	1,219.74
Super Tin 8 fl oz + Topsin 10 fl oz/ Inspire XT 7 fl oz + Manzate 1.6 qt/ Super Tin 8 fl oz + Manzate 1.6 qt/ Minerva Duo 16 fl oz	4.8	34.63	17.00	313.9	10,847	1,206.47
Super Tin 8 fl oz + Topsin 10 fl oz/ Inspire XT 7 fl oz + Badge 4 pt/ Super Tin 8 fl oz + Manzate 1.6 qt/ Minerva Duo 16 fl oz + Badge 1.6 qt	4.8	34.70	17.66	329.8	11,439	1,154.38
Super Tin 6 fl oz + Manzate 1.2 qt + Topsin 7.6 fl oz/ Inspire XT 5.3 fl oz + Manzate 1.2 qt	5.3	34.03	16.86	314.3	10,696	1,122.37
Untreated Check	10.0	29.90	15.13	277.0	8,289	831.06
LSD (P=0.05)	0.75	3.68	0.69	17.18	1,160	225.93

*Cercospora leaf spot measured on 1-10 scale (1 = 1- 5 spots/leaf or 0.1% severity and 10 = 50% severity) on 8 September.

**Returns based on American Crystal payment system and subtracting fungicide costs and application.

***Treatment applied on 10-12 day interval.

References

- Jones, R. K., Windels, C. E. 1991. A management model for *Cercospora* leaf spot of sugarbeets. Minnesota Extension Service. University of Minnesota. AG-FO-5643-E
- Khan, J., del Rio, L.E., Nelson, R., Khan, M.F.R. 2007. Improving the *Cercospora* leaf spot management model for sugar beet in Minnesota and North Dakota. *Plant Dis.* 91, 1105-1108.
- Khan, M.F.R., Smith, L.J. 2005. Evaluating fungicides for controlling *Cercospora* leaf spot on sugarbeet. *J. Crop Prot.* 24, 79-86.
- Lamey, H. A., Cattnach, A.W., Bugbee, W.M., Windels, C.E. 1996. *Cercospora* leaf spot of sugarbeet. North Dakota State Univ. Ext. Circ. PP- 764 Revised, 4 pp.
- Miller, S.S., Rekoske, M., Quinn, A., 1994. Genetic resistance, fungicide protection and variety approval policies for controlling yield losses from *Cercospora* leaf spot infection. *J. Sugar Beet Res.* 31, 7-12.
- Shane, W.W., Teng, P.S., 1992. Impact of *Cercospora* leaf spot on root weight, sugar yield and purity. *Plant Dis.* 76, 812-820.
- Smith, G.A., Campbell, L.G., 1996. Association between resistance to *Cercospora* and yield in commercial sugarbeet. *Plant Breed.* 115, 28-32.
- Smith, G.A., Ruppel, E.G., 1973. Association of *Cercospora* leaf spot, gross sugar, percentage sucrose and root weight in sugarbeet. *Can. J. Plant Sci.* 53, 695-696.

PLANT-PARASITIC NEMATODES ON SUGARBEET IN NORTH DAKOTA AND MINNESOTA

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INTRODUCTION AND OBJECTIVES

Plant-parasitic nematodes are one of the important groups of pests on sugarbeet. The sugarbeet cyst nematode (*Heterodera schachtii*) is a major pest affecting sugarbeet production in the world (Khan et al. 2016a). This nematode was identified to be the major cause of “beet weariness” which was responsible for the closure of many sugarbeet processing factories in Germany (Harveson and Jackson 2008). Sugarbeet cyst nematode (SBCN) was first discovered in the U. S. in Utah in 1895 and has spread to at least 17 states (Stewart et al. 2014). In 2012 the SBCN was first reported officially in the Yellowstone Valley of western North Dakota (Nelson et al. 2012). Sugarbeet production in Utah and Washington has been terminated largely due to heavy infestations of SBCN which has made growing of sugarbeet unprofitable. In Michigan, this nematode significantly lowered sugarbeet yield and quality, and the estimate of the annual economic loss caused by SBCN to the Michigan Sugar Cooperative is about 5-10 million dollars (Stewart et al. 2014).

Apart from the SBCN, several other nematodes such as stubby root, sting, needle, spiral, sheath, stem and bulb, root knot, false root rot, and potato rot nematodes have been reported as pests on sugarbeet in California, Idaho, Colorado, and other parts of the world. However, they are not known to be a factor for sugarbeet production in North Dakota and Minnesota. Infection with plant-parasitic nematodes often enhances infection by *Rhizoctonia* and other root disease pathogens, which increases the overall effect of the nematode damage.

In June 2015, we received approximately 50 samples from the agriculturists at American Crystal Sugar Company and other extension people. Some of the samples looked like injury from stubby root nematode, needle nematode, or sting nematode. In general these plants were stunted compared to the rest of the field and the roots had very short necrotic lateral roots. Some of the samples were pulled from “sand syndrome” fields in certain areas of the Red River Valley. Six groups of plant-parasitic nematodes were detected including soybean cyst nematode, stubby-root, root-lesion, pin, spiral, and stunt nematodes. In one field with sand syndrome, stubby root nematodes were detected from the area of small and stunted plants but were not detected in the area with healthy plants, which led to the first detection of the stubby root nematode *Paratrichodorus allius* on sugar beet in Minnesota (Yan et al. 2016, Khan et al. 2016b). However, the information on incidence, distribution and species of the plant-parasitic nematodes across North Dakota and Minnesota is limited. The host range of northern-grown crops to the stubby root nematode and effect of the vermiform plant-parasitic nematodes on sugar beet plant growth and crop yield are also not known.

The objectives of this research were to 1) conduct a survey of sugarbeet fields in North Dakota and Minnesota to determine the incidence, abundance and distribution of cyst nematodes and vermiform plant-parasitic nematodes; 2) determine the effect of vermiform plant-parasitic nematodes on plant growth of five sugarbeet cultivars commonly grown in ND and MN; 3) determine the host range of stubby root nematode (*P. allius*), especially for those crops such as wheat, corn, barley, soybean, and sunflower grown in rotation with sugar beet; and 4) evaluate sugarbeet varieties in ND and MN for resistance to *P. allius*.

MATERIALS AND METHODS

Soil and root samples were collected three times once during spring, once in late summer, and once during harvest from sugarbeet fields in the Red River Valley of ND and MN. We worked in collaboration with sugar beet company representatives, sugarbeet producers and extension personnel to identify fields which might be infested with SBCN. Fields with poor sugarbeet growth possibly due to plant-parasitic nematodes were targeted for sampling. A total of 109 soil samples were collected from sugarbeet fields in eight counties in ND, four counties in MN, and one county in Montana. A soil sample consisted of 15-20 soil cores each in 2.5 cm in diameter by 30 cm deep.

Standard laboratory protocols were used in our lab for extracting nematodes from all of the samples and plant-parasitic nematodes including cyst nematodes and vermiform plant-parasitic nematodes were quantified using microscopy. Molecular procedures were optimized and utilized to differentiate SBCN from soybean cyst nematode that were found in sugarbeet fields (Ye 2012). Economically important vermiform plant-parasitic nematodes or nematode pathogens in high densities were attempted to be identified to species using molecular and morphological methods. A panel of nematode control species were requested and obtained from the USDA-ARS Nematology Laboratory in Beltsville MD.

Vermiform plant-parasitic nematode populations were extracted from soil from a naturally infested field to evaluate their effects on plant growth of five sugarbeet cultivars (BTS 8337, Crystal M375, BTS 80RR52, Maribo MA305, BTS 73MNRP). The nematode inoculum were used to inoculate sugarbeet plants under controlled conditions in the greenhouse. At harvest, plants are assessed for emergence rate, plant height, shoot dry weight, root dry weight, and final nematode density. The nematode reproductive factor will be determined by dividing the final nematode population by the initial population inoculated into each pot.

Hosting abilities of sugarbeet and rotational crops to the stubby root nematode will be determined. Northern-grown crops, including wheat, corn, soybean, barley, and sunflower, which are commonly grown in rotation with sugarbeet were evaluated as hosts for the stubby root nematode. Seven sugarbeet cultivars and five rotational crops were included; sugarbeet cultivars: BTS 8337, Crystal M375, BTS 80RR52, BTS 73MNRP, BTS 82RR28, Maribo MA305 and BTS 8500; wheat cultivars: Faller, Glenn, Elgin, Barlow and Brenan; corn cultivars: DK 43-46, DK 43-48, DK 44-13, 1392VT2P and LR9487VT2PRIB; soybean cultivars: Sheyenne, Barnes, HO9X7, SB-8807N and LS-1335NRR2X; barley: Quest and ND-Genesis; and sunflower: Croplan 306 and Mycogen 8N270. A sugarbeet cultivar (BTS 73MNRP) with resistance/tolerance to sugarbeet cyst nematode were included in evaluation for resistance to stubby root nematode.

RESULTS AND DISCUSSIONS

In 2017, soil samples (109) were collected from sugarbeet fields in 8 counties (73 samples from Richland, Walsh, Pembina, Grand Forks, Cass, Traill, Benson, Williams) in ND, 4 counties (34 samples from Clay, Norman, Carver, Aitkin) in MN, and one county (2 samples from Richland) in Montana. Nine groups of plant-parasitic nematodes were detected including cyst nematode, stubby-root, root-lesion, pin, spiral, stunt, dagger, ring and lance nematodes. Thirty-eight soil samples (35%) were infested with stunt nematodes ranging from 20 to 620/100 cc of soil (Table 1, Figure 1). Thirty-five soil samples (32%) contained pin nematodes from 15 to 500/100 cc of soil. Twenty-six soil samples (24%) had spiral nematodes at 15 - 720/100 cc of soil, 11 soil samples (10%) had stubby root nematodes at 15 - 100/100 cc of soil, four samples had root-lesion nematodes at 20 - 60/100cc soil, one sample (1%) had ring nematode at 23/100cc soil, one sample (1%) had dagger nematode at 20/100cc soil, and one sample (1%) had lance nematode at 20/100cc soil (Table 1, Figure 1). Twenty soil samples (18%) were found to have cyst nematodes at 100-8,560/100 cc of soil. The average population densities of these nine groups of plant-parasitic nematodes were calculated, ranging from 20 to 1,196 (Table 1).

Soybean cyst nematode was first detected in ND in 2003 and in MN in 1978 (Bradley et al. 2004, Porter and Chen 2005). Infestation of soybean cyst nematode has spread to many soybean fields in which soybean is a rotational crop of sugarbeet. The soybean cyst nematode and the SBCN have very similar morphology and distinction between them is difficult and time consuming based on morphology using microscopic methods. Molecular procedures were optimized and utilized to identify the cyst nematodes to the species level. The cyst nematodes in nine soil samples were tested using species-specific PCR assays and DNA sequencing. Seven of the samples from ND and MN showed PCR bands specific for soybean cyst nematode using soybean cyst nematode-specific primers but did not produce amplification using sugarbeet cyst nematode-specific PCR primers, indicating these cyst nematodes were soybean cyst nematode but not sugarbeet cyst nematode. Two of the samples from Montana close to the border of ND showed PCR bands specific for sugarbeet cyst nematode using sugarbeet cyst nematode-specific primers but did not produce amplification using soybean cyst nematode-specific primers, indicating these cyst nematodes were sugarbeet cyst nematode. DNA sequencing results confirmed that the samples from ND and MN sugarbeet fields are soybean cyst nematode and the samples from MT sugarbeet fields are sugarbeet cyst nematode.

Likewise, the stubby root nematode we found was identified as *Paratrichodorus allius* using species-specific PCR. This confirms the presence of the stubby root nematode *P. allius* in sugarbeet fields in ND and MN. To determine the species identity of other plant-parasitic nematodes, PCR products from these samples were purified and sequenced. The root-lesion nematode in two samples was identified as *Pratylenchus neglectus*. The spiral nematode in one sample was identified as *Helicotylenchus pseudorobustus*, and pin nematode in one sample was determined as *Paratylenchus nanus*. The stunt nematodes in three samples were identified as a new species that haven't been reported in any literature. More work is needed to further validate the species identity of these plant-parasitic nematodes.

On September 15, 2017, one composite soil sample with 67 stubby root nematodes/100 cc soil along with 160 pin, 160 stunt and 220 spiral nematodes, collected from a field (Cavalier, ND) with a history of "sand syndrome", was used to inoculate seven varieties of sugarbeet, five varieties of each of wheat, corn and soybean crops, and two varieties of each of barley and sunflower. Each of these entries plus one unplanted control were planted in 5 replicates. This set of experiments was harvested on December 22, 2017 and the nematodes are being extracted, identified and counted to determine the resistance reactions of the sugarbeet varieties and hosting abilities of the crop species and varieties.

A soil sample was collected from a field infested with stubby root nematodes. This field (Cavalier, ND) has a history of "sand syndrome". Stubby root nematodes with other vermiform nematodes were extracted from 138 subsamples of the soil for obtaining enough inoculum. The soil was pasteurized to plant the five sugarbeet varieties for determining the effect of nematodes on plant growth by comparing to the plants inoculated with the vermiform nematodes extracted. Two sets of experiments were set up on November 1, 2017 and November 27, 2017, and will be harvested in February.

Table 1. The population densities of plant-parasitic nematodes in 100 cc of soil from 109 soil samples collected from sugarbeet fields.

Nematode Common Name	Nematode Scientific Name	Lowest Density	Highest Density	Average Density
Spiral	<i>Helicotylenchus</i>	15	720	133
Stunt	<i>Tylenchorhynchus</i>	20	620	83
Pin	<i>Paratylenchus</i>	15	500	104
Lesion	<i>Pratylenchus</i>	20	60	40
Dagger	<i>Xiphinema</i>	20	20	20
Stubby root	<i>Paratrichodoros</i>	15	100	36
Cyst nematode	<i>Heterodera</i>	100	8,560	1,196
Ring	<i>Mesocriconema</i>	23	23	23
Lance	<i>Hoplolaimus</i>	20	20	20

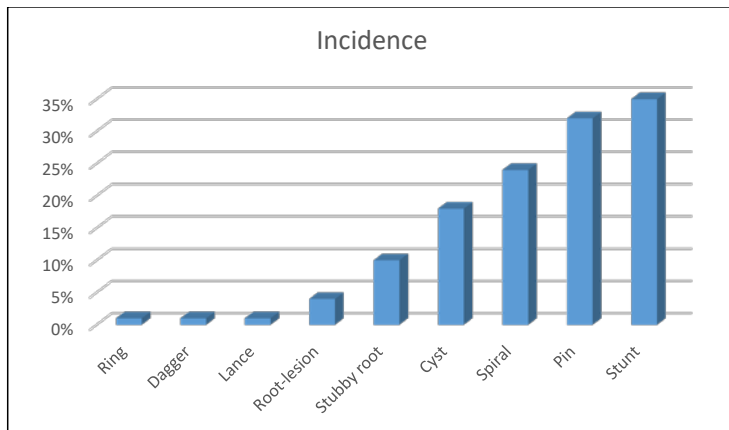


Figure 1. The occurrence frequency (incidence) of plant-parasitic nematodes in 109 soil samples collected from sugarbeet fields.

REFERENCES

- Bradley, C. A., Biller, C. R., and Nelson, B. D. 2004. First report of soybean cyst nematode (*Heterodera glycines*) on soybean in North Dakota. *Plant Disease* 88:1287.
- Harveson, R. M. and Jackson, T. A. 2008. Sugar beet cyst nematode. NebGuide, University of Nebraska-Lincoln Extension, Institute of Agriculture and Natural Resources, Lincoln, NE. 4 p.
- Khan, M., Arabiat, S., Chanda, A. K., and Yan, G. P. 2016a. Sugar beet cyst nematode. North Dakota Extension Bulletin PP1788, North Dakota State Univ., Fargo, ND. 2 p.
- Khan, M., Arabiat, S., Yan, G. P., and Chanda, A. K. 2016b. Stubby root nematode and sampling in sugarbeet. North Dakota Extension Bulletin A1821, North Dakota State Univ., Fargo, ND. 4p.
- Nelson, B. D., Bolton, M. D., Lopez-Nicora, H. D., Niblack, T. L., and Mendoza, L. del Rio. 2012. First confirmed report of sugar beet cyst nematode, *Heterodera schachtii*, in North Dakota. *Plant Disease* 96:772.
- Porter, P. M. and Chen, S. Y. 2005. Sugarbeet cyst nematode not detected in the Red River Valley of Minnesota and North Dakota. *Journal of Sugar Beet Research* 42:79-85.
- Stewart, J., Clark, G., Poindexter, S., and Hubbell, L. 2014. Sugarbeet cyst nematode (BCN) management guide. Michigan Sugarbeet REAch, Research & Education Advisory Council, Bay City, MI. 4 p.
- Yan, G. P., Khan, M., Huang, D., Lai, X., and Handoo, Z. A. 2016. First report of the stubby root nematode *Paratrichodorus allius* on sugar beet in Minnesota. *Plant Disease* 100:1022.
- Ye, W. 2012. Development of primetime-real-time PCR for species identification of soybean cyst nematode (*Heterodera glycines* Ichinohe, 1952) in North Carolina. *Journal of Nematology* 44:284-290.

SCREENING OF SUGAR BEET GERmplasm FOR RESISTANCE TO FUSARIUM YELLOWING DECLINE

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Fusarium spp. can lead to significant economic losses for sugar beet growers throughout the United States production region by causing reductions in yield from several associated diseases (Campbell, Fugate, and Niehaus 2011; Hanson and Hill 2004; Hanson and Jacobsen 2009; Stewart 1931) including Fusarium yellows (Stewart 1931) and Fusarium tip root (Harveson and Rush 1998; Martyn et al. 1989). In 2008, a new sugar beet disease was found in the Red River Valley of MN and ND which caused *Fusarium* yellows-like symptoms but turned out to be more aggressive than Fusarium yellows (Rivera et al. 2008). Symptoms differed from the traditional Fusarium yellows by causing discoloration of petiole vascular elements as well as seedling infection and rapid death of plants earlier in the season. Subsequent studies confirmed that the causal agent of this disease was different from any previously described *Fusarium* species and was therefore named *F. secorum* and the disease it causes as Fusarium yellowing decline (Secor et al. 2014).

F. secorum was shown to belong to the *Fusarium fujikuroi* species complex whereas Fusarium yellows is primarily caused by *Fusarium oxysporum* f. sp. *betae* (Ruppel 1991; Snyder and Hansen 1940) but can be caused by other *Fusarium* spp. including *F. acuminatum*, *F. avenaceum*, *F. solani*, and *F. moniliforme* (Hanson and Hill 2004). Currently, the most effective management strategy for the more common Fusarium yellows is through the use of resistant cultivars and crop rotations with non-hosts (Harveson, Hanson, and Hein 2009) with several sugar beet germplasm being reported to have some resistance (Hanson et al. 2009). However it is unknown if the resistance found in sugar beet to the more common Fusarium yellows will provide any protection against the emerging Fusarium yellowing decline. Therefore this project proposes to screen multiple sugar beet germplasm for resistance against *F. secorum* which causes Fusarium yellowing decline.

Objectives:

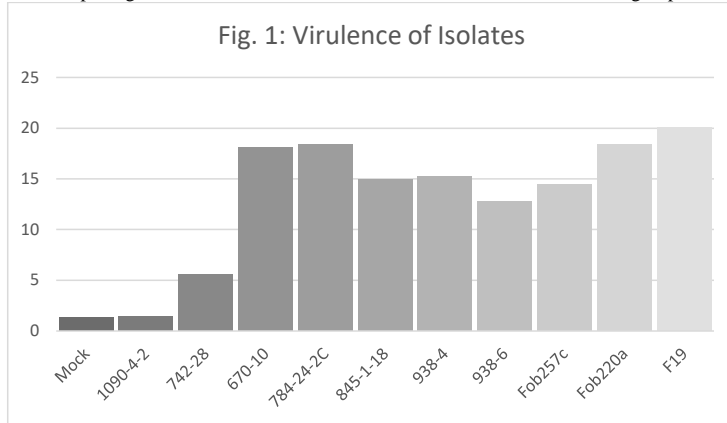
Objective 1: Screen select USDA-ARS, Fort Collins Sugar beet breeding program sugar beet germplasm with known resistance to Fusarium yellows for resistance to Fusarium yellowing decline caused by *F. secorum*. (in progress)

Materials and Methods

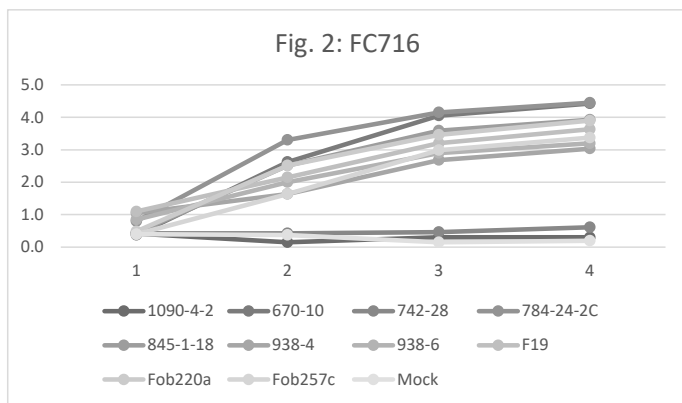
Plant treatment(s). Fifteen sugar beet lines/germplasm will be provided by the breeding program of Dr. Leonard Panella, USDA-ARS, Fort Collins, CO. Additionally, three sugar beet germplasm (Monohikori; FC716; and USH20) will be included as Fusarium yellows susceptible controls. Additional sugar beet lines provided by commercial sugar beet seed companies will be included as requested through lifetime of project. Experiments will be performed as previously described by Secor et al. (2014). Briefly, sugar beet seed will be planted into 6.5cm black plastic “conetainers” using pasteurized potting soil supplemented with Osmocote 14-14-14 slow release fertilizer (Scotts, Marysville, OH). Plants will be grown in a greenhouse with an average daytime temperature of 24°C and average nighttime temperature of 18°C and a 16h photoperiod for 4 weeks. Ten plants will be used for each treatment and will be performed using an augmented split block experimental design (Federer 2005). Briefly, germplasm will be randomly assigned to one of multiple “sets” of inoculations which will be based on the final number of sugar beet germplasm and *F. secorum* isolates. “Sets” will then represent the blocking for the statistical analysis for this experiment. Each inoculation “set” will then be used for two inoculation dates (experiments). At each inoculation date, two replicates will be performed where each isolate is inoculated to a block of five sugar beet plants per germplasm and replicate two times. Statistical analyses will be conducted using SAS Proc Glimmix (SAS Institute, version 9.2, Cary, NC, USA) and the best linear unbiased estimates (Blups) compared to the respective negative and positive controls.

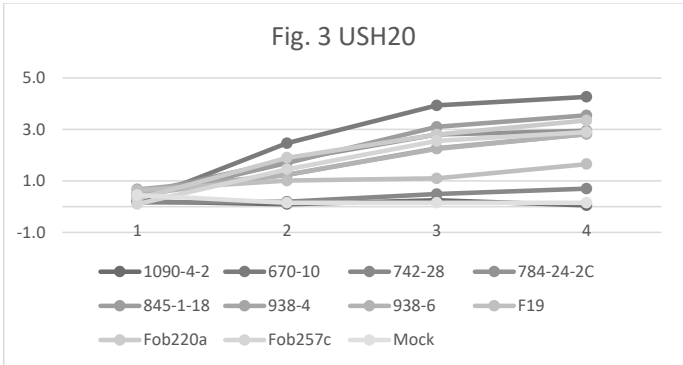
***Fusarium secorum* inoculations.** At inoculation, sugar beet plants that are at 4 weeks after planting will be inoculated by dipping the root into a spore suspension of 1×10^5 conidia ml^{-1} for 2-8 min without agitation (Burlakoti et al. 2012; Secor et al. 2014). Plants will be inoculated with multiple isolates of *F. secorum* including the wild type *F. secorum* (670-10; Secor et al. 2014) and which represent the diversity of the pathogen population throughout the Red River Valley. *F. oxysporum* f. sp. *betae* isolate “F19” will be used as a known positive control for Fusarium yellows and distilled water as the negative control. Treated plants will be maintained in the greenhouse and evaluated for Fusarium yellowing decline symptoms on a weekly basis for 4 weeks after inoculation. Fusarium yellowing decline symptoms will be evaluated using a modified 0-5 Fusarium yellows disease severity rating (Hanson et al. 2009).

Results and Discussion. We were interested in two primary questions 1) Is there a variation in virulence of the *F. secorum* population and 2) Do sugar beet lines and/or cultivars differ in severity of Fusarium yellowing decline. In preliminary experiments, we determined that the *F. secorum* varies in virulence to sugar beet and that this is influenced by variety (Fig. 1). *F. oxysporum* isolates F19 and Fob220a and *F. secorum* isolates 670-10 and 784-24-2C are highly virulent, and mostly end up completely killing the sugar beet plants after 4weeks. *F. secorum* isolates 845-1-18, 938-4, and 938-6 and *F. oxysporum* Fob257c are moderately virulent. One *F. secorum* isolate 742-28, is weakly virulent only causing minor symptoms and is dependent on the cultivar in question. One isolate, 1090-4-2 was non-pathogenic in our studies and will not be included in future screening of potential resistant varieties.



Preliminary results have also indicated that sugar beet cultivars did react differently to the *F. secorum* isolates with some lines having more severe disease symptoms than others. In general, the most susceptible cultivar in these tests was VDH46177 and the least susceptible variety was USH20. Symptoms associated with *F. secorum* such as the half leaf yellowing also appeared to be associated with the cultivar being tested rather than based on isolate however, this trait was not specifically recorded. In future experiments we will record these observations for each isolate by genotype interaction. In general, each cultivar reacted differently to the isolates inoculated. For example, on sugar beet line FC716 all isolates tested caused generally the same amount of disease (Fig. 2) whereas on other lines such as USH20, a clear difference in the susceptibility to some isolates was observed (Fig. 3).





In conclusion, it appears that the *F. secorum* population varies in virulence to sugar beet but that this is similar to the variation that we see for Fusarium yellows caused by *F. oxysporum*. Likewise, there are differences in susceptibility of sugar beet and therefore it is important to screen for both *F. secorum* and *F. oxysporum* populations for each sugar beet production region. Testing for resistance to *F. secorum* was proposed for FY18-19 and findings will be reported in the future.

METABOLOMIC ANALYSIS FOR IDENTIFICATION OF BIOLOGICAL FUNCTIONS ASSOCIATED WITH INFECTION BY AND RESISTANCE TO *BET* NECROTIC YELLOW VEIN VIRUS IN SUGARBEET

William M. Wintermantel, Kimberly Webb, Naveet Kaur and Corey Broeckling

Background:

Rhizomania, caused by *Beet necrotic yellow vein virus* (BNYVV), is one of the most economically important diseases affecting sugarbeet, and is widely distributed in most sugarbeet growing areas of the world. Fields remain infested with BNYVV indefinitely in *P. betae* cystosori that can remain dormant up to 25 years. Rotation to non-host crops or lengthening rotations is ineffective at reducing disease incidence, and to date the only viable means of control has been natural host-plant resistance. The *Rz1* source of resistance was introduced widely to commercial sugarbeet in the 1990s, and for several years has effectively controlled the virus. However, this resistance does not completely eliminate virus replication, but rather suppresses it to low levels compared to what one would find in a susceptible variety. The low level of replication in *Rz1* sugarbeet, has led to the emergence of new variants that overcome or “break” the resistance, since the main forms of the virus that can replicate in *Rz1* varieties are those few variants that have the ability to replicate in the presence of *Rz1*. In the early 2000s an *Rz1* resistance-breaking variant emerged in the Imperial Valley of California (Liu et al., 2005; Rush et al., 2006), and subsequent studies identified the presence of limited numbers of isolated resistance-breaking (RB) variants from most American production regions (Liu and Lewellen, 2007). RB isolates are increasingly affecting production throughout the US industry, and this can be expected to continue.

In addition to the well-known *Rz1* gene, several additional sources of genetic resistance to BNYVV have been identified, and they hold promise. Although these additional genes are being incorporated into sugarbeet varieties, the inability of any of these genes to completely eliminate BNYVV replication leaves all known resistance genes prone to eventual breakdown, even when the genes are “stacked” or combined in order to enhance resistance and make it more difficult for RB strains to establish in plants. It is critically important that rhizomania be studied to allow it to remain under control, and new advances in research approaches create the opportunity for completely new strategies for control of pathogens. This proposal uses one of those new approaches, metabolome analysis, to enhance knowledge of what happens in sugarbeet during infection, as well as to learn how resistance changes the sugarbeet plant to reduce virus accumulation and prevent symptom development. The result of this project will clarify information gained using other technologies and lead to new strategies to enhance and stabilize known forms of resistance. The information generated should also lead to new screening methods that can be applied for identification of varieties with enhanced resistance, as well as for the identification of new approaches to protect sugarbeet from rhizomania.

The different sources of resistance to BNYVV map to different chromosomal positions and although some may be allelic to one other, others appear to be distinct (Scholten et al., 1997; 1999; Gidner et al., 2005). Furthermore, several minor genes may contribute to enhanced resistance (Gidner et al., 2005), although further characterization of how this works is necessary. With the introduction of Roundup-Ready sugarbeets a few years ago, we may begin to see new opportunities for the application of biotechnology-based resistance at least in the US and Canada. Furthermore, the sequencing of the sugarbeet genome and the aggressive development of genetic markers creates additional opportunities for selective breeding that can target development and selection of sugarbeet with specific and unique traits that may lead to enhanced resistance as we learn more about how BNYVV infects sugarbeet and overcomes known sources of resistance. Finally, the emergence of new gene editing technologies may also lead to the ability to specifically target individual genes for up-or down-regulation that may enhance pathogen resistance or yield related traits.

With these things in mind, we recently completed studies evaluating resistant and susceptible sugarbeet using proteomics methods in order to gain an understanding of what BNYVV does at the cellular level to allow it to infect and cause disease in sugarbeet. These studies led to new knowledge on how BNYVV infection and virus resistance alter protein expression associated with infection and resistance in sugarbeet. (Larson et al., 2008; Webb et al., 2014 & 2015). Others have used different strategies to identify other protein interactions that may contribute to infection (Thiel and Varrelmann, 2009). Recent studies have begun to examine how gene activity (transcriptomics) is influenced by BNYVV infection (Fan et al., 2014; 2015). All of this information is informative on its own, but

studies on gene expression (RNA and protein) only provide part of the picture. By utilizing a metabolomics approach, we can complement knowledge of gene expression provided through previous studies, and obtain a more complete picture of what is happening during BNYVV infection and resistance.

Metabolomics provides an analytic tool that enables the qualitative and quantitative profiling of metabolites in a biological system and serves as a link between the plant genotype and phenotypic (visible or obvious) responses (Fiehn et al., 2000; Heuberger et al., 2014). A better understanding of plant metabolism in response to different biotic stresses (including pathogen infections) facilitates a better understanding of plant physiology which is crucial for the development of future applications in plant breeding, biotechnology, and crop protection (Aliferis and Jabaji, 2012a). Metabolomic profiling was previously used in sugar beet to complement proteomic studies characterizing the response of carbohydrate metabolism and the TCA cycle to iron deficiency (Rellan-Alvarez et al., 2010). Aliferis and Jabaji (2012b) have described the metabolome of potato in response to infection with *R. solani* AG 3; however, to date there have been no known reported studies on the effects of plant pathogens on the metabolome of sugar beet. Additionally, metabolomic approaches have been utilized in characterizing compounds found in *R. solani* that are directly important to the fungus during critical life stages as well as characterizing the metabolites found in fungal exudates which may contain clues to secreted effectors (Aliferis and Jabaji, 2010a,b). Application of this technology to BNYVV in sugarbeet should lead to the identification of compounds necessary for both infection and resistance.

The information provided through our studies has the potential to lead to a new era in management of BNYVV. By utilizing knowledge of how the virus infects sugarbeet, combined with increasing knowledge of the sugarbeet genome and a growing number of molecular markers, it should be possible to enhance performance of existing resistance genes through selective, marker based breeding practices (Laurent et al., 2007). This could include selection for sugarbeet varieties enhanced for specific metabolites, as well as identifying new ways to reduce or even prevent BNYVV from infecting sugarbeet through targeted breeding directed at enhancing or manipulating specific biological pathways within the plant through specific gene editing approaches.

Objectives:

1. Compare the metabolome of a near isogenic line of susceptible (*rz1*) sugarbeet with those of sugarbeet with each of two resistance genes against BNYVV (*Rz1* and *Rz2*) at specific time points in the infection process.
2. Identify important compounds/cellular chemicals that may be critical to BNYVV infection of susceptible sugarbeet and for suppression of BNYVV in resistant sugarbeet.
3. Compare results with existing knowledge of RNA and protein expression changes associated with infection and resistance from previous studies to identify targets for interference and potential resistance.

Summary of Project to Date:

Three sugarbeet lines with closely related genetic backgrounds (near isogenic lines) from the ARS-Salinas sugarbeet breeding program (developed by Lewellen and Richardson) were used in seedling grow-out experiments; one with the *Rz1* resistance gene (C79-1), another with the *Rz2* resistance gene (C79-3), and one susceptible to BNYVV (C37). All three lines share the same genetic background and differ by the type of *Rz* gene they carry. Soil containing a well established isolate of BNYVV pathotype A (source collected from USDA-ARS, Spence Field in Salinas in 2006) was mixed with equal parts sterile builders sand and placed into new Styrofoam cups. Parallel studies were performed with an *Rz1* resistance breaking strain of BNYVV (Imperial Valley, CA - Rockwood 158 RB isolate), as well as with virus-free (healthy) soil. For each sugar beet variety, 50 seeds were planted per cup, with two cups per treatment, and grown in a growth chamber at 24°C with 16-hour days and approximately 220 $\mu\text{M m}^{-1}\text{s}^{-2}$ light until 3 weeks after sowing. At each time point, seedlings from each cup was handled independently for each treatment to assure good infection of each sample/treatment. Foliar and root portions of the plant were separated at the crown and lyophilized (freeze dried), then stored at -80 until further analysis. Root samples from each plot were tested by RT-PCR to confirm BNYVV infection prior to use in metabolome analysis, and remaining roots from the same samples were used for metabolite extractions.

Roots from all three experiments were freeze-dried and stored at -80C so that metabolites could be extracted from all samples at the same time. Upon completion of the last replication, dried root samples were pulverized in liquid nitrogen and sent to the Core Laboratory at Colorado State University (CSU) in Ft. Collins, CO for methanol extraction of metabolites. Metabolome analysis was completed at CSU during the fall, and results of analyses provided to USDA-ARS in late November 2017.

Overall 746 metabolites were found and these were annotated to known compounds or to unknown compounds with a specified mass. These metabolites were examined in all possible combinations of treatments to look for statistically different levels of expression among treatments, including patterns of expression indicating how traditional or RB-BNYVV influence resistant and susceptible sugarbeet during infection, as well as for identification of “interesting” compounds that may play an important role in rhizomania disease development. Metabolite levels were compared among treatments using a 95 percent confidence interval to distinguish compounds with statistically different levels of expression among treatments. Results demonstrated the most important difference in metabolite levels was between healthy sugarbeet plants and sugarbeet plants infected with BNYVV. Results also demonstrated differences between traditional BNYVV and RB-BNYVV. Overall, comparative studies indicated 32% of differences in metabolite levels among treatments were based on the presence or absence of BNYVV (Fig. 1). In contrast, only 3% of variation among treatments could be explained by differences in sugarbeet variety (i.e. the different resistance genes) (Fig. 2). Essentially, results indicate most metabolic differences are caused by the BNYVV infection, and are not influenced much by the presence or absence of either resistance gene. This contrasts with what was observed with our recent proteomics analysis of similar sugarbeet near isogenic lines, in which differences that occurred were influenced by both virus strain and the resistance genes.

In our previous proteomics analysis comparing BNYVV infection of Rz1 and Rz2 sugarbeet with susceptible sugarbeet, we identified a number of proteins with differential expression not only between RB- and traditional strains of BNYVV, but also between sugarbeet genotypes (Rz1, Rz2, and susceptible). Results of those studies demonstrated that abundance of select proteins in sugarbeet is significantly altered based on the presence or absence of the two resistance genes (Webb et al., 2015), whereas in the current metabolomics study very limited (3%) differences in the metabolome were determined by the presence or absence of rhizomania resistance genes.

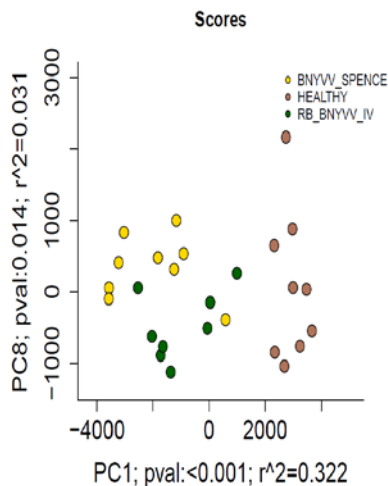


Figure 1. Principle component analysis plot generated from 27 samples derived from 9 treatments showing clear separation by virus type. Yellow: BNYVV-Spence (traditional/wild type BNYVV), Green: BNYVV-IV (Rz1 Resistance breaking BNYVV), Red: Healthy (virus-free sugarbeet).

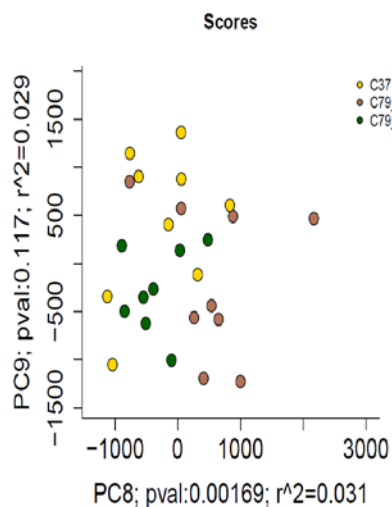


Figure 2. Principle component analysis plot generated from 27 samples derived from 9 treatments showing little separation by sugarbeet genotype (resistance gene or not). Yellow: C37 (susceptible sugarbeet [rz1rz2]), Red: C79-1 (Rz1 resistant sugarbeet [Rz1rz2]), Green: C79-3 (Rz2 resistant sugarbeet [rz1Rz2]).

Continuing studies are focusing on identification of specific compounds that differ among treatments. Although these detailed studies are just beginning, some interesting results have already been identified, including compound $C_{40}H_{107}N_{17}OS_4$ (Fig. 3). This compound had low expression in the absence of virus in both susceptible (rz1rz2) and resistant varieties (both Rz1 and Rz2), but higher expression with virus infection when either traditional or Rz1-resistance-breaking BNYVV strains were present. In general, the expression of this compound mimics what would be “expected” in a traditional gene-for-gene type of resistance. The highest expression of compound $C_{40}H_{107}N_{17}OS_4$ was observed in the susceptible line (C37) with the traditional BNYVV strain (Spence), but expression differences were also significant with the RB BNYVV strain (which we believe is generally less fit overall than traditional BNYVV based on its performance in field situations). The fact that this compound is expressed at elevated levels in all varieties indicates its expression is a response to infection, but not necessarily associated with ability of the plant to resist infection (no strong differential effect with resistant beets).

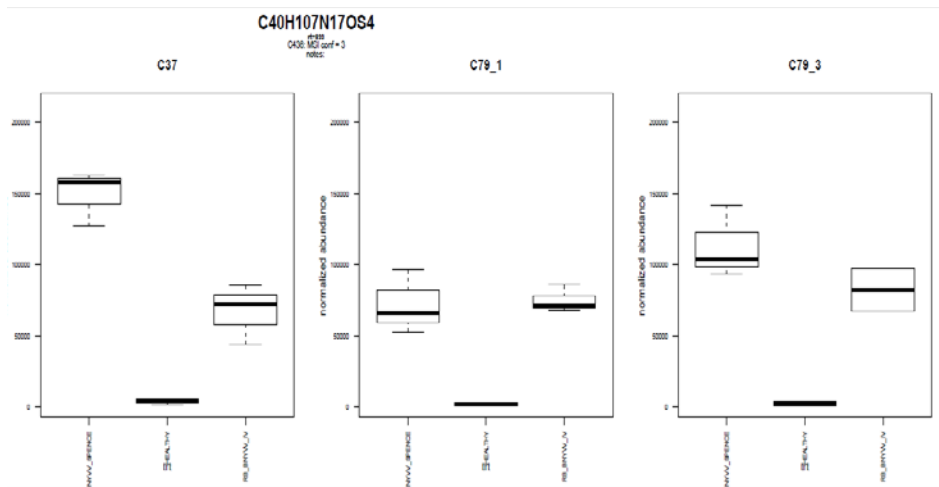


Figure 3. Abundance of compound C₄₀H₁₀₇N₁₇OS₄. C37 = susceptible sugarbeet (rz1rz2), C79-1 = RZ1 resistant sugarbeet (Rz1rz2), C79-3 = RZ2 resistant sugarbeet (rz1Rz2).

Further Research:

Although we have not requested additional funding for this project we will be continuing data analysis and interpretation of results. Through characterization of differential abundance of compounds and identification of these compounds, we expect to improve our knowledge of what is happening biochemically in sugarbeet during BNYVV infection and development of rhizomania disease. We will also examine results of this metabolome analysis in comparison to those of our previous studies on proteomics (Larson et al., 2008; Webb et al., 2014, 2015), and studies by others on gene expression and protein interactions (Fan et al., 2014, 2015; Thiel and Varrelmann, 2009). This should allow us to begin to piece together how BNYVV causes disease in plants by determining changes that occur in infected vs. healthy sugarbeet. Ultimately we anticipate gaining insight into how resistance genes are able to suppress BNYVV levels by identifying differences in biochemicals produced (this study) along with changes in gene expression (previous studies). This information will be useful toward application of marker-based selection of traits that may enhance performance of resistance genes, as well as for identification of targets for use of new biotechnology-based methods that should lead to novel methods to prevent rhizomania disease in sugarbeet.

Literature Cited:

Acosta-Leal, R., and C.M. Rush. 2007. Mutations associated with resistance-breaking isolates of Beet necrotic yellow vein virus and their allelic discrimination using TaqMan technology. *Phytopathology* 97: 325-330.

Aliferis, K. A. and S. Jabaji. 2010a. H NMR and GC-MS metabolic fingerprinting of developmental stages of *Rhizoctonia solani* sclerotia. *Metabolomics* 6:96-108.

Aliferis, K. A. and S. Jabaji. 2010b. Metabolite composition and bioactivity of *Rhizoctonia solani* sclerotial exudates. *Journal of Agricultural and Food Chemistry* 58:7604-7615.

Aliferis, K. A. and S. Jabaji. 2012a. Deciphering plant-pathogen interactions applying metabolomics: principles and applications. *Canadian Journal of Plant Pathology* 34:29-33.

Field Code Changed

- Aliferis, K. A. and S. Jabaji. 2012b. FT-ICR/MS and GC-ESI/MS metabolomics networking unravels global potato sprout's responses to *Rhizoctonia solani* infection. *PLOS one* 7, no. 8:1-8.
- Fan H, Sun H, Wang Y, Zhang Y, Wang X, Li D, et al. (2014) Deep sequencing-based transcriptome profiling reveals comprehensive insights into the responses of *Nicotiana benthamiana* to *Beet necrotic yellow vein virus* infections containing or lacking RNA4. *PLoS One* 9: e85284. doi: 10.1371/journal.pone.0085284. pmid:24416380
- Fan, H., Zhang, Y., Sun, H., Liu, J., Wang, Y., Wang, X., Li, D., Yu, J., and Han, C. 2015. Transcriptome analysis of *Beta macrocarpa* and identification of differentially expressed transcripts in response to Beet necrotic yellow vein virus Infection. *PLoS One* DOI: 10.1371/journal.pone.0132277
- Fiehn, Oliver, Joachim Kopka, Peter Dormann, Thomas Altmann, Richard N. Trethewey, and Lothar Willmitzer. 2000. Metabolite profiling for plant functional genomics. *Nat Biotech* 18, no. 11:1157-1161.
- Gidner, S., B. L. Lennefors, N. O. Nilsson, J. Bensefelt, E. Johansson, U. Gyllenspetz, and T. Kraft. 2005. QTL mapping of BNYVV resistance from the WB41 source in sugar beet. *Genome* 48:279-285.
- Heuberger, Adam L., Corey D. Broeckling, Kaylyn R. Kirkpatrick, and Jessica E. Prenni. 2013. Application of nontargeted metabolite profiling to discover novel markers of quality traits in an advanced population of malting barley. *Plant Biotechnol J*:n/a.
- Heuberger, Adam L., Faith M. Robison, Sarah Marie A. Lyons, Corey D. Broeckling, and Jessica E. Prenni. 2014. Evaluating plant immunity using mass spectrometry-based metabolomics workflows. *Frontiers in Plant Science* 5:1-11.
- Larson, R. L., W. M. Wintermantel, A. L. Hill, L. Fortis, and A. Nunez. 2008. Proteome changes in sugar beet in response to *Beet necrotic yellow vein virus*. *Physiol. and Mol. Plant Pathol.* 72:62-72.
- Laurent V, Devaux P, Thiel T, Viard F, Mielordt S, Touzet P, et al. (2007) Comparative effectiveness of sugar beet microsatellite markers isolated from genomic libraries and GenBank ESTs to map the sugar beet genome. *Theor Appl Genet* 115: 793–805. pmid:17646961 doi: 10.1007/s00122-007-0609-y
- Liu, H., J. L. Sears, and R. T. Lewellen. 2005. Occurrence of resistance breaking *Beet necrotic yellow vein virus* of sugar beet. *Plant Disease* 89:464-468.
- Liu, H. Y. and R. T. Lewellen. 2007. Distribution and molecular characterization of resistance-breaking isolates of Beet necrotic yellow vein virus in the United States. *Plant Dis.* 91:847-851.
- Rellan-Alvarez, R., S. Andaluz, J. Rodriguez-Celma, G. Wohlgemuth, G. Zocchi, A. Alvarez-Fernandez, O. Fiehn, A. F. Lopez-Millan, and J. Abadia. 2010. Changes in the proteomic and metabolomic profiles of *Beta vulgaris* root tips in response to iron deficiency and resupply. *BMC Plant Biology* 10:120.
- Rush, C. M., H. Y. Liu, R. T. Lewellen, and R. Acosta-Leal. 2006. The continuing saga of rhizomania of sugar beets in the United States. *Plant Disease* 90:4-15.
- Scholten, O. E., T. S. De Bock, R. M. Klein-Lankhorst, and W. Lange. 1999. Inheritance of resistance to *Beet necrotic yellow vein virus* in *Beta vulgaris* conferred by a second gene for resistance. *TAG* 99:740-746.
- Scholten, O. E., R. M. Klein-Lankhorst, D. G. Esselink, T. S. De Bock, and W. Lange. 1997. Identification and mapping of random amplified polymorphic DNA (RAPD) markers linked to resistance against *Beet necrotic yellow vein virus* (BNYVV) in *Beta* accessions. *TAG* 94:123-130.
- Thiel, H., and Varrelmann, N. 2009. Identification of Beet necrotic yellow vein virus P25 pathogenicity factor-interacting sugar beet proteins that represent putative virus targets or components of plant resistance. *Mol Plant-Microbe Interact.* 22(8): 999-1010.

- Webb, K. M., C. J. Brocardo, J. E. Prenni, and W. M. Wintermantel. 2014. Proteomic profiling of sugar beet (*Beta vulgaris*) leaves during rhizomania compatible interactions. *Proteomes* 2:208-223.
- Webb, K. M., W. M. Wintermantel, N. Kaur, J. E. Prenni, C. J. Brocardo, L. M. Wolfe, and L. L. Hladky. 2015. Differential abundance of proteins in response to *Beet necrotic yellow vein virus* during compatible and incompatible interactions in sugar beet containing *Rz1* or *Rz2*. *Physiological and Molecular Plant Pathology* 91:96-105.
- Wisler, G. C., R. T. Lewellen, J. L. Sears, H. Liu, and J. E. Duffus. 1999. Specificity of TAS-ELISA for *Beet necrotic yellow vein virus* and its application for determining rhizomania resistance in field-grown sugar beets. *Plant Disease* 83:864-870.

Budget Justification: Funds for general laboratory supplies, as well as kits and reagents necessary for metabolite extraction, and other metabolome analyses were provided through a combination of BSDF funds and USDA-ARS in-house funds (Not SBREB) during 2016. These charges were covered with 2016 funds. Plant growth work at Salinas is nearly completed, and all samples will be sent for analysis once the current and final experiment is completed this month (Dec. 2016).

A GS-11 USDA-ARS postdoctoral research associate (Dr. Navneet Kaur, ARS Salinas) will conduct data analysis, with guidance and assistance from Drs. Broekling (CSU) and Webb (USDA) in Ft. Collins. Dr. Kaur's salary for sample preparation and research on this project was provided by SBREB in 2016. We are only requesting \$6,000 from SBREB in 2017 to support Dr. Kaur's salary (additional salary funds were requested from BSDF). Dr. Kaur cannot be paid with USDA in-house funds due to her nationality (India). Therefore we are requesting limited funds from BSDF to assist with Dr. Kaur's salary to finish out the project involving data analysis and interpretation of results. An existing agreement is in place between USDA-ARS and BSDF to utilize BSDF funds for ARS salaries.

Budget:	USDA	BSDF	SBREB
Labor	\$25,000	\$10,000	\$6,000
Equipment (over \$250.00)	\$0	\$0	\$0
Supplies	\$800	\$0	\$0
Service (metabolomics analysis)	\$9,000	\$0	\$0
Travel	\$0	\$0	\$0
TOTALS:	\$34,800	\$10,000	\$6,000

SUGARBEET VARIETIES / QUALITY TESTING

NOTES

RESULTS OF AMERICAN CRYSTAL SUGAR COMPANY'S 2017 CODED OFFICIAL VARIETY TRIALS

William S. Niehaus, Official Trial Manager
American Crystal Sugar Company
Moorhead, Minnesota

American Crystal Sugar Company's (ACSC) coded Official Variety Trials (OVT) are designed to provide an unbiased evaluation of the genetic potential of sugarbeet variety entries under several different environments. The two-year average of these evaluations then are used to establish a list of approved varieties which ensures the use of high quality, productive varieties to maximize returns for growers and the cooperative as a whole. This report presents data from the 2017 American Crystal OVTs and describes the procedures and cultural practices involved in the trials.

Table	Information in the Table
1	ACSC approved varieties for 2017
2	Multi-year performance of approved varieties (all locations combined)
3	Performance of ACSC Aphanomyces specialty varieties
4	Performance data of approved conventional varieties (all locations combined)
5	Disease ratings for ACSC tested varieties (multiple diseases)
6	Official trial sites, cooperators, plant and harvest dates, soil types and disease notes
7	Seed treatments applied to seed used in the OVTs
8-18	2017 Roundup Ready variety trials and combined trials
19-25	2017 Conventional variety trials and combined trials
26-29	Approval calculations for ACSC market
30	Aphanomyces disease nursery ratings
31	Cercospora disease nursery ratings
32	Rhizoctonia disease nursery ratings
33	Fusarium disease nursery ratings
34	Herbicides and fungicides applied to official trials

Procedures and Cultural Practices

Sugarbeet official variety testing was conducted the ACSC growing region areas of the Red River Valley by ACSC personnel at the Technical Services Center.

All entries were assigned a code number by KayJay Ag Services. The seed then was sent to ACSC Technical Services Center at Moorhead for official testing.

Thirteen official yield trial sites were planted in the ACSC area with eleven harvested. Plant-to-stand trials (4.5 inch spacing) were used to evaluate the commercial, experimental and conventional varieties. Seed companies had the option of treating seed with Tachigaren, insecticide and a Rhizoctonia seed treatment fungicide. The treatments used on the seed planted in the official variety yield trials can be found in table 7.

Ten ACSC sites were used for variety approval calculations (Felton, Georgetown, Hendrum, Hillsboro, Climax, Grand Forks, Scandia, St. Thomas, Stephen and Bathgate). One site was abandoned due to erratic emergence (Casselton) and two were abandoned due to soil compaction (Humboldt and Argyle). Rhizoctonia was less prevalent in 2017 yet showed an increase from 2016 in yield trials. Based upon susceptible plot observations, root aphids had only a slight effect on varieties in 2017.

Plots were planted crosswise (90°) to the cooperators' normal farming operations, where possible. Plot row lengths for all official trials were maintained at 44 feet with about 39 feet harvested. All trials had two or four-row plots planted in four or six replications. Planting was performed with a 12-row SRES vacuum planter. The GPS controlled planter gave good single seed spacing which facilitated emergence counting. Emergence counts were taken on 24 feet of each plot. Multiple seedlings were counted as a single plant if they emerged less than one inch apart. The stands in all yield trials were refined by removing doubles (multiple seedlings less than 1.5 inch apart) by hand but were not further reduced.

Roundup Powermax with Event and full rates of fungicides were applied using a pickup sprayer driven down the alleys. Hand weeding was used where necessary. The micro rate program was used on conventional trials. All yield trials were treated with Quadris in a band during the 2 leaf (9 oz) and 6-10 leaf stage (14 oz) for Rhizoctonia control. Treatments used for Cercospora control in 2017 included Inspire XT/Penncozeb, Agritin/Incognito, Penncozeb, and Headline/Agritin. Ground spraying was conducted by ACSC technical staff.

Roundup Ready (RR) varieties with commercial seed were planted in four-row, six replication trials. The RR experimental entries were planted in smaller two-row, four replication trials. Two applications of Roundup were made in the 4-6 (32 oz) and 8-12 (22 oz) leaf stages.

ACSC Conventional OVT's were reinstated in 2016 and repeated again in 2017. Approval was based on one year of data with eighteen varieties approved for 2018 sales. Three conventional varieties were previously approved and have data from the 2012 Sugarbeet Research and Extension Report.

All plot rows were measured for total length after approximately 2.5 feet at each end were removed at the end of August, with skips greater than 60 inches being measured for adjustment purposes. Harvest was performed with two modified four-row harvesters (4310 and 4310A John Deere). All harvested beets of each plot were used for yield determination while one sample (approx 25 lbs) for sugar and impurity analysis was obtained from each plot. Quality analysis was performed at the ACSC Technical Services quality lab in Moorhead.

Varieties were planted in disease nurseries in North Dakota, Minnesota and Michigan to evaluate varieties for disease tolerance.

The ACSC official variety trial program attempts to utilize multiple disease nurseries adjusting the Cercospora, Aphanomyces, Rhizoctonia and Fusarium nursery data each year to provide consistency to the disease ratings. In 2017, the disease ratings for Aphanomyces (Shakopee) and Rhizoctonia (Michigan) were limited to a single location due to lack of disease pressure in the RRV. Consider reviewing all available disease ratings when evaluating variety performance.

Acknowledgements

Thanks to the beet seed companies for their participation in the official variety testing program and to all grower-cooperators, agricultural, and beet seed staff for their assistance. Special thanks are extended to Dr. Mohamed Khan for Cercospora nursery infection, Dr. Albert Sims for hosting a Rhizoctonia nursery, Randy Nelson for RRV disease ratings, USDA staff in Michigan for Cercospora and Rhizoctonia nursery ratings. The Betaseed staff for Aphanomyces and Cercospora ratings in the Shakopee area, and Kay Jay Ag Services for sampling and coding all variety entries.

Table 2. Performance Data of RR Varieties During 2015, 2016, 2017 Growing Seasons (All Locations Combined)+++

Variety #	Yrs	Rev/Tn ++				Rev/Acs ++				Sugar				Yield				Miles/acre	Emerg	Bolls/Ac			CR +	Ash Rcks			Ribs/cv			Fusarium/Rm										
		17	2015	2016	2017	17	2015	2016	2017	17	2015	17	2015	17	2015	17	2015			17	2015	17		2015	17	2015	17	2015	17	2015	17	2015	17	2015	17	2015	17	2015	17	2015
		10	19	29		10	19	29		10	19	10	19	10	19	10	19			10	19	10		19	10	19	10	19	10	19	10	19	10	19	10	19	10	19	10	19
Previous Approved																																								
BTS 80R52	6	52.79	52.12	100	52.82	100	1699	1830	102	1787	103	334	326	10789	11432	17.94	17.45	32.4	35.3	1.22	1.17	78	75	5	2	4.37	4.33	4.4	4.2	4.1	4.3	2.7	2.7	H						
BTS 8337	3	57.43	55.76	107	56.89	108	1842	1860	104	1825	105	350	337	11209	11247	18.55	17.96	32.1	33.4	1.06	1.10	76	72	5	2	4.36	4.49	3.8	3.5	4.3	4.2	3.8	3.9	H						
BTS 8383	3	51.14	50.20	97	50.68	96	1770	1854	104	1813	104	329	319	11391	11772	17.53	17.03	34.7	37.0	1.09	1.08	78	76	0	0	4.50	4.21	4.6	4.8	4.9	4.6	3.5	3.3	H						
BTS 8500	1	53.24	51.10	98	51.62	98	1862	1914	107	1855	107	336	322	11741	12068	17.90	17.22	35.0	37.6	1.11	1.10	78	76	0	0	4.29	4.41	4.5	4.4	4.6	4.5	2.1	2.0	H						
BTS 8512	1	54.51	52.80	102	53.37	101	1749	1833	103	1793	103	340	328	10921	11360	18.08	17.48	32.2	34.8	1.06	1.08	79	77	14	7	3.69	3.86	3.8	4.0	4.3	4.4	3.0	2.8	H						
BTS 8524	1	51.51	49.79	96	50.15	95	1736	1875	105	1831	105	330	318	11506	11961	17.84	17.03	34.9	37.7	1.14	1.14	79	79	5	2	4.38	4.56	4.5	4.2	4.4	4.3	3.2	3.3	H						
BTS 8572	1	56.57	54.96	106	55.68	106	1817	1865	104	1816	105	347	335	11147	11365	18.41	17.81	32.2	34.0	1.07	1.05	78	77	0	0	4.14	4.27	3.8	4.1	4.3	4.4	2.5	2.4	H						
Cystal 933RR	6	57.63	54.91	106	55.51	105	1866	1904	107	1850	106	350	335	11329	11633	18.60	17.84	32.4	34.8	1.06	1.10	76	74	0	0	4.49	4.72	4.4	4.4	4.5	4.4	3.5	3.4	H						
Cystal 9101R	6	51.29	49.71	96	50.79	96	1718	1794	100	1728	99	329	318	11040	11400	17.66	17.10	33.6	36.0	1.20	1.22	77	73	0	0	4.57	4.58	3.9	3.7	4.8	4.6	2.7	2.6	H						
Cystal 246RR	4	52.05	49.94	96	50.68	96	1775	1810	101	1774	102	332	319	11322	11534	17.67	17.01	34.2	36.3	1.09	1.09	74	75	0	0	4.63	4.72	5.1	5.0	4.2	4.3	3.2	3.2	H						
Cystal 247RR	4	53.09	51.91	100	52.76	100	1852	1923	108	1886	109	335	325	11575	12031	17.79	17.28	34.6	37.1	1.03	1.04	76	72	18	9	4.55	4.60	5.4	5.1	4.5	4.4	3.0	2.9	H						
Cystal 255RR	2	54.56	53.87	104	54.20	103	1711	1829	102	1781	101	340	331	10883	11243	18.16	17.70	31.5	34.0	1.15	1.14	75	76	0	0	4.36	4.48	4.8	4.7	4.1	4.0	2.8	2.7	H						
Cystal 467RR	1	51.86	49.09	94	50.11	95	1804	1825	102	1825	104	330	316	11588	11754	17.63	16.91	35.2	37.4	1.12	1.14	80	75	0	0	4.49	4.57	4.0	4.0	4.5	4.4	2.0	1.9	H						
Cystal 672RR	1	58.99	56.37	108	56.67	108	1891	1937	108	1866	107	355	340	11379	11673	18.74	18.01	32.1	34.5	1.01	1.02	81	79	0	0	4.27	4.42	4.7	4.7	4.5	4.3	2.6	2.2	H						
Cystal 673RR	NC	55.68	54.22	104	54.82	104	1786	1877	105	1837	106	344	330	11039	11512	18.28	17.71	32.1	34.7	1.08	1.07	75	74	0	0	4.15	4.26	3.8	4.0	4.6	4.6	3.1	3.3	H						
Cystal 674RR	1	52.84	50.76	98	51.24	97	1875	1973	110	1915	110	334	321	11851	12453	17.79	17.14	35.4	38.8	1.08	1.09	79	79	0	0	4.35	4.43	4.7	4.2	4.2	4.3	2.2	2.0	H						
Cystal 679RR	NC	54.05	52.68	101	53.46	101	1899	1958	110	1904	110	338	328	11908	12160	18.00	17.43	35.3	37.2	1.07	1.05	80	78	0	0	4.91	4.89	4.6	4.5	4.4	4.4	2.4	2.2	H						
Cystal 989RR	6	54.89	53.45	103	54.13	103	1776	1856	103	1772	103	341	330	11058	11298	18.09	17.51	32.2	34.2	1.08	1.01	78	77	0	0	4.77	4.76	4.1	4.2	4.4	4.4	4.7	4.8	Rzm						
Hilleshov HL9707	1	49.79	48.85	94	50.46	96	1862	1716	96	1661	96	324	315	11020	11042	17.35	16.86	34.0	35.1	1.14	1.13	72	66	5	2	4.96	4.74	4.7	4.3	4.4	4.4	4.1	4.5	H						
Hilleshov HL9708	NC	54.11	52.09	100	53.34	101	1840	1749	98	1730	100	339	326	10290	10933	18.02	17.34	30.4	33.7	1.07	1.06	74	76	9	5	4.61	4.68	5.9	5.4	4.2	4.2	4.6	4.4	Rzm						
Hilleshov 4302RR	4	52.73	50.18	100	53.06	101	1897	1899	95	1674	95	334	326	10993	10956	17.75	17.33	30.1	32.5	1.06	1.04	68	65	0	0	3.93	4.03	6.7	5.6	3.6	3.6	5.1	5.1	Rzm						
Hilleshov 4448RR	4	53.93	51.47	99	53.10	101	1829	1851	104	1840	106	338	324	11456	11817	17.97	17.24	33.9	35.9	1.06	1.06	70	67	5	2	5.28	5.24	6.3	5.1	4.6	4.6	5.3	5.3	Rzm						
Hilleshov 5259RR	3	54.35	53.26	102	54.10	103	1785	1884	106	1843	106	339	329	11544	11631	18.02	17.51	32.9	35.4	1.05	1.05	74	71	5	2	4.99	4.86	5.8	4.7	4.2	4.2	4.2	4.4	H						
Marbo 109	2	56.86	56.60	109	57.45	109	1596	1729	97	1675	96	348	340	9579	10365	18.43	18.03	27.5	30.5	1.06	1.04	67	68	0	0	4.14	4.14	5.1	4.7	3.6	3.7	4.2	4.4	H						
Marbo 305	2	52.03	50.29	97	50.67	96	1731	1752	98	1713	99	332	320	11018	11121	17.60	17.00	33.2	34.8	1.02	1.02	67	65	0	0	4.98	4.85	5.7	5.0	4.6	4.5	5.9	5.9	Rzm						
Marbo M502	1	51.46	49.31	95	50.47	96	1642	1733	97	1716	99	330	316	10539	11124	17.66	17.00	32.0	35.4	1.17	1.20	74	72	66	34	5.01	4.90	3.5	3.3	4.8	4.8	3.0	2.5	H						
Marbo M504	NC	52.79	50.34	97	51.06	98	1830	1879	105	1875	108	334	320	11842	11946	17.77	17.07	34.8	37.6	1.07	1.08	77	75	0	0	5.50	5.27	6.0	5.4	4.4	4.6	4.5	4.6	H						
SV RR447T	2	52.83	52.31	101	52.78	100	1796	1837	103	1787	103	335	326	11339	11427	17.79	17.36	33.8	35.0	1.05	1.05	72	69	5	2	4.85	4.85	4.9	4.9	4.5	4.5	3.7	3.9	H						
SV RR333	2	54.21	53.06	102	53.63	102	1823	1887	106	1849	106	350	329	11399	11670	17.98	17.46	33.7	35.6	1.04	1.03	72	71	0	0	4.84	4.84	5.0	4.8	4.4	4.4	5.3	5.1	H						
SV RR351	1	53.73	52.02	100	53.06	101	1783	1877	105	1792	103	337	325	11196	11723	17.91	17.30	33.2	36.1	1.05	1.04	74	73	0	0	4.41	4.46	4.2	4.3	4.2	4.2	5.0	4.9	H						
SX Avalanche RR(856)	1	55.22	53.89	104	54.90	104	1860	1803	101	1761	101	342	331	10472	11077	18.13	17.58	30.6	33.5	1.02	1.00	72	72	9	5	4.64	4.69	4.0	4.2	4.3	4.4	5.8	5.6	H						
SX Canyon RR	2	55.26	53.44	103	53.37	101	1829	1878	105	1812	104	342	330	11330	11574	18.15	17.51	33.1	35.1	1.03	1.02	71	71	0	0	4.92	4.84	4.3	4.3	4.5	4.5	5.1	5.2	H						
SX Cayuga RR	2	48.04	47.03	90	48.53	92	1658	1704	95	1663	97	318	308	11222	11197	17.65	16.68	35.5	38.3	1.13	1.11	77	74	5	2	4.97	5.01	4.8	4.1	4.4	4.5	4.0	3.4	Rzm						
SX Meridian RR(856)	1	54.66	52.81	102	53.90	102	1815	1825	108	1894	109	340	328	11258	11945	18.04	17.43	33.2	36.0	1.02	1.04	72	72	5	2	4.54	4.49	4.5	4.5	4.4	4.4	4.8	4.9	H						
SX Winchester RR	3	51.84	52.22	100	53.49	101	1580	1706	98	1664	96	331	326	10987	10615	17.59	17.29	30.5	32.6	1.04	1.01	68	67	5	2	4.07	4.02	4.4	4.1	4.5	4.6	4.6	4.4	Rzm						
Newly Approved																																								
BTS 8606	NC	54.65	53.10	102	--	--	1882	1941	109	--	--	340	329	11739	12018	18.13	17.54	34.6	36.6	1.09	1.08	79	75	0	0	4.73	4.92	4.9	4.8	5.0	4.7	2.8	2.7	H						
BTS 8629	NC	52.38	50.48	97	--	--	1884</																																	

Table 3. Performance Data of RR Aphanomyces Specialty Varieties - Under Aphanomyces Conditions (Relative to Susceptible Checks) approved for 2018 Growing Season +++																											
Variety	Years Comm	Rev/Ton			Rev/Acre			Rec/Ton			Rec/Acre			Sugar		Yield		CR Rating +			Aph Root +			Rhizoctonia +		Fusarium +	
		2017	2016	%Sus	2017	2016	%Sus	2017	2016	2017	2016	2017	2016	2017	2016	17	16	17	16	2 Yr	17	16	17	16	17	16	
# of locations		0	2	2	0	2	2	0	2	0	2	0	2	0	2	0	2	3	3	1	2	3	1	4	2	2	
Previously Approved																											
BTS 80RR52	6	--	47.73	99	--	1406	137	--	305.0	--	8994	--	16.32	--	29.5	4.37	4.28	4.4	4.1	4.2	4.1	4.4	2.7	2.8			
BTS 8337	3	--	49.32	102	--	1372	134	--	310.0	--	8626	--	16.59	--	27.9	4.36	4.62	3.8	3.3	3.5	4.3	4.1	3.8	4.0			
BTS 8500	1	--	44.32	92	--	1328	130	--	293.9	--	8817	--	15.79	--	30.1	4.29	4.54	4.5	4.2	4.4	4.6	4.4	2.1	1.9			
BTS 8512	1	--	45.42	94	--	1291	126	--	297.6	--	8488	--	15.97	--	28.6	3.69	4.04	3.8	4.2	4.0	4.3	4.4	3.0	2.7			
BTS 8524	1	--	44.53	92	--	1417	138	--	294.6	--	9385	--	15.85	--	31.9	4.38	4.74	4.5	3.9	4.2	4.4	4.2	3.2	3.4			
BTS 8572	1	--	49.62	103	--	1285	125	--	311.6	--	8094	--	16.59	--	26.1	4.14	4.41	3.8	4.5	4.1	4.3	4.5	2.5	2.2			
Crystal 093RR	6	--	49.26	102	--	1380	135	--	309.9	--	8685	--	16.61	--	28.1	4.49	4.95	4.4	4.3	4.4	4.5	4.4	3.5	3.4			
Crystal 101RR	6	--	42.78	89	--	1332	130	--	289.2	--	9012	--	15.70	--	31.2	4.57	4.59	3.9	3.4	3.7	4.8	4.8	2.7	2.4			
Crystal 355RR	2	--	49.37	102	--	1278	125	--	310.2	--	8071	--	16.58	--	26.1	4.36	4.60	4.8	4.5	4.7	4.1	4.0	2.8	2.7			
Crystal 467RR	1	--	42.00	87	--	1244	121	--	286.1	--	8510	--	15.48	--	29.9	4.46	4.69	4.0	4.0	4.0	4.5	4.3	2.0	1.8			
Crystal 573RR	NC	--	48.78	101	--	1303	127	--	308.8	--	8294	--	16.51	--	27.0	4.15	4.35	3.8	4.1	4.0	4.6	4.5	3.1	3.5			
Crystal 574RR	1	--	44.17	92	--	1361	133	--	293.4	--	9003	--	15.76	--	30.5	4.35	4.51	4.7	3.7	4.2	4.2	4.5	2.2	1.8			
Crystal 986RR	6	--	49.30	102	--	1428	139	--	310.0	--	8981	--	16.53	--	29.0	4.77	4.75	4.1	4.4	4.2	4.4	4.4	4.7	4.9			
Hilleshög HL9707	1	--	44.36	92	--	1256	123	--	294.0	--	8345	--	15.78	--	28.4	4.96	4.53	4.7	4.0	4.3	4.4	4.4	4.1	4.9			
Hilleshög 4302RR	4	--	47.43	98	--	1096	107	--	304.0	--	6975	--	16.25	--	22.9	3.93	4.13	6.7	4.6	5.6	3.6	3.7	5.1	5.1			
Hilleshög 9528RR	3	--	48.08	100	--	1379	134	--	306.1	--	8772	--	16.38	--	28.6	4.99	4.73	5.6	3.8	4.7	4.2	4.2	4.2	4.5			
Marbo 109	2	--	51.46	107	--	1180	115	--	316.9	--	7271	--	16.91	--	23.0	4.14	4.14	5.1	4.3	4.7	3.6	3.7	4.2	4.5			
Marbo MA502	1	--	44.36	92	--	1350	132	--	294.0	--	8945	--	15.88	--	30.4	5.01	4.79	3.5	3.1	3.3	4.8	4.7	3.0	1.9			
SV RRR333	2	--	46.56	97	--	1241	121	--	301.2	--	8010	--	16.08	--	26.5	4.84	4.85	5.0	4.7	4.8	4.4	4.4	5.3	4.8			
SV RRR351	1	--	46.82	97	--	1386	135	--	302.2	--	8971	--	16.16	--	29.7	4.41	4.50	4.2	4.4	4.3	4.2	4.2	5.0	4.8			
SX Avalanche RR(858)	1	--	48.30	100	--	1330	130	--	307.2	--	8473	--	16.37	--	27.6	4.64	4.74	4.0	4.4	4.2	4.3	4.5	5.8	5.4			
SX Canyon RR	2	--	44.98	93	--	1201	117	--	296.2	--	7852	--	15.86	--	26.3	4.92	4.76	4.3	4.3	4.3	4.5	4.4	5.1	5.3			
SX Cruze RR	2	--	42.40	88	--	1321	129	--	288.0	--	8957	--	15.51	--	31.0	5.37	4.65	4.8	3.4	4.1	4.4	4.7	4.0	2.8			
SX Winchester RR	3	--	47.53	99	--	1311	128	--	304.3	--	8395	--	16.23	--	27.6	4.07	3.97	4.4	3.9	4.1	4.5	4.6	4.6	4.1			
Newly Approved																											
Crystal 684RR	NC	--	44.83	93	--	1517	148	--	295.6	--	9986	--	15.89	--	33.7	4.34	4.57	4.3	3.7	4.0	4.6	4.4	2.0	1.8			
Hilleshög HL9895	NC	--	46.60	97	--	1344	131	--	301.5	--	8726	--	16.20	--	29.0	4.84	4.49	4.4	3.6	4.0	4.3	4.6	4.1	2.4			
Marbo IM611	NC	--	48.38	100	--	1278	125	--	307.5	--	8119	--	16.50	--	26.4	5.03	4.47	4.0	3.9	4.0	4.4	4.6	3.8	2.0			
SX RRR1863	NC	--	50.16	104	--	1349	132	--	313.4	--	8434	--	16.62	--	26.9	4.08	4.35	4.9	3.6	4.2	4.2	4.5	6.0	5.8			
SV RRR268	NC	--	48.64	101	--	1306	127	--	308.4	--	8262	--	16.40	--	26.7	5.06	5.13	4.7	4.0	4.4	4.6	4.7	5.0	5.2			
Aph Susc Checks		--	48.17		--	1025		--	306.8	--	6529	--	16.49	--	21.3												
Mean of Aph Specialty Varieties		--	46.81		--	1320		--	302.1	--	8533	--	16.18	--	28.3												

%Susc = % of susceptible varieties.
+ Aph ratings from RRV & Shakopee (res.-4-4, susc-5.5). CR from Randolph MN, Foxhome MN & Michigan (res.-4-4, susc-5.5). Fusarium from RRV (res.-3.0, susc-5.0). Rhizoc: from Mhd.
NWROC & Mich (res.-3-3, susc-5). It may perform better under severe Rhz.

+++ 2016 Revenue estimates based on a \$2.44 beet payment at 17.5% sugar and 1.5% loss to molasses. Revenue does not consider hauling or production costs.

++++ 2016 Data from Perley and Cavalier.

++++Lack of Aphanomyces pressure at any of the OVT sites prevented collection of Aphanomyces Yield Data for 2017.

Table 4. Performance Data of Conventional Varieties During 2016 and 2017 Growing Seasons (All Locations Combined)+++

Variety	Yrs Com	Rcv/Ton **		Rcv/Acre **		Rcv/Ton		Rcv/Acre		Sugar		Yield		Molasses		Embrn.		Boiler Ac		CR *		Ash Stock		Rhozc *		Fgs		Rzm		
		17	2 Yr	21%	17	2 Yr	21%	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr	17	2 Yr			
Previous Approved																														
BETA EXP 687	NC	56.11	54.82	121	1633	1781	116	345	334	10123	10894	18.47	17.94	29.6	32.8	1.22	1.22	72	73	0	0	3.99	4.07	4.3	4.6	4.2	4.2	3.5	3.5	H
BETA EXP 698	NC	53.21	52.37	116	1615	1786	116	336	326	10304	11185	17.92	17.45	31.1	34.6	1.14	1.14	76	73	0	0	4.18	4.23	3.6	3.7	4.5	4.4	3.1	2.9	H
Crystal 620	NC	53.86	53.05	117	1706	1825	119	339	329	10783	11322	18.05	17.56	32.2	34.6	1.15	1.13	69	70	0	0	4.14	4.17	4.1	4.2	4.4	4.5	2.8	2.8	H
Crystal 622	NC	54.64	54.57	120	1532	1665	108	340	333	9650	10239	18.26	17.89	28.7	30.9	1.25	1.22	66	66	0	0	3.72	3.84	4.0	4.2	4.5	4.3	3.5	3.6	H
Crystal R761	8	51.12	50.18	111	1691	1749	114	329	319	10896	11128	17.72	17.25	33.2	35.0	1.28	1.28	74	72	0	0	4.93	4.96	4.0	3.8	4.5	4.6	3.2	3.2	H
Heshiq 3035Rz	11	54.33	54.57	120	1457	1617	105	339	333	9182	9906	18.17	17.84	27.3	29.9	1.21	1.18	80	79	18	77	4.42	4.47	5.2	4.8	4.1	4.0	3.7	3.7	Rzm
Heshiq 9891Rz	NC	54.95	53.90	119	1481	1585	103	341	331	9288	9781	18.26	17.77	27.4	29.7	1.19	1.20	77	77	0	0	4.13	4.27	4.9	4.7	4.5	4.3	3.7	3.7	Rzm
Marbo M615Rz	NC	51.71	51.79	114	1586	1778	116	331	324	10191	11127	17.80	17.42	31.0	34.5	1.27	1.20	80	76	0	0	4.81	4.92	5.3	5.0	4.7	4.6	4.7	4.9	Rzm
Seedex 9889	NC	54.07	53.32	118	1741	1874	122	338	329	10942	11385	18.02	17.83	32.5	35.3	1.09	1.08	75	77	0	0	5.21	4.99	5.0	4.8	4.4	4.5	3.8	3.2	H
Seedex Deuce	NC	53.90	53.65	118	1790	1882	122	338	330	11246	11584	18.00	17.58	33.4	35.1	1.10	1.09	75	75	18	18	4.78	4.72	5.0	5.9	4.4	4.5	4.5	4.6	H
SV 48611	NC	55.52	54.71	121	1669	1793	117	343	334	10325	10925	18.30	17.83	30.1	32.8	1.13	1.13	69	68	0	0	5.28	5.06	4.2	4.4	4.3	4.5	5.7	5.5	H
Newly Approved																														
BETA EXP 747	NC	52.59	--	--	1652	--	--	334	--	10556	--	17.83	--	31.9	--	1.15	--	75	--	0	0	4.40	--	3.6	--	3.9	--	4.6	--	H
BETA EXP 758	NC	53.88	--	--	1638	--	--	336	--	10331	--	18.02	--	30.8	--	1.13	--	78	--	0	0	4.52	--	3.3	--	4.3	--	3.9	--	H
Crystal 735	NC	58.13	--	--	1616	--	--	352	--	9832	--	18.69	--	28.1	--	1.09	--	68	--	0	0	4.84	--	3.9	--	4.6	--	3.6	--	H
Crystal 737	NC	53.07	--	--	1555	--	--	337	--	9878	--	18.00	--	29.7	--	1.25	--	69	--	0	0	3.92	--	2.2	--	4.2	--	3.5	--	Rzm
Marbo M6720Rz	NC	55.19	--	--	1586	--	--	342	--	9919	--	18.23	--	29.3	--	1.13	--	84	--	0	0	4.54	--	5.2	--	4.5	--	3.3	--	H
SV 48777	NC	57.39	--	--	1701	--	--	349	--	10409	--	18.48	--	30.0	--	1.02	--	72	--	0	0	4.78	--	4.2	--	4.6	--	4.0	--	H
Strube 13722	NC	50.40	--	--	1696	--	--	326	--	11043	--	17.46	--	34.0	--	1.15	--	79	--	0	0	4.06	--	7.5	--	4.7	--	6.6	--	Rzm
Benchmark var. mean		52.84	51.50		1681	1780		334	324	10653	11056	17.88	17.32	31.9	34.3	1.16	1.14	74	71											

Embrn is % of planted seeds producing a full leaf
 * Ash ratings from Shakopee (res=4.4, suscc=5.5), CR from Randolph MN, Foxhome MN & Michigan (res=4.5, suscc=5.2), Fusarium from RRV (res=3.0, suscc=6.0), Rhozc: from Mkt, NWVROC & Mich (res=3.8, suscc=5). H may perform better under severe Rzm.
 ** 2017 Revenue estimate based on a \$48.48 bush payment (\$11.48) at 17.5% sugar and 1.5% loss to molasses. 2016 Revenue estimate based on a \$52.44 bush payment. Revenue does not consider hauling or production costs.
 *** Sites include Casselton, Ada, Crookston, Grand Forks, St. Thomas in 2016.
 +++ Sites include Casselton, Hendrum, Grand Forks, Grandis, St. Thomas, Humbolt in 2017.
 Boilers (Ac) are based upon a plant stand of 45,000.
 - data not available.

Created 10/31/2017

Table 5. Official Trial Disease Nurseries 2015 - 2017 (Varieties tested in 2017)																						
Cercospora, Aphanomyces, Rhizoctonia, Fusarium & Rhizomania																						
Code	Variety	< 4.5 CR > 5.2					< 4.4 Aph > 5.5					< 3.82 Rhizoctonia > 5.0					< 3.0 Fusarium > 5.0					High Rzm
		17	16	15	2 Yr	3 Yr	17	16	15	2 Yr	3 Yr	17	16	15	2 Yr	3 Yr	17	16	15	2 Yr	3 Yr	
ACSC Commercial																						
529	BTS 80RR52	4.37	4.28	4.11	4.33	4.26	4.36	4.11	3.24	4.23	3.90	4.14	4.41	3.95	4.27	4.17	2.69	2.81	2.83	2.75	2.77	Hi Rzm
545	BTS 8337	4.36	4.62	4.49	4.49	4.49	3.78	3.26	2.55	3.52	3.19	4.30	4.08	3.87	4.19	4.08	3.83	4.01	3.72	3.92	3.85	Hi Rzm
562	BTS 8363	4.10	4.33	3.83	4.21	4.09	4.60	4.93	4.77	4.76	4.76	4.85	4.34	4.12	4.59	4.44	3.49	3.11	2.85	3.30	3.15	Hi Rzm
513	BTS 8500	4.29	4.54	4.45	4.41	4.43	4.52	4.22	3.54	4.37	4.09	4.57	4.43	4.19	4.50	4.40	2.14	1.90	2.41	2.02	2.15	Hi Rzm
533	BTS 8512	3.69	4.04	4.12	3.86	3.95	3.78	4.17	3.91	3.97	3.95	4.28	4.44	4.28	4.36	4.33	2.96	2.71	2.70	2.83	2.79	Hi Rzm
550	BTS 8524	4.38	4.74	4.40	4.56	4.51	4.49	3.89	3.33	4.19	3.90	4.41	4.20	4.14	4.31	4.25	3.24	3.38	2.88	3.31	3.17	Hi Rzm
570	BTS 8572	4.14	4.41	4.60	4.27	4.38	3.76	4.46	4.05	4.11	4.09	4.32	4.54	3.85	4.43	4.24	2.54	2.23	2.54	2.39	2.44	Hi Rzm
549	Crystal 093RR	4.49	4.95	4.76	4.72	4.73	4.43	4.32	3.86	4.38	4.21	4.50	4.37	3.96	4.44	4.28	3.48	3.35	3.22	3.42	3.35	Hi Rzm
551	Crystal 101RR	4.57	4.59	4.65	4.58	4.60	3.92	3.42	3.31	3.67	3.55	4.78	4.78	4.64	4.78	4.73	2.72	2.40	2.64	2.56	2.59	Hi Rzm
507	Crystal 246RR	4.63	4.81	4.49	4.72	4.64	5.13	4.85	4.99	4.99	4.99	4.23	4.32	4.19	4.28	4.25	3.24	3.10	3.00	3.17	3.11	Hi Rzm
560	Crystal 247RR	4.55	4.65	4.19	4.60	4.47	5.35	4.77	4.94	5.06	5.02	4.49	4.32	4.33	4.40	4.38	3.00	2.80	2.51	2.90	2.77	Hi Rzm
565	Crystal 355RR	4.36	4.60	4.43	4.48	4.46	4.84	4.46	3.26	4.65	4.19	4.09	3.96	NE	4.02	NE	2.76	2.65	NE	2.71	NE	Hi Rzm
523	Crystal 467RR	4.46	4.69	4.34	4.57	4.49	3.96	4.04	3.55	4.00	3.85	4.47	4.26	3.97	4.37	4.23	1.98	1.84	2.46	1.91	2.09	Hi Rzm
503	Crystal 572RR	4.27	4.57	4.65	4.42	4.50	4.69	4.74	4.33	4.71	4.59	4.47	4.21	3.89	4.34	4.19	2.64	1.82	2.36	2.23	2.27	Hi Rzm
544	Crystal 574RR	4.35	4.51	4.30	4.43	4.39	4.72	3.69	2.93	4.21	3.78	4.16	4.47	4.16	4.31	4.26	2.23	1.82	2.00	2.02	2.02	Hi Rzm
532	Crystal 986RR	4.77	4.75	4.97	4.76	4.83	4.09	4.41	3.87	4.25	4.12	4.39	4.38	4.06	4.38	4.28	4.73	4.86	3.89	4.79	4.49	Hi Rzm
505	Hilleshög 4302RR	3.93	4.13	4.13	4.03	4.06	6.66	4.63	4.02	5.65	5.10	3.60	3.65	3.70	3.63	3.65	5.09	5.09	4.05	5.09	4.74	Rzm
542	Hilleshög 4448RR	5.28	5.21	5.29	5.24	5.26	6.29	3.90	2.80	5.09	4.33	4.63	4.51	3.92	4.57	4.35	5.35	5.26	NE	5.30	NE	Rzm
531	Hilleshög 9528RR	4.99	4.73	5.16	4.86	4.96	5.63	3.77	2.97	4.70	4.12	4.21	4.21	4.10	4.21	4.18	4.25	4.52	4.00	4.39	4.26	Hi Rzm
559	Hilleshög HL9707	4.96	4.53	4.60	4.74	4.70	4.70	3.99	3.52	4.34	4.07	4.43	4.40	4.21	4.41	4.35	4.09	4.88	3.68	4.49	4.22	Hi Rzm
556	Maribo 109	4.14	4.14	4.56	4.14	4.28	5.06	4.27	3.54	4.66	4.29	3.63	3.69	3.67	3.66	3.66	4.23	4.50	3.58	4.37	4.11	Hi Rzm
539	Maribo 305	4.98	4.72	4.76	4.85	4.82	5.67	4.42	4.76	5.05	4.95	4.60	4.40	3.83	4.50	4.28	5.89	5.89	5.02	5.89	5.60	Rzm
526	Maribo MA502	5.01	4.79	5.04	4.90	4.95	3.53	3.06	2.93	3.29	3.17	4.78	4.73	4.14	4.76	4.55	3.02	1.92	2.33	2.47	2.42	Hi Rzm
537	SX Avalanche RR(858)	4.64	4.74	4.15	4.69	4.51	4.00	4.44	3.40	4.22	3.95	4.29	4.52	4.21	4.40	4.34	5.75	5.38	5.12	5.57	5.42	Rzm
548	SX Canyon RR	4.92	4.76	4.02	4.84	4.56	4.33	4.28	3.59	4.31	4.07	4.51	4.40	4.22	4.45	4.38	5.12	5.26	3.85	5.19	4.74	Rzm
535	SX Cruze RR	5.37	4.65	4.57	5.01	4.87	4.79	3.41	4.14	4.10	4.11	4.39	4.69	4.18	4.54	4.42	3.98	2.80	NE	3.39	NE	Rzm
519	SX Marathon RR(856)	4.54	4.44	5.37	4.49	4.78	4.52	4.38	4.53	4.45	4.48	4.40	4.47	4.16	4.43	4.34	4.84	4.90	4.87	4.87	4.87	Rzm
575	SX Winchester RR	4.07	3.97	3.67	4.02	3.90	4.36	3.85	3.07	4.11	3.76	4.47	4.63	4.28	4.55	4.46	4.64	4.11	3.95	4.38	4.23	Rzm
564	SV RR244TT	4.85	4.46	4.17	4.65	4.49	4.91	4.97	4.23	4.94	4.70	4.50	4.45	4.18	4.48	4.38	3.74	4.14	3.86	3.94	3.91	Hi Rzm
541	SV RR333	4.84	4.85	4.54	4.84	4.74	4.99	4.71	3.46	4.85	4.39	4.44	4.44	4.11	4.44	4.33	5.35	4.84	NE	5.09	NE	Hi Rzm
573	SV RR351	4.41	4.50	4.62	4.46	4.51	4.18	4.38	3.53	4.28	4.03	4.25	4.17	NE	4.21	NE	4.96	4.75	NE	4.86	NE	Hi Rzm
ACSC Experimental																						
509	BTS 8606	4.73	5.12	--	4.92	--	4.91	4.60	--	4.75	--	5.00	4.48	--	4.74	--	2.81	2.69	--	2.75	--	Hi Rzm
525	BTS 8629	4.29	4.59	--	4.44	--	4.68	4.14	--	4.41	--	4.21	3.73	--	3.97	--	4.20	4.04	--	4.12	--	Hi Rzm
577	BTS 8735	4.22	--	--	--	--	4.74	--	--	--	--	4.38	--	--	--	--	3.93	--	--	--	--	Hi Rzm
506	BTS 8742	4.36	--	--	--	--	5.02	--	--	--	--	4.23	--	--	--	--	2.59	--	--	--	--	Hi Rzm
536	BTS 8749	4.05	--	--	--	--	3.53	--	--	--	--	3.95	--	--	--	--	3.28	--	--	--	--	Hi Rzm
540	BTS 8756	4.01	--	--	--	--	5.23	--	--	--	--	4.34	--	--	--	--	2.67	--	--	--	--	Hi Rzm
521	BTS 8767	4.16	--	--	--	--	4.80	--	--	--	--	4.75	--	--	--	--	2.71	--	--	--	--	Hi Rzm
518	BTS 8770	4.30	--	--	--	--	4.97	--	--	--	--	4.57	--	--	--	--	2.82	--	--	--	--	Hi Rzm
567	BTS 8784	3.65	--	--	--	--	4.59	--	--	--	--	4.64	--	--	--	--	2.63	--	--	--	--	Hi Rzm
502	BTS 8787	4.03	--	--	--	--	4.71	--	--	--	--	4.31	--	--	--	--	2.50	--	--	--	--	Hi Rzm
512	BTS 8798	4.30	--	--	--	--	4.92	--	--	--	--	4.52	--	--	--	--	3.37	--	--	--	--	Hi Rzm
554	Crystal 573RR	4.15	4.35	4.15	4.25	4.22	3.84	4.06	3.69	3.95	3.86	4.57	4.55	4.25	4.56	4.45	3.10	3.49	3.02	3.29	3.20	Hi Rzm
571	Crystal 578RR	4.91	4.87	4.93	4.89	4.91	4.56	4.44	4.52	4.50	4.51	4.40	4.32	4.03	4.36	4.25	2.41	1.99	2.42	2.20	2.27	Hi Rzm
510	Crystal 684RR	4.34	4.57	--	4.45	--	4.31	3.74	--	4.02	--	4.57	4.41	--	4.49	--	2.01	1.76	--	1.89	--	Hi Rzm
547	Crystal 792RR	3.94	--	--	--	--	4.73	--	--	--	--	3.88	--	--	--	--	2.81	--	--	--	--	Hi Rzm
557	Crystal 793RR	3.93	--	--	--	--	3.02	--	--	--	--	4.26	--	--	--	--	2.95	--	--	--	--	Hi Rzm
534	Crystal 794RR	4.92	--	--	--	--	4.65	--	--	--	--	4.15	--	--	--	--	2.45	--	--	--	--	Hi Rzm
522	Crystal 795RR	4.39	--	--	--	--	4.40	--	--	--	--	3.94	--	--	--	--	2.66	--	--	--	--	Hi Rzm
553	Crystal 796RR	4.85	--	--	--	--	3.11	--	--	--	--	4.23	--	--	--	--	2.34	--	--	--	--	Hi Rzm
528	Crystal 797RR	4.17	--	--	--	--	5.21	--	--	--	--	4.26	--	--	--	--	3.18	--	--	--	--	Hi Rzm
576	Hilleshög HL9708	4.61	4.74	5.04	4.68	4.80	5.94	4.82	4.69	5.38	5.15	4.21	4.28	4.04	4.25	4.18	4.61	4.29	3.69	4.45	4.20	Hi Rzm
561	Hilleshög HL9895	4.84	4.49	--	4.67	--	4.39	3.65	--	4.02	--	4.34	4.56	--	4.45	--	4.15	2.40	--	3.27	--	Hi Rzm
566	Hilleshög HL9920	4.89	--	--	--	--	4.94	--	--	--	--	4.48	--	--	--	--	5.92	--	--	--	--	Hi Rzm
563	Hilleshög HL9921	4.47	--	--	--	--	5.41	--	--	--	--	3.85	--	--	--	--	4.66	--	--	--	--	Hi Rzm
504	Hilleshög HL9922	4.02	--	--	--	--	5.79	--	--	--	--	4.39	--	--	--	--	4.49	--	--	--	--	Hi Rzm
543	Hilleshög HL9923	4.81	--	--	--	--	5.06	--	--	--	--	4.58	--	--	--	--	5.29	--	--	--	--	Hi Rzm
517	Hilleshög HL9924	4.09	--	--	--	--	5.37	--	--	--	--	4.62	--	--	--	--	4.58	--	--	--	--	Hi Rzm
514	Maribo MA504	5.50	5.04	5.25	5.27	5.26	6.20	4.54	4.60	5.37	5.11	4.37	4.58	3.98	4.47	4.31	4.52	4.60	4.11	4.56	4.41	Hi Rzm
568	Maribo MA611	5.03	4.47	--	4.75	--	4.00	3.94	--	3.97	--	4.44	4.63	--	4.53	--	3.78	1.96	--	2.87	--	Hi Rzm
574	Maribo MA717	4.85	--	--	--	--	5.31	--	--	--	--	4.28	--	--	--	--	4.95	--	--	--	--	Rzm
530	Maribo MA718	4.39	--	--	--	--	4.46	--	--	--	--	4.13	--	--	--	--	4.61	--	--	--	--	Rzm
538	Maribo MA719	4.41	--	--	--	--	4.75	--	--	--	--	4.28	--	--	--	--	5.76	--	--	--	--	Rzm

558	SX RR1861	4.74	4.52	--	4.63	--	5.71	4.40	--	5.05	--	4.50	4.59	--	4.55	--	5.05	4.75	--	4.90	--	Hi Rzm
527	SX RR1863	4.08	4.35	--	4.21	--	4.88	3.55	--	4.21	--	4.23	4.54	--	4.39	--	6.04	5.80	--	5.92	--	Hi Rzm
516	SX RR1875	4.06	--	--	--	--	4.13	--	--	--	--	4.34	--	--	--	--	3.57	--	--	--	--	Hi Rzm
520	SX RR1876	4.31	--	--	--	--	4.73	--	--	--	--	4.42	--	--	--	--	3.85	--	--	--	--	Hi Rzm
569	SX RR1877	4.62	--	--	--	--	3.84	--	--	--	--	4.42	--	--	--	--	4.21	--	--	--	--	Hi Rzm
552	SX RR1878	4.71	--	--	--	--	5.54	--	--	--	--	4.31	--	--	--	--	5.03	--	--	--	--	Hi Rzm
524	SX RR1879	4.88	--	--	--	--	4.18	--	--	--	--	4.36	--	--	--	--	4.64	--	--	--	--	Hi Rzm
511	SV RR265	5.19	5.00	--	5.09	--	5.35	4.54	--	4.95	--	4.42	4.44	--	4.43	--	5.32	5.26	--	5.29	--	Hi Rzm
555	SV RR266	4.61	4.74	--	4.67	--	5.64	4.62	--	5.13	--	4.39	4.20	--	4.30	--	5.64	5.18	--	5.41	--	Hi Rzm
572	SV RR268	5.06	5.13	--	5.10	--	4.71	4.00	--	4.36	--	4.57	4.70	--	4.63	--	5.01	5.20	--	5.11	--	Hi Rzm
515	SV RR371	4.59	--	--	--	--	4.55	--	--	--	--	4.31	--	--	--	--	4.91	--	--	--	--	Hi Rzm
501	SV RR372	4.23	--	--	--	--	4.42	--	--	--	--	4.47	--	--	--	--	4.19	--	--	--	--	Hi Rzm
508	SV RR373	4.31	--	--	--	--	4.93	--	--	--	--	4.38	--	--	--	--	5.17	--	--	--	--	Hi Rzm
578	SV RR374	4.71	--	--	--	--	5.20	--	--	--	--	4.30	--	--	--	--	4.44	--	--	--	--	Hi Rzm
546	SV RR375	5.08	--	--	--	--	4.54	--	--	--	--	4.25	--	--	--	--	5.44	--	--	--	--	Hi Rzm
ACSC Conventional																						
807	BETA EXP 687	3.99	4.14	--	4.07	--	4.30	4.88	--	4.59	--	4.20	4.16	--	4.18	--	3.51	3.41	--	3.46	--	Hi Rzm
808	BETA EXP 698	4.18	4.27	--	4.23	--	3.62	3.69	--	3.65	--	4.45	4.35	--	4.40	--	3.06	2.74	--	2.90	--	Hi Rzm
810	BETA EXP 747	4.40	--	--	--	--	3.60	--	--	--	--	3.93	--	--	--	--	4.58	--	--	--	--	Hi Rzm
817	BETA EXP 758	4.52	--	--	--	--	3.29	--	--	--	--	4.31	--	--	--	--	3.91	--	--	--	--	Hi Rzm
811	Crystal 620	4.14	4.19	--	4.17	--	4.09	4.28	--	4.18	--	4.37	4.54	--	4.45	--	2.79	2.73	--	2.76	--	Hi Rzm
801	Crystal 622	3.72	3.96	--	3.84	--	4.05	4.36	--	4.20	--	4.49	4.14	--	4.31	--	3.53	3.57	--	3.55	--	Hi Rzm
814	Crystal 735	4.44	--	--	--	--	3.93	--	--	--	--	4.61	--	--	--	--	3.62	--	--	--	--	Hi Rzm
806	Crystal 737	3.92	--	--	--	--	2.25	--	--	--	--	4.25	--	--	--	--	3.52	--	--	--	--	Hi Rzm
819	Crystal R761	4.93	4.99	--	4.96	--	4.01	3.57	--	3.79	--	4.54	4.57	--	4.55	--	3.23	3.25	--	3.24	--	Hi Rzm
805	Hilleshög 3035Rz	4.42	4.53	--	4.47	--	5.18	4.40	--	4.79	--	4.07	3.93	--	4.00	--	3.70	3.65	--	3.67	--	Rzm
812	Hilleshög 9891Rz	4.13	4.42	--	4.27	--	4.89	4.45	--	4.67	--	4.46	4.22	--	4.34	--	3.66	3.76	--	3.71	--	Rzm
818	Maribo MA615Rz	4.81	5.04	--	4.92	--	5.30	4.80	--	5.05	--	4.73	4.54	--	4.63	--	4.72	5.11	--	4.92	--	Rzm
816	Maribo MA720Rz	4.54	--	--	--	--	5.15	--	--	--	--	4.55	--	--	--	--	3.31	--	--	--	--	Rzm
809	Seedex 8869	5.21	4.76	--	4.99	--	4.99	4.70	--	4.85	--	4.40	4.67	--	4.53	--	3.53	2.92	--	3.23	--	Rzm
802	Seedex Deuce	4.76	4.68	--	4.72	--	6.04	5.70	--	5.87	--	4.39	4.66	--	4.52	--	4.54	4.68	--	4.61	--	Rzm
815	SV 48611	5.28	4.85	--	5.06	--	4.25	4.47	--	4.36	--	4.35	4.66	--	4.50	--	5.74	5.24	--	5.49	--	Hi Rzm
803	SV 48777	4.76	--	--	--	--	4.20	--	--	--	--	4.59	--	--	--	--	3.96	--	--	--	--	Rzm
813	Strube 12720	5.65	--	--	--	--	8.11	--	--	--	--	4.59	--	--	--	--	5.60	--	--	--	--	Rzm
804	Strube 13722	4.06	--	--	--	--	7.54	--	--	--	--	4.73	--	--	--	--	6.63	--	--	--	--	Rzm
CR ratings on a scale of 1-9.																						
Aph root ratings on a scale of 1-9.																						
Rhizoctonia ratings on a scale of 1-7.																						
Fusarium ratings on a scale of 1-9.																						
NE indicates variety was not entered into disease nursery.																						
Hi Rzm = may perform better under severe Rzm.																						
Created 11/8/2017																						

Table 6. Planting & Harvest Dates, Previous Crop and Disease Levels for 2017 ACSC Official Trial Sites *

Location	District / Trial Type	Cooperator	Planting Date	Harvest Date	Preceding Crop	Soil Type	Diseases Present @						Comments
							Aph	Rhc	Rzm	Fus	Maggot	Rt Aphid	
Cassellon ND	Mhd/Hb	Todd Weber	5/1	9/8	Barley	Medium/Light	N	N	N	N	N	N	Late emergence, Cow Harvested Only
Felton MN	Mhd/Hb	Menholt Farms	4/22	10/19	Wheat	Medium/Light	N	N	N	N	N	N	Uniform
Georgetown MN	Mhd/Hb	Hoff Farms	5/9	10/18	Soybeans	Medium	L	L	N	N	N	N	AP in NE
Hendrum MN	EGF/Crk	Mark Maring	4/17	10/15	Wheat	Medium	N	L-M	L-M	N	N	N	Uniform Very little disease
Hillsboro ND	Mhd/Hb	Cotton Farms	4/22	10/16	Wheat	Medium	N	L	L-M	N	N	N	Some Weak Stands
Climax MN	EGF/Crk	Curt Knutson	5/10	10/13	Wheat	Medium	L	M	L	N	L	L	Nursery Abandoned
Grand Forks ND	EGF/Crk	Drees Farming Associaior	5/5	10/10	Wheat	Medium/Light	L	M	L	N	N	N	Slight water stunting, Some weaker stand
Scandia MN	EGF/Crk	Dennis Deboer	5/2	9/13	Wheat	Medium	L-M	L	M	N	N	N	Some Weak Stands
Stephen MN	EGF/Crk	Hvidsten Farms	5/4	10/6	Wheat	Medium/Light	N	L	L	N	L	N	Uniform Canopy, Some Wilting in August
Argyle MN	EGF/Crk	Brent Rippelle	5/3	Abandon	Wheat	Medium/Heavy	L	M	M	NA	NA	L	Abandoned
St Thomas ND	Dtn	Kennelly Farms	5/8	9/30	Wheat	Medium/Light	L	L	L	N	N	N	Gappy due to Hail Damage
Humboldt MN	Dtn	Waise Farms	5/8	10/5	Wheat	Medium/Heavy	L	L	L	N	N	N	Cow Harvested Only
Bathgala ND	Dtn	Shady Bend Farms	5/7	10/1	Wheat	Medium/Heavy	L-M	M	N	L	L	L	Shorter Yellow Canopy
Mhd Rhc-S	Rhc Nurs	Jon Hickel	5/11	Abandon	Soybeans	Medium/Heavy	L	V	N	L	N	N	
Mhd Rhc-E	Rhc Nurs	Jon Hickel	5/11	Abandon	Soybeans	Medium/Heavy	L	L-M	N	L	N	N	
Mhd Rhc-W	Rhc Nurs	Jon Hickel	5/11	Abandon	Soybeans	Medium/Heavy	L	L-M	N	L-M	N	N	
NWROC Rhc	Rhc Nurs	Albert Sims	5/10	8/30	Soybeans	Medium	N	L-M	N	N	N	N	
SSDF Rhc	Rhc Nurs	Mich McCreath	5/11	8/16	NA	NA	NA	NA	NA	NA	NA	NA	
Mhd SE Fus	Fusarium	Ernie Oberg	5/11	7/18	Soybeans	Medium	NA	L	N	V	NA	NA	
Mhd Fus	Fusarium	Kevin Nelson	5/12	7/20	Soybeans	Medium	NA	N	N	V	NA	NA	
Shakopee MN	Aph Nurs	Patrick O'Boyle	5/9	8/30	NA	NA	NA	NA	NA	NA	NA	NA	
Longmont CO	RA Nurs	Eric Runkle	4/20	10/10	NA	NA	NA	NA	NA	NA	NA	NA	
Fonthome CR	Cercospora	Kevin Etzler	5/9	8/29	Wheat	Medium	NA	L-M	NA	L	NA	NA	
SSDF CR	CR Nurs	Mich McCreath	5/11	8/30	NA	NA	NA	NA	NA	NA	NA	NA	
Randolph MN CR	Cercospora	Patrick O'Boyle	5/5	8/9	NA	Medium/Light	NA	NA	NA	NA	NA	NA	

* Fertilizer applied in accordance to cooperative recommendations.

@ Disease notes for Aph, Rhizoc., Rhizomania, Fusarium, Root Maggot and Root Aphids were based upon visual evaluations (N=none, L=light, M=moderate, V=severe, NA=not observed)

Created 10-31-2017

Table 7. Seed Treatments Used on Varieties in ACSC Official Trials in 2017

Variety	Years in Trial	Years ** Comm.	Fungicide (Rhizoctonia)	Insecticide Spring Tails & Maggots	Tachigaren Rate (Aphanomyces)	Priming (Emergence)	Fungicide (Damping Off)
ACSC Commercial							
BTS 80RR52	8	6	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8337	5	3	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8363	5	3	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8500	3	1	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8512	3	1	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8524	3	1	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8572	3	1	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
Crystal 093RR	8	6	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 101RR	7	6	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 246RR	6	4	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 247RR	6	4	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 355RR	5	2	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 467RR	4	1	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 572RR	3	1	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 574RR	3	1	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 986RR	9	6	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Hilleshög 4302RR	7	4	Vibrance	Cruiser Maxx	0	XBEEET	Apron XL Maxim
Hilleshög 4448RR	6	4	Vibrance	Cruiser Maxx	0	XBEEET	Apron XL Maxim
Hilleshög 9528RR	5	3	Vibrance	Cruiser Maxx	0	XBEEET	Apron XL Maxim
Hilleshög HIL9707	3	1	Vibrance	Cruiser Maxx	45	XBEEET	Apron XL Maxim
Maribo 109	4	2	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo 305	5	2	Vibrance	Cruiser Maxx	20	XBEEET	Apron XL Maxim
Maribo MA502	3	1	Vibrance	Cruiser Maxx	20	XBEEET	Apron XL Maxim
SX Avalanche RR(858)	3	1	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SX Canyon RR	4	2	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SX Cruze RR	4	2	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SX Marathon RR(856)	3	1	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SX Winchester RR	5	3	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SV RR244TT	4	2	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SV RR333	5	2	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
SV RR351	3	1	Metlock/Rizolex/Kabina 7g	NipsIt	20	XBEEET	Sebring Thiram
ACSC Experimental							
BTS 8606	2	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8629	2	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8735	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8742	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8749	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8756	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8767	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8770	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8784	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8787	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BTS 8798	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
Crystal 573RR	3	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 578RR	3	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 684RR	2	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 792RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 793RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 794RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 795RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 796RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Crystal 797RR	1	NC	Kabina 14g	Poncho Beta	45	XBEEET	Allegiance Thiram
Hilleshög HIL9708	3	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9895	2	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9920	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9921	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9922	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9923	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög HIL9924	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA504	3	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA611	2	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA717	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA718	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA719	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
SX RR1861	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1863	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1875	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1876	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1877	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1878	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SX RR1879	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR265	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR266	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR268	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR371	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR372	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR373	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR374	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV RR375	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram

Conventional							
BETA EXP 687	2	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BETA EXP 698	2	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BETA EXP 747	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
BETA EXP 758	1	NC	Systiva	Poncho Beta	35	Ultipro	Allegiance Thiram
Crystal 620	2	NC	Kabina 14g	Poncho Beta	45	XBEET	Allegiance Thiram
Crystal 622	2	NC	Kabina 14g	Poncho Beta	45	XBEET	Allegiance Thiram
Crystal 735	1	NC	Kabina 14g	Poncho Beta	45	XBEET	Allegiance Thiram
Crystal 737	1	NC	Kabina 14g	Poncho Beta	45	XBEET	Allegiance Thiram
Crystal R761	11	8	Kabina 14g	Poncho Beta	45	XBEET	Allegiance Thiram
Hilleshög 3035Rz	13	11	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Hilleshög 9891Rz	2	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA615Rz	2	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Maribo MA720Rz	1	NC	Vibrance	Cruiser Maxx	20	NA	Apron XL Maxim
Seedex 8869	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
Seedex Deuce	10	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV 48611	2	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
SV 48777	1	NC	Kabina 14g	NipsIt	20	NA	Sebring Thiram
Strube 12720	1	NC	NA	Poncho Beta	14	3D Plus	Thiram
Strube 13722	1	NC	NA	Poncho Beta	14	3D Plus	Thiram
NA indicates no treatment applied in this category.							Created 11/9/2017

Table 8. 2017 Performance of All RR Varieties - ACSC Official Trial

		10 sites																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %	Tare %	
Commercial Trial																			
BTS 80RR52	103	334.2	100	10789	101	1.22	52.79	100	1699	101	17.94	32.39	206	1650	432	5	78.2	5.5	
BTS 8337	116	349.5	105	11209	105	1.08	57.43	109	1842	110	18.55	32.08	174	1574	352	5	76.0	6.3	
BTS 8363	130	328.7	98	11391	107	1.09	51.14	97	1770	105	17.53	34.70	214	1555	354	0	77.9	5.9	
BTS 8500	119	335.7	100	11741	110	1.11	53.24	101	1862	111	17.90	34.98	192	1606	366	0	76.2	5.5	
BTS 8512	109	339.0	102	110921	103	1.08	54.51	103	1749	104	18.08	32.19	182	1557	359	14	79.5	5.5	
BTS 8524	101	330.0	99	11026	108	1.14	51.51	97	1796	107	17.64	34.88	197	1697	362	5	78.7	6.1	
BTS 8572	111	346.7	104	11147	105	1.07	56.57	107	1817	108	18.41	32.19	159	1501	377	0	77.6	6.0	
Crystal 093RR	115	350.3	105	11339	106	1.08	57.65	109	1866	111	18.60	32.40	153	1575	366	0	76.4	5.8	
Crystal 101RR	107	329.3	98	11040	104	1.20	51.29	97	1718	102	17.66	33.57	238	1768	376	0	76.9	5.7	
Crystal 246RR	108	331.7	99	11322	106	1.09	52.05	99	1775	106	17.67	34.16	226	1553	349	0	74.3	6.2	
Crystal 247RR	131	335.0	100	11675	109	1.03	53.09	100	1832	109	17.79	34.57	202	1568	305	18	76.1	5.7	
Crystal 355RR	104	340.0	102	10689	100	1.15	54.56	103	1711	102	18.16	31.53	195	1601	400	0	75.1	6.3	
Crystal 467RR	129	330.1	99	11588	109	1.12	51.56	98	1804	107	17.63	35.22	256	1647	338	0	79.6	5.7	
Crystal 572RR	126	354.7	106	11379	107	1.01	58.99	112	1891	112	18.74	32.12	148	1456	341	0	81.5	6.5	
Crystal 574RR	110	334.4	100	11851	111	1.08	52.84	100	1875	112	17.79	35.41	175	1587	346	0	79.5	5.2	
Crystal 986RR	128	341.1	102	11008	103	1.03	54.89	104	1776	106	18.09	32.18	210	1440	339	0	78.4	6.2	
Hilleshög 4302RR	105	334.0	100	10093	95	1.05	52.73	100	1597	95	17.75	30.13	223	1584	310	0	85.2	6.2	
Hilleshög 4448RR	112	338.0	101	11456	108	1.06	53.93	102	1829	109	17.97	33.89	191	1516	351	5	69.8	5.1	
Hilleshög 9528RR	123	339.3	101	11154	105	1.05	54.35	103	1785	106	18.02	32.88	193	1512	343	5	74.1	5.4	
Hilleshög HIL9707	114	324.3	97	11020	103	1.14	49.79	94	1692	101	17.35	33.98	226	1598	375	5	71.8	5.7	
Maribo 109	122	347.6	104	9579	90	1.06	56.86	108	1569	93	18.43	27.50	184	1495	355	0	67.5	5.8	
Maribo 305	106	331.7	99	11018	103	1.02	52.03	98	1731	103	17.60	33.15	194	1457	333	0	66.8	5.0	
Maribo MA502	121	329.8	99	10539	99	1.17	51.46	97	1642	98	17.66	32.02	257	1672	370	68	73.6	5.7	
SX Avalanche RR(858)	127	342.2	102	10472	98	1.02	55.22	104	1690	101	18.13	30.60	200	1526	310	9	71.6	6.3	
SX Canyon RR	118	342.4	102	11330	106	1.03	55.26	105	1829	109	18.15	33.11	171	1534	331	0	71.4	5.8	
SX Cruze RR	117	318.4	95	11272	106	1.13	48.01	91	1696	101	17.05	35.47	202	1564	388	5	76.8	5.7	
SX Marathon RR(856)	102	340.4	102	11296	106	1.02	54.66	103	1812	108	18.04	33.22	170	1534	323	5	71.7	6.1	
SX Winchester RR	125	331.1	99	10087	95	1.04	51.84	98	1580	94	17.59	30.46	195	1566	320	5	65.7	5.8	
SV RR244TT	120	334.7	100	11339	106	1.05	52.93	100	1796	107	17.79	33.83	178	1577	331	5	71.9	5.7	
SV RR333	124	338.9	101	11399	107	1.04	54.21	103	1823	108	17.98	33.65	178	1556	325	0	72.4	5.8	
SV RR351	113	337.3	101	11196	105	1.05	53.73	102	1783	106	17.91	33.19	185	1549	332	0	73.9	5.8	
RR Filler #01s	132	330.8	99	11182	105	1.19	51.76	98	1747	104	17.73	33.86	234	1738	375	0	78.6	6.0	
RR Filler #01v	133	330.2	99	11242	106	1.20	51.58	98	1754	104	17.71	34.10	234	1753	376	0	78.0	5.6	
Experimental Trial (Comm status)																			
BTS 8606	242	340.5	102	11739	110	1.09	54.65	103	1882	112	18.13	34.57	207	1576	356	0	78.9	4.8	
BTS 8629	224	332.8	100	11986	113	1.08	52.38	99	1884	112	17.72	36.12	215	1450	377	0	81.2	4.1	
BTS 8735	234	335.7	100	11581	109	1.05	53.23	101	1836	109	17.84	34.49	199	1421	364	0	79.2	3.8	
BTS 8742	207	333.4	100	10461	98	1.23	52.55	99	1646	98	17.89	31.40	210	1624	443	0	76.4	4.7	
BTS 8749	202	337.7	101	10812	101	1.14	53.82	102	1719	102	18.04	32.11	203	1629	381	9	77.3	4.9	
BTS 8756	241	338.4	101	10818	102	1.25	54.06	102	1724	103	18.16	32.02	202	1698	443	0	81.6	5.0	
BTS 8767	235	339.2	101	11755	110	1.08	54.27	103	1878	112	18.05	34.75	203	1590	342	0	81.4	5.0	
BTS 8770	247	337.4	101	11328	106	1.07	53.72	102	1801	107	17.95	33.62	211	1609	325	9	73.2	4.4	
BTS 8784	236	351.4	105	10874	102	1.04	57.86	109	1787	106	18.62	31.00	160	1451	359	0	77.3	4.4	
BTS 8787	219	331.5	99	11071	104	1.09	52.00	98	1733	103	17.68	33.45	198	1583	351	0	73.4	4.5	
BTS 8798	223	338.8	101	10627	100	1.07	54.16	103	1695	101	18.03	31.42	166	1485	377	0	80.1	5.1	
Crystal 573RR	225	343.9	103	11039	104	1.08	55.66	105	1785	106	18.28	32.14	176	1511	368	0	75.0	4.4	
Crystal 578RR	220	338.4	101	11908	112	1.07	54.05	102	1899	113	18.00	35.28	206	1579	335	0	80.1	4.6	
Crystal 684RR	239	333.7	100	12057	113	1.12	52.65	100	1899	113	17.81	36.22	216	1658	354	0	80.0	4.9	
Crystal 792RR	218	344.0	103	11139	105	1.05	55.67	105	1799	107	18.26	32.45	161	1485	367	0	77.6	4.2	
Crystal 793RR	246	347.5	104	11636	109	1.00	56.69	107	1896	113	18.39	33.52	172	1441	329	18	78.0	4.8	
Crystal 794RR	208	333.8	100	11629	109	1.09	52.66	100	1835	109	17.79	34.82	206	1534	368	0	70.7	4.1	
Crystal 795RR	215	340.1	102	10685	100	1.13	54.53	103	1708	102	18.14	31.53	191	1557	395	0	75.7	4.6	
Crystal 796RR	238	337.0	101	12237	115	1.07	53.63	101	1950	116	17.93	36.27	190	1570	342	0	80.0	5.0	
Crystal 797RR	201	330.1	99	11595	109	1.11	51.58	98	1809	108	17.63	35.22	233	1660	339	9	69.6	4.1	
Hilleshög HIL9708	222	338.6	101	10290	97	1.07	54.11	102	1640	98	18.02	30.41	214	1493	364	9	74.3	3.7	
Hilleshög HIL9895	209	326.3	98	10024	94	1.17	50.46	95	1547	92	17.48	30.84	223	1604	400	41	73.1	4.3	
Hilleshög HIL9920	203	347.2	104	10968	103	1.02	56.61	107	1785	106	18.40	31.64	190	1574	304	9	74.1	4.4	
Hilleshög HIL9921	204	345.2	103	9779	92	1.08	56.04	106	1585	94	18.35	28.35	205	1481	368	0	76.5	4.2	
Hilleshög HIL9922	231	325.4	97	10144	95	1.18	50.19	95	1560	93	17.44	31.26	215	1627	403	68	78.0	4.8	
Hilleshög HIL9923	243	337.5	101	9412	88	1.24	53.77	102	1497	89	18.11	27.92	248	1644	441	0	62.1	3.8	
Hilleshög HIL9924	237	335.0	100	9186	86	1.27	53.00	100	1455	87	18.02	27.42	219	1622	482	0	57.5	4.6	
Maribo MA504	229	333.9	100	11632	109	1.07	52.70	100	1830	109	17.77	34.93	210	1592	357	0	76.9	3.4	
Maribo MA611	245	325.9	97	10000	94	1.16	50.37	95	1542	92	17.45	30.75	225	1623	378	0	75.3	3.9	
Maribo MA717	232	342.0	102	10828	102	1.08	55.10</												

Table 9. 2017 Performance of All RR Varieties - ACSC Official Trial

Felton MN																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	298.0	98	11136	102	1.95	41.82	95	1578	100	16.83	37.05	552	1779	842	0	58.3
BTS 8337	116	340.1	111	12185	111	1.51	54.57	124	1953	123	18.55	35.96	305	1598	626	0	60.8
BTS 8363	130	306.9	100	12019	110	1.67	44.51	101	1746	110	17.02	39.03	483	1698	669	0	64.6
BTS 8500	119	325.8	107	12953	118	1.58	50.23	114	1994	126	17.87	39.89	357	1618	658	0	65.3
BTS 8512	109	321.8	105	11792	108	1.49	49.03	111	1791	113	17.61	36.59	351	1565	602	32	60.4
BTS 8524	101	315.0	103	12812	117	1.58	46.98	107	1913	121	17.35	40.62	407	1728	614	0	63.8
BTS 8572	111	329.4	108	11975	109	1.55	51.35	116	1874	118	17.98	36.28	286	1557	685	0	62.2
Crystal 093RR	115	329.8	108	12237	112	1.49	51.46	117	1907	120	18.03	37.21	319	1625	597	0	57.8
Crystal 101RR	107	303.5	99	11549	106	1.69	43.51	99	1655	104	16.87	38.23	456	1816	659	0	64.0
Crystal 246RR	108	315.0	103	12362	113	1.65	46.99	107	1849	117	17.39	39.13	491	1629	664	0	57.9
Crystal 247RR	131	322.1	105	12591	115	1.44	49.12	111	1924	121	17.56	38.88	418	1661	524	32	65.9
Crystal 355RR	104	312.1	102	11247	103	1.70	46.10	105	1667	105	17.28	35.84	478	1650	708	0	59.9
Crystal 467RR	129	308.6	101	12418	113	1.60	45.04	102	1818	115	16.99	40.06	533	1677	607	0	68.9
Crystal 572RR	126	339.4	111	12424	114	1.39	54.37	123	1994	126	18.39	36.46	266	1439	595	0	72.8
Crystal 574RR	110	325.0	106	12873	118	1.53	50.00	113	1982	125	17.78	39.86	331	1612	626	0	68.6
Crystal 966RR	128	307.4	101	11073	101	1.54	44.67	101	1617	102	16.92	35.65	455	1446	644	0	61.8
Hilleshq 4302RR	105	308.5	101	9842	90	1.57	45.00	102	1442	91	16.99	31.73	492	1696	589	0	45.2
Hilleshq 4448RR	112	312.5	102	12012	110	1.38	46.22	105	1779	112	17.03	38.28	387	1531	530	0	50.3
Hilleshq 9528RR	123	314.9	103	12138	111	1.48	46.94	106	1813	114	17.20	38.59	392	1489	608	0	63.5
Hilleshq HIL9707	114	300.7	98	11587	106	1.65	42.66	97	1655	104	16.65	38.37	437	1650	687	0	58.9
Maribo 109	122	328.3	107	10166	93	1.50	51.01	116	1574	99	17.93	31.25	392	1455	627	0	49.6
Maribo 305	106	301.5	99	11335	104	1.45	42.89	97	1617	102	16.51	37.68	424	1449	581	0	51.7
Maribo MA502	121	303.5	99	11253	103	1.68	43.48	99	1617	102	16.81	37.07	552	1707	644	0	64.1
SX Avalanche RR(858)	127	324.2	106	11058	101	1.50	49.76	113	1699	107	17.71	34.03	410	1578	587	0	58.1
SX Canyon RR	118	311.3	102	11569	106	1.62	45.86	104	1707	108	17.24	37.06	373	1565	703	0	53.4
SX Cruze RR	117	293.3	96	11905	109	1.78	40.41	92	1651	104	16.43	40.28	444	1647	773	0	60.2
SX Marathon RR(856)	102	325.7	107	11876	109	1.39	50.22	114	1833	116	17.66	36.30	288	1610	551	0	59.4
SX Winchester RR	125	307.6	101	10074	92	1.59	44.73	101	1452	92	17.00	33.03	427	1656	624	0	51.1
SV RR244TT	120	319.4	105	11780	108	1.46	48.30	110	1776	112	17.44	37.01	336	1631	567	0	60.3
SV RR333	124	319.4	105	11868	108	1.53	48.32	110	1795	113	17.49	37.07	356	1633	619	0	58.2
SV RR351	113	319.0	104	11716	107	1.53	48.19	109	1778	112	17.44	36.61	350	1618	623	0	59.2
RR Filler #01s	132	307.3	101	12007	110	1.64	44.64	101	1749	110	17.01	38.95	441	1747	636	0	66.2
RR Filler #01v	133	302.8	99	11523	105	1.73	43.28	98	1653	104	16.88	37.92	494	1724	706	0	59.5
Experimental Trial (Comm status)																	
BTS 8606	242	308.6	101	11660	107	1.83	44.94	102	1701	107	17.20	37.91	562	1684	780	0	70.4
BTS 8629	224	306.8	100	12344	113	1.83	44.46	101	1799	113	17.12	40.15	575	1513	849	0	72.1
BTS 8735	234	311.4	102	11728	107	1.72	45.70	104	1729	109	17.28	37.42	499	1555	742	0	68.9
BTS 8742	207	303.7	99	11411	104	1.78	43.60	99	1641	103	16.93	37.63	529	1546	803	0	70.6
BTS 8749	202	307.7	101	11004	101	1.85	44.68	101	1609	101	17.20	35.36	536	1745	778	0	73.4
BTS 8756	241	302.8	99	10842	99	2.12	43.36	98	1549	98	17.11	35.90	523	1775	1013	0	72.5
BTS 8767	235	310.7	102	11603	106	1.78	45.50	103	1697	107	17.29	37.41	505	1711	753	0	68.6
BTS 8770	247	309.3	101	11770	108	1.64	45.12	102	1722	109	17.12	37.95	506	1642	641	0	69.8
BTS 8784	236	320.6	105	11359	104	1.54	48.22	109	1708	108	17.64	35.36	383	1537	639	0	66.8
BTS 8787	219	311.2	102	11853	108	1.54	45.66	104	1747	110	17.16	37.90	414	1640	587	0	67.3
BTS 8798	223	316.7	104	11294	103	1.62	47.15	107	1671	105	17.49	36.04	361	1565	699	0	75.0
Crystal 573RR	225	319.2	104	11375	104	1.53	47.85	108	1711	108	17.56	35.47	362	1512	642	0	68.5
Crystal 578RR	220	312.1	102	11446	105	1.64	45.90	104	1689	107	17.26	36.43	439	1665	652	0	69.1
Crystal 684RR	239	304.2	100	12620	115	1.57	43.74	99	1827	115	16.82	41.08	477	1663	583	0	80.8
Crystal 792RR	218	321.8	105	11337	104	1.43	48.56	110	1704	107	17.61	35.76	294	1472	606	0	72.1
Crystal 793RR	246	324.6	106	12103	111	1.42	49.31	112	1842	116	17.75	37.29	364	1374	603	0	72.3
Crystal 794RR	208	311.9	102	11980	109	1.58	45.86	104	1757	111	17.22	38.58	485	1614	604	0	54.5
Crystal 795RR	215	306.2	100	10998	100	1.83	44.28	100	1598	101	17.09	35.78	489	1594	846	0	66.5
Crystal 796RR	238	312.9	102	12477	114	1.64	46.11	105	1846	116	17.30	39.79	442	1602	678	0	67.8
Crystal 797RR	201	294.4	96	11842	108	1.87	41.06	93	1669	105	16.51	39.58	611	1761	761	0	57.1
Hilleshq HIL9708	222	307.1	101	10134	93	1.69	44.51	101	1468	93	17.06	32.92	490	1560	739	0	67.5
Hilleshq HIL9895	209	291.7	95	10106	92	1.87	40.35	91	1405	89	16.37	34.72	574	1650	831	95	64.4
Hilleshq HIL9920	203	319.0	104	11027	101	1.58	47.80	108	1648	104	17.58	34.95	449	1666	609	0	69.7
Hilleshq HIL9921	204	317.0	104	10546	96	1.56	47.23	107	1567	99	17.43	33.76	508	1516	618	0	73.6
Hilleshq HIL9922	231	290.4	95	10514	96	1.85	39.97	91	1449	91	16.29	36.12	471	1650	841	0	69.3
Hilleshq HIL9923	243	307.8	101	9722	89	2.00	44.72	101	1405	89	17.28	31.98	616	1766	894	0	41.4
Hilleshq HIL9924	237	306.6	100	10123	93	2.04	44.41	101	1471	93	17.25	32.84	543	1713	970	0	36.4
Maribo MA504	229	301.4	99	11828	108	1.66	42.98	97	1675	106	16.71	39.80	505	1535	715	0	63.0
Maribo MA611	245	291.3	95	9843	90	1.85	40.21	91	1362	86	16.34	33.64	613	1745	765	0	64.5
Maribo MA717	232	310.5	102	10832	99	1.76	45.46	103	1589	100	17.26	34.58	453	1562	798	0	67.8
Maribo MA718	221	292.2	96	9572	87	2.03	40.46	92	1328	84	16.49	32.76	796	1916	793	0	52.2
Maribo MA719	213	307.0	100	10574	97	1.96	44.51	101	1524	96	17.21	34.89	507	1804	876	0	43.3
SX RR1861	233	304.3	100	11087	101	1.62	43.76	99	1601	101	1						

Table 10. 2017 Performance of All RR Varieties - ACSC Official Trial

Georgetown MN

Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$ ++	Rev/T %Bnch	Rev/A \$ ++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg. %
103	349.4	100	11554	102	1.07	57.40	101	1903	103	18.55	33.00	121	1682	341	0	89.7
116	356.5	102	11522	102	1.03	59.55	104	1916	104	18.86	32.45	128	1666	312	0	90.8
130	343.2	99	12044	107	0.94	55.53	97	1941	105	18.10	35.23	130	1555	266	0	92.4
119	347.5	100	12424	110	1.02	56.83	100	2038	110	18.40	35.86	132	1669	301	0	94.4
109	352.6	101	11322	100	0.97	58.37	102	1869	101	18.59	32.14	116	1607	283	63	89.9
101	344.8	99	12365	110	1.03	56.01	98	2013	109	18.28	35.92	156	1718	285	0	92.9
111	354.7	102	11421	101	0.99	59.00	104	1893	102	18.71	32.20	122	1502	323	0	88.4
115	362.2	104	11434	101	1.06	61.27	107	1943	105	19.18	31.51	122	1664	333	0	89.3
107	347.9	100	11842	105	1.11	56.93	100	1941	105	18.50	34.07	168	1845	307	0	91.4
108	344.5	99	12215	108	1.02	55.91	98	1987	108	18.25	35.22	162	1582	312	0	89.5
131	347.7	100	12745	113	0.94	56.88	100	2083	113	18.33	36.61	120	1600	264	0	88.8
104	347.9	100	11035	98	1.09	56.95	100	1800	97	18.48	31.85	128	1675	352	0	92.3
129	346.3	99	12099	107	1.02	56.44	99	1971	107	18.32	34.94	140	1698	286	0	91.6
126	364.3	105	11309	100	1.01	61.91	109	1918	104	19.24	31.23	128	1620	308	0	92.5
110	345.9	99	12657	112	1.01	56.34	99	2063	112	18.30	36.55	138	1685	282	0	89.5
128	353.9	102	11582	103	0.97	58.75	103	1924	104	18.66	32.82	162	1509	290	0	92.9
105	347.2	100	10693	95	0.94	56.71	99	1746	95	18.29	30.86	149	1643	242	0	83.4
112	356.2	102	12467	111	0.95	59.44	104	2085	113	18.76	34.89	132	1536	281	32	89.3
123	353.2	101	11768	104	0.94	58.55	103	1958	106	18.60	33.18	131	1541	270	0	84.2
114	340.6	98	12329	109	1.05	54.72	96	1986	107	18.08	36.11	165	1696	307	0	91.3
122	360.8	104	10953	97	1.01	60.85	107	1850	100	19.06	30.14	128	1541	328	0	87.8
106	347.2	100	12014	106	1.02	56.71	99	1953	106	18.38	34.77	168	1474	337	0	86.1
121	348.3	100	10953	97	1.10	57.05	100	1797	97	18.52	31.51	164	1732	333	0	87.4
127	357.1	103	11035	98	0.98	59.72	105	1839	100	18.82	31.08	143	1621	275	63	86.6
118	359.7	103	11895	105	0.98	60.51	106	1992	108	18.95	33.11	117	1649	278	0	87.8
117	333.2	96	11542	102	1.05	52.48	92	1815	98	17.71	34.70	126	1620	338	0	90.8
102	361.1	104	11765	104	0.98	60.93	107	1983	107	19.03	32.57	126	1610	282	0	91.8
125	347.2	100	11202	99	0.94	56.73	100	1823	99	18.30	32.37	121	1621	257	32	85.3
120	350.5	101	11955	106	1.04	57.73	101	1968	107	18.56	34.10	129	1649	321	0	88.4
124	355.9	102	11978	106	0.92	59.37	104	2001	108	18.72	33.51	111	1603	249	0	91.6
113	351.2	101	11472	102	1.03	57.95	102	1902	103	18.60	32.72	143	1603	318	0	89.2
132	343.2	99	11211	99	1.14	55.53	97	1803	98	18.30	32.77	182	1810	338	0	93.8
133	341.3	98	12048	107	1.13	54.94	96	1927	104	18.19	35.37	185	1801	330	0	92.2
(status)																
242	355.8	102	11822	105	0.96	59.31	104	1970	107	18.76	33.27	131	1601	279	0	89.2
224	343.8	99	12811	114	0.99	55.68	98	2075	112	18.18	37.34	143	1441	337	0	96.6
234	347.5	100	13070	116	0.92	56.81	100	2139	116	18.30	37.64	117	1453	294	0	95.6
207	347.7	100	10853	96	1.10	56.84	100	1770	96	18.48	31.35	125	1676	360	0	91.8
202	344.5	99	11186	99	1.03	55.91	98	1814	98	18.26	32.56	155	1635	315	0	88.5
241	350.9	101	11264	100	1.14	57.83	101	1858	101	18.69	32.12	144	1812	358	0	95.5
235	363.5	104	12271	109	0.91	61.64	108	2082	113	19.10	33.79	139	1526	254	0	88.0
247	348.8	100	12565	111	0.95	57.22	100	2060	112	18.39	36.09	136	1567	274	0	88.1
236	359.1	103	11309	100	0.99	60.30	106	1901	103	18.96	31.54	130	1599	312	0	94.1
219	327.1	94	11591	103	0.91	50.68	89	1794	97	17.26	35.53	135	1551	247	0	87.7
223	350.3	101	11050	98	0.96	57.66	101	1819	98	18.48	31.58	137	1562	283	0	95.3
225	359.8	103	11935	106	1.00	60.51	106	2008	109	18.98	33.22	134	1575	304	0	93.9
220	357.2	103	12635	112	0.90	59.72	105	2113	114	18.76	35.43	158	1527	239	0	92.7
239	354.5	102	13245	117	0.99	58.91	103	2201	119	18.73	37.47	151	1737	270	0	93.6
218	354.3	102	11743	104	1.00	58.87	103	1950	106	18.72	33.23	126	1505	331	0	89.6
246	358.2	103	12495	111	0.97	60.03	105	2094	113	18.88	34.94	135	1444	320	0	91.7
208	342.2	98	11941	106	1.02	55.22	97	1927	104	18.12	34.96	174	1493	333	0	93.4
215	347.6	100	11185	99	1.05	56.82	100	1828	99	18.42	32.27	126	1592	347	0	93.7
238	352.7	101	12775	113	0.94	58.38	102	2117	115	18.58	36.22	144	1563	265	0	95.0
201	336.4	97	12051	107	1.01	53.48	94	1913	104	17.84	35.95	156	1736	276	0	81.1
222	352.0	101	11740	104	0.94	58.15	102	1940	105	18.55	33.40	160	1441	294	0	92.8
209	354.3	102	10964	97	1.07	58.85	103	1821	99	18.79	31.02	143	1560	367	0	89.3
203	360.5	104	11557	102	0.98	60.70	106	1945	105	19.01	32.16	142	1709	258	0	92.8
204	364.0	105	10942	97	0.96	61.78	108	1855	100	19.16	30.16	147	1560	283	0	91.3
231	341.0	98	10710	95	1.27	54.85	96	1724	93	18.32	31.45	188	1711	471	0	95.8
243	349.6	100	10935	97	1.16	57.44	101	1800	97	18.64	31.28	199	1702	379	0	85.4
237	348.4	100	9906	88	1.19	57.09	100	1623	88	18.62	28.48	161	1710	416	0	74.4
229	349.2	100	12500	111	1.02	57.35	101	2049	111	18.49	35.91	155	1545	333	0	96.0
245	347.4	100	10398	92	1.00	56.76	100	1701	92	18.37	29.96	123	1588	311	0	93.9
232	352.5	101	11890	105	1.08	58.31	102	1969	107	18.72	33.75	187	1554	369	0	84.6
221	351.3	101	10802	96	1.04	57.97	102	1781	96	18.61	30.81	162	1738	292	0	81.7
213	354.8	102	11128	99	1.14	59.01	104	1849	100	18.89	31.42	176	1756	357	0	87.8
233	356.1	102	11972	106	0.91	59.40	104	1997	108	18.72	33.70	123	1575	246	0	95.7
244	359.2	103	11810	105	0.88	60.31	106	1982	107	18.85	32.95	136	1575	221	0	85.9
210	349.1	100	11044	98	0.93	57.32	101	1815	98	18.40	31.63	154	1603	247	0	93.6
228	347.1	100	11659	103	0.94	56.67	99	1904	103	18.30	33.67	140	1573	270	0	89.8
227	345.1	99	11501	102	1.12	56.06	98	1869	101	18.38	33.39	142	1786	350	0	90.2
206	348.0	100	12511	111	0.96	56.94	100	2049	111	18.37	35.98	129	1572	291	0	97.0
205	354.2	102	12700	113	0.95	58.83	103	2109	114	18.66	35.93	133	1590	266	0	90.6
240	357.6	103	11828	105	0.95	59.85	105	1981	107	18.83	33.12	111	1605	269	0	93.3
216	355.0	102	12197	108	1.04	59.06	104	2029	110	18.79	34.42	148	1708	301	0	92.3
226	357.5	103	11851	105	0.97	59.81	105	1983	107	18.86	33.19	143	1584	284	0	96.8
212	361.2	104	12088	107	0.97	60.93	107	2039	110	19.03	33.51	126	1599	277	0	94.6
217	347.5	100	11772	104	0.94	56.80	100	1927	104	18.32	33.89	140	1550	274	0	88.6

Table 11. 2017 Performance of All RR Varieties - ACSC Official Trial

Hendrum MN																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	329.0	100	11362	102	1.24	51.23	100	1762	102	17.69	34.63	168	1540	487	0	72.9
BTS 8337	116	342.8	104	11353	102	1.02	55.39	108	1836	107	18.16	33.21	138	1422	363	32	67.7
BTS 8363	130	323.7	98	11969	108	1.04	49.62	97	1832	106	17.23	36.97	168	1403	374	0	68.5
BTS 8500	119	334.0	102	12307	111	1.01	52.72	103	1939	113	17.70	36.79	140	1434	352	0	62.6
BTS 8512	109	337.6	103	11595	105	1.03	53.82	105	1846	107	17.90	34.39	144	1380	379	0	75.8
BTS 8524	101	322.3	98	12590	114	1.16	49.20	96	1919	111	17.26	39.11	153	1617	411	0	71.5
BTS 8572	111	339.5	103	11642	105	1.10	54.40	106	1864	108	18.08	34.29	144	1424	423	0	71.4
Crystal 093RR	115	343.2	104	11318	102	1.11	55.52	108	1833	106	18.28	33.00	123	1463	428	0	72.2
Crystal 101RR	107	325.1	99	11347	102	1.21	50.02	98	1747	101	17.47	34.94	201	1608	436	0	70.3
Crystal 246RR	108	327.6	100	12345	111	1.00	50.79	99	1913	111	17.38	37.71	171	1385	346	0	68.2
Crystal 247RR	131	326.4	99	12461	112	1.02	50.44	99	1923	112	17.33	38.18	166	1482	333	0	68.7
Crystal 355RR	104	330.9	101	11048	100	1.14	51.79	101	1725	100	17.68	33.42	145	1499	431	0	61.0
Crystal 467RR	129	321.6	98	11800	106	1.08	48.97	96	1794	104	17.16	36.74	198	1540	354	0	75.5
Crystal 572RR	126	343.4	104	11653	105	0.98	55.56	109	1885	109	18.14	33.94	129	1346	356	0	78.6
Crystal 574RR	110	326.7	99	12505	113	1.03	50.52	99	1930	112	17.36	38.31	135	1432	365	0	70.5
Crystal 966RR	128	344.5	105	11434	103	0.98	55.92	109	1854	108	18.22	33.18	155	1263	369	0	74.9
Hilleshoq 4302RR	105	330.6	101	10607	96	0.96	51.69	101	1659	96	17.49	32.03	169	1421	309	0	59.2
Hilleshoq 4448RR	112	341.2	104	12251	110	1.05	54.91	107	1973	114	18.11	36.00	152	1321	409	32	67.1
Hilleshoq 9528RR	123	340.2	103	11559	104	1.01	54.59	107	1857	108	18.02	34.03	156	1332	368	0	65.6
Hilleshoq HIL9707	114	326.0	99	11997	108	1.08	50.30	98	1846	107	17.38	36.87	159	1453	390	0	69.5
Maribo 109	122	348.1	106	10104	91	0.95	57.01	111	1658	96	18.36	29.06	134	1322	340	0	55.9
Maribo 305	106	329.9	100	11944	108	0.99	51.50	101	1866	108	17.47	36.20	144	1301	366	0	54.3
Maribo MA502	121	336.7	102	11257	101	1.09	53.55	105	1792	104	17.92	33.99	172	1510	383	32	62.3
SX Avalanche RR(858)	127	335.0	102	10300	93	0.96	53.03	104	1632	95	17.71	30.74	183	1432	300	0	56.9
SX Canyon RR	118	339.5	103	11764	106	0.98	54.41	106	1888	110	17.95	34.60	131	1413	337	0	61.0
SX Cruze RR	117	317.2	96	11559	104	1.05	47.64	93	1735	101	16.91	36.42	131	1388	396	0	64.8
SX Marathon RR(856)	102	336.4	102	11556	104	0.96	53.45	104	1837	107	17.79	34.39	143	1393	324	0	67.1
SX Winchester RR	125	322.4	98	10091	91	0.94	49.22	96	1541	89	17.07	31.30	154	1363	315	0	51.8
SV RR244TT	120	331.7	101	11906	107	0.94	52.05	102	1870	109	17.53	35.82	115	1405	316	32	68.8
SV RR333	124	336.9	102	11520	104	1.05	53.60	105	1829	106	17.90	34.26	160	1430	372	0	63.7
SV RR351	113	340.7	104	11589	104	0.93	54.76	107	1866	108	17.97	33.93	119	1400	310	0	63.6
RR Filler #01s	132	319.8	97	11455	103	1.23	48.44	95	1734	101	17.22	35.83	202	1637	444	0	71.0
RR Filler #01v	133	325.6	99	11333	102	1.14	50.20	98	1746	101	17.43	34.86	173	1604	392	0	71.0
Experimental Trial (Comm status)																	
BTS 8606	242	336.0	102	12185	110	1.06	53.26	104	1927	112	17.86	36.33	185	1401	369	0	70.4
BTS 8629	224	327.5	100	12596	114	1.07	50.79	99	1947	113	17.46	38.71	184	1310	411	0	80.6
BTS 8735	234	331.0	101	13054	118	1.01	51.82	101	2038	118	17.57	39.62	164	1258	383	0	71.7
BTS 8742	207	325.0	99	10608	96	1.24	50.06	98	1626	94	17.49	32.67	179	1565	477	0	62.9
BTS 8749	202	326.2	99	10949	99	1.15	50.37	98	1697	98	17.49	33.51	163	1506	431	0	71.2
BTS 8756	241	331.7	101	10836	98	1.22	51.99	102	1692	98	17.80	32.75	147	1543	473	0	75.1
BTS 8752	235	335.2	102	12746	115	0.98	53.02	104	2013	117	17.77	38.14	158	1378	340	0	80.5
BTS 8770	247	328.4	100	12133	109	1.03	51.03	100	1886	109	17.48	36.97	183	1428	348	95	63.5
BTS 8784	236	342.1	104	11235	101	1.07	55.04	108	1806	105	18.19	32.83	140	1289	442	0	75.0
BTS 8787	219	331.3	101	11837	107	1.03	51.91	101	1845	107	17.60	35.97	158	1420	359	0	54.9
BTS 8798	223	327.7	100	11059	100	1.08	50.85	99	1708	99	17.46	33.71	180	1385	397	0	66.9
Crystal 573RR	225	335.0	102	11119	100	1.14	52.99	104	1759	102	17.90	33.21	179	1401	443	0	71.5
Crystal 578RR	220	329.8	100	13060	118	1.05	51.44	100	2035	118	17.55	39.59	161	1466	358	0	72.2
Crystal 684RR	239	330.0	100	12435	112	1.13	51.49	101	1938	112	17.63	37.88	179	1514	406	0	79.1
Crystal 792RR	218	336.8	102	11717	106	1.05	53.52	105	1863	108	17.91	34.85	136	1413	390	0	68.4
Crystal 793RR	246	342.5	104	12135	109	0.89	55.17	108	1944	113	18.02	35.52	137	1272	306	0	76.8
Crystal 794RR	208	328.2	100	12186	110	1.10	50.98	100	1884	109	17.50	37.33	207	1430	401	0	57.8
Crystal 795RR	215	328.3	100	10968	99	1.22	51.02	100	1702	99	17.64	33.53	179	1469	484	0	68.1
Crystal 796RR	238	325.8	99	12767	115	1.07	50.26	98	1980	115	17.40	39.15	166	1421	394	0	65.7
Crystal 797RR	201	325.4	99	11968	108	1.02	50.16	98	1837	107	17.30	36.98	197	1428	328	95	58.7
Hilleshoq HIL9708	222	331.4	101	10529	95	1.04	51.93	101	1641	95	17.60	31.83	182	1369	369	0	68.4
Hilleshoq HIL9895	209	321.3	98	10165	92	1.10	48.94	96	1547	90	17.16	31.73	155	1407	415	0	69.2
Hilleshoq HIL9920	203	340.7	104	10765	97	1.04	54.65	107	1725	100	18.09	31.74	158	1455	351	0	64.8
Hilleshoq HIL9921	204	339.3	103	10113	91	1.10	54.23	106	1615	94	18.09	29.99	178	1316	438	0	75.2
Hilleshoq HIL9922	231	323.2	98	10516	95	1.12	49.49	97	1604	93	17.28	32.61	150	1478	424	0	64.1
Hilleshoq HIL9923	243	334.3	102	10349	93	1.20	52.77	103	1633	95	17.91	30.88	183	1537	451	0	55.8
Hilleshoq HIL9924	237	336.4	102	9299	84	1.27	53.38	104	1473	85	18.08	27.51	173	1516	516	0	44.1
Maribo MA504	229	329.3	100	12433	112	1.05	51.30	100	1924	112	17.51	37.99	178	1334	385	0	70.6
Maribo MA611	245	324.4	99	10535	95	1.16	49.86	97	1611	93	17.38	32.68	186	1494	436	95	61.3
Maribo MA717	232	333.4	101	11605	105	1.09	52.48	103	1823	106	17.77	34.96	179	1370	409	0	70.5
Maribo MA718	221	323.6	98	10404	94	1.11	49.62	97	1602	93	17.33	32.15	230	1602	353	0	56.2
Maribo MA719	213	338.5	103	10750	97	1.17	54.01	106	1715	99	18.11	31.62	178	1545	440	0	56.9
SX RR1861	233	333.4	101	11042	100	1.09	52.49	103	1735	101							

Table 12. 2017 Performance of All RR Varieties - ACSC Official Trial

Hillsboro ND																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	356.4	99	13386	101	1.26	59.53	99	2234	101	19.09	37.68	131	1790	454	32	77.6
BTS 8337	116	368.6	103	13793	105	1.05	63.22	105	2371	107	19.51	37.45	114	1662	335	0	79.5
BTS 8363	130	359.5	100	14098	107	1.01	60.46	100	2352	106	18.98	39.64	108	1596	324	0	71.4
BTS 8500	119	355.0	99	14164	107	1.14	59.08	98	2361	107	18.88	39.74	140	1681	385	0	73.7
BTS 8512	109	355.7	99	12773	97	1.07	59.30	99	2136	96	18.85	35.66	103	1614	363	0	76.5
BTS 8524	101	349.0	97	13794	105	1.17	57.28	95	2260	102	18.62	39.65	129	1806	382	0	75.2
BTS 8572	111	367.9	103	13654	104	1.07	62.99	105	2334	105	19.47	37.25	108	1638	360	0	75.2
Crystal 093RR	115	372.7	104	13505	102	1.16	64.45	107	2337	106	19.80	36.14	128	1707	400	0	80.8
Crystal 101RR	107	349.0	97	13275	101	1.26	57.29	95	2172	98	18.71	38.12	138	1986	398	0	78.2
Crystal 246RR	108	354.9	99	13525	103	1.02	59.07	98	2252	102	18.77	38.15	135	1626	310	0	74.8
Crystal 247RR	131	369.0	103	13898	105	1.02	63.32	105	2385	108	19.47	37.70	144	1674	291	126	74.5
Crystal 355RR	104	367.9	103	13561	103	1.16	63.01	105	2330	105	19.57	36.67	114	1741	400	0	77.5
Crystal 467RR	129	353.6	99	14147	107	1.12	58.66	97	2339	106	18.80	40.16	141	1815	334	0	78.1
Crystal 572RR	126	375.6	105	13453	102	1.06	65.32	109	2338	106	19.84	35.94	95	1625	356	0	79.2
Crystal 574RR	110	357.7	100	14353	109	1.08	59.90	100	2403	109	18.96	40.17	114	1672	350	0	75.8
Crystal 986RR	128	379.2	106	13843	105	1.02	66.43	110	2416	109	19.97	36.64	118	1540	340	0	75.3
Hillesq 4302RR	105	361.1	101	12542	95	1.08	60.93	101	2122	96	19.14	34.57	143	1786	312	0	53.2
Hillesq 4448RR	112	360.0	100	13334	101	1.17	60.61	101	2252	102	19.17	36.81	120	1635	429	0	64.2
Hillesq 9528RR	123	362.7	101	13311	101	1.10	61.42	102	2262	102	19.22	36.39	126	1710	351	0	69.1
Hillesq HIL9707	114	343.1	96	11863	90	1.26	55.48	92	1913	86	18.42	34.95	185	1795	432	0	52.1
Maribo 109	122	365.1	102	11059	84	1.19	62.13	103	1889	85	19.44	30.17	129	1659	435	0	56.4
Maribo 305	106	353.9	99	12738	97	1.05	58.75	98	2116	96	18.73	35.91	124	1559	356	0	59.8
Maribo MA502	121	348.4	97	12558	95	1.29	57.08	95	2061	93	18.72	36.02	156	1914	437	639	64.6
SX Avalanche RR(858)	127	366.4	102	12841	97	0.96	62.53	104	2200	99	19.29	34.90	112	1597	282	0	66.5
SX Canyon RR	118	362.6	101	13325	101	1.06	61.38	102	2254	102	19.19	36.79	118	1684	331	0	68.6
SX Cruze RR	117	333.2	93	13302	101	1.15	52.48	87	2094	95	17.81	40.00	130	1693	392	0	71.2
SX Marathon RR(856)	102	364.8	102	13317	101	1.03	62.05	103	2269	102	19.28	36.51	110	1633	325	0	65.8
SX Winchester RR	125	356.4	99	12170	92	1.05	59.52	99	2027	92	18.86	34.22	128	1686	316	0	59.9
SV RR244TT	120	361.0	101	13564	103	1.07	60.89	101	2292	103	19.13	37.32	97	1719	341	0	67.2
SV RR333	124	361.3	101	13712	104	1.05	61.00	101	2306	104	19.12	38.06	109	1650	339	0	66.7
SV RR351	113	358.3	100	13727	104	1.13	60.09	100	2301	104	19.03	38.28	133	1735	364	0	66.5
RR Filler #01s	132	359.8	100	13954	106	1.21	60.54	101	2344	106	19.19	38.93	132	1892	385	0	77.5
RR Filler #01v	133	346.9	97	13284	101	1.26	56.64	94	2166	98	18.61	38.35	179	1957	394	0	79.3
Experimental Trial (Comm status)																	
BTS 8606	242	370.7	103	15760	119	1.04	63.81	106	2710	122	19.58	42.49	101	1663	323	0	75.3
BTS 8629	224	362.0	101	15145	115	1.06	61.20	102	2559	116	19.16	41.84	131	1522	364	0	74.2
BTS 8735	234	368.0	103	15603	118	1.01	62.97	105	2666	120	19.42	42.49	107	1495	341	0	79.1
BTS 8742	207	351.9	98	13067	99	1.37	58.19	97	2148	97	18.98	37.35	116	1873	524	0	74.7
BTS 8749	202	363.4	101	13685	104	1.14	61.60	102	2306	104	19.33	37.91	103	1761	383	0	70.3
BTS 8756	241	362.8	101	13509	102	1.22	61.45	102	2275	103	19.38	37.53	100	1838	426	0	76.2
BTS 8775	235	373.5	104	15618	118	1.05	64.64	107	2699	122	19.74	41.76	102	1709	324	0	80.5
BTS 8770	247	367.5	102	15537	118	1.01	62.84	104	2650	120	19.41	42.38	104	1719	291	0	69.7
BTS 8784	236	373.8	104	14296	108	1.04	64.72	108	2470	112	19.74	38.39	89	1529	367	0	73.8
BTS 8787	219	361.0	101	14727	112	1.13	60.92	101	2478	112	19.20	40.87	112	1784	365	0	71.7
BTS 8798	223	366.2	102	14194	108	1.06	62.46	104	2411	109	19.39	38.92	81	1587	368	0	76.7
Crystal 573RR	225	368.3	103	14881	113	1.15	63.06	105	2542	115	19.57	40.48	102	1721	396	0	75.1
Crystal 578RR	220	368.4	103	15986	121	1.03	63.09	105	2728	123	19.48	43.52	114	1685	310	0	79.1
Crystal 684RR	239	359.1	100	15049	114	1.08	60.35	100	2532	114	19.04	41.92	115	1728	340	0	71.3
Crystal 792RR	218	368.0	103	14213	108	1.06	62.99	105	2426	110	19.47	38.84	88	1607	359	0	80.5
Crystal 793RR	246	365.5	102	15055	114	1.08	62.25	103	2555	115	19.38	41.31	120	1598	368	95	75.1
Crystal 794RR	208	355.4	99	14750	112	1.18	59.26	98	2457	111	18.97	41.50	130	1713	418	0	65.7
Crystal 795RR	215	362.5	101	14247	108	1.12	61.35	102	2408	109	19.24	39.40	102	1702	379	0	79.3
Crystal 796RR	238	367.6	103	15358	116	1.03	62.84	104	2618	118	19.43	41.91	98	1730	302	0	81.6
Crystal 797RR	201	360.0	100	15579	118	1.06	60.63	101	2612	118	19.09	43.40	108	1779	309	0	66.5
Hillesq HIL9708	222	362.6	101	13181	100	1.10	61.37	102	2220	100	19.24	36.65	126	1627	373	0	68.8
Hillesq HIL9895	209	349.2	97	12490	95	1.30	57.41	95	2052	93	18.75	35.83	132	1799	477	0	59.7
Hillesq HIL9920	203	377.6	105	14614	111	1.03	65.85	109	2539	115	19.94	38.90	102	1727	302	0	68.8
Hillesq HIL9921	204	370.0	103	12201	92	1.14	63.58	106	2090	94	19.66	33.15	112	1628	408	0	71.4
Hillesq HIL9922	231	352.4	98	12529	95	1.18	58.36	97	2061	93	18.82	35.79	118	1765	403	675	67.9
Hillesq HIL9923	243	365.8	102	12255	93	1.30	62.34	104	2081	94	19.59	33.71	137	1731	491	0	62.7
Hillesq HIL9924	237	367.2	102	12317	93	1.23	62.74	104	2100	95	19.60	33.64	111	1745	450	0	60.2
Maribo MA504	229	363.2	101	15013	114	1.10	61.57	102	2534	114	19.27	41.66	117	1643	373	0	72.3
Maribo MA611	245	348.2	97	12546	95	1.24	57.11	95	2063	93	18.65	35.85	140	1829	426	0	68.3
Maribo MA717	232	363.4	101	13081	99	1.16	61.60	102	2211	100	19.33	36.25	112	1643	424	0	65.4
Maribo MA718	221	358.7	100	11964	91	1.14	60.22	100	2007	91	19.08	33.43	146	1812	346	0	54.5
Maribo MA719	213	363.5	101	12494	95	1.23	61.63	102	2116	96	19.42	34.44	104	1790	446	576	67.5
SX RR1861	233	355.1	99	14491	110	1.05	59.14	98	2409	109	18.81	4					

Table 13. 2017 Performance of All RR Varieties - ACSC Official Trial

Climax MN																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	335.5	102	9304	100	1.01	53.17	104	1473	102	17.78	27.80	109	1437	363	0	90.6
BTS 8337	116	350.5	107	10245	110	1.04	57.73	113	1688	117	18.56	29.19	114	1407	389	0	85.7
BTS 8363	130	330.7	101	9615	103	0.95	51.73	101	1507	105	17.49	29.05	118	1351	334	0	84.9
BTS 8500	119	330.9	101	10891	117	1.02	51.79	102	1705	118	17.56	32.90	118	1484	349	0	89.4
BTS 8512	109	336.8	103	9910	107	0.97	53.57	105	1578	109	17.82	29.42	122	1362	348	32	88.3
BTS 8524	101	320.6	98	9790	105	1.12	48.68	95	1480	103	17.14	30.64	131	1552	407	32	87.9
BTS 8572	111	347.7	106	10141	109	0.96	56.87	112	1657	115	18.33	29.22	104	1364	341	0	86.9
Crystal 093RR	115	337.8	103	10270	111	0.96	53.87	106	1637	114	17.85	30.47	110	1404	331	0	83.2
Crystal 101RR	107	322.8	98	9965	107	1.08	49.34	97	1524	106	17.23	30.87	143	1573	367	0	86.3
Crystal 246RR	108	332.8	101	8974	97	0.98	52.35	103	1413	98	17.62	26.96	135	1387	341	0	83.0
Crystal 247RR	131	329.8	100	10090	109	0.94	51.45	101	1573	109	17.43	30.60	131	1368	318	0	85.2
Crystal 355RR	104	337.5	103	9765	105	1.07	53.79	106	1550	108	17.94	29.10	118	1479	393	0	88.4
Crystal 467RR	129	326.7	100	9973	107	1.01	50.52	99	1540	107	17.33	30.57	147	1457	342	0	92.5
Crystal 572RR	126	348.4	106	10547	114	0.92	57.09	112	1726	120	18.35	30.35	98	1280	340	0	90.2
Crystal 574RR	110	325.1	99	10850	117	1.00	50.04	98	1665	116	17.24	33.42	111	1436	352	0	89.3
Crystal 966RR	128	307.1	94	7275	78	0.90	44.60	87	1060	74	16.26	23.61	172	1324	283	0	86.8
Hilleshög 4302RR	105	317.2	97	8134	88	0.94	47.64	93	1218	85	16.79	25.71	166	1396	297	0	77.2
Hilleshög 4448RR	112	320.6	98	8699	94	0.90	48.69	95	1324	92	16.94	27.08	123	1356	297	0	77.2
Hilleshög 9528RR	123	326.2	99	9517	102	0.87	50.36	99	1465	102	17.17	29.23	120	1320	279	0	81.6
Hilleshög HIL9707	114	310.6	95	8371	90	0.94	45.65	90	1228	85	16.47	27.01	161	1378	303	0	86.4
Maribo 109	122	335.9	102	7871	85	0.87	53.32	105	1249	87	17.67	23.43	119	1320	283	0	80.4
Maribo 305	106	312.7	95	8361	90	0.89	46.27	91	1237	86	16.52	26.69	127	1309	294	0	80.8
Maribo MA502	121	318.2	97	8196	88	0.95	47.96	94	1236	86	16.88	25.75	145	1471	295	0	82.4
SX Avalanche RR(858)	127	333.8	102	8715	94	0.94	52.67	103	1376	95	17.63	26.09	120	1393	316	32	83.7
SX Canyon RR	118	336.5	103	10135	109	0.91	53.48	105	1612	112	17.74	30.07	110	1357	305	0	84.0
SX Cruze RR	117	318.3	97	9981	107	1.03	47.97	94	1502	104	16.94	31.43	128	1432	373	0	86.0
SX Marathon RR(856)	102	337.3	103	10467	113	0.92	53.63	105	1662	115	17.76	31.12	114	1334	316	0	82.5
SX Winchester RR	125	317.1	97	8110	87	0.89	47.63	93	1218	85	16.75	25.55	118	1400	277	0	79.4
SV RR244TT	120	330.8	101	10167	109	0.97	51.75	102	1588	110	17.51	30.81	124	1389	337	0	84.3
SV RR333	124	341.5	104	10049	108	0.86	54.99	108	1619	112	17.94	29.39	99	1330	277	0	86.3
SV RR351	113	325.8	99	10007	108	0.89	50.24	99	1542	107	17.18	30.76	120	1316	296	0	86.3
RR Filler #01s	132	324.4	99	9896	107	1.10	49.84	98	1524	106	17.33	30.47	141	1551	385	0	89.7
RR Filler #01v	133	322.0	98	10126	109	1.10	49.09	96	1545	107	17.20	31.44	140	1570	382	0	89.5
Experimental Trial (Comm status)																	
BTS 8606	242	337.0	103	9393	101	0.90	53.62	105	1491	103	17.76	27.91	118	1348	299	0	87.6
BTS 8629	224	337.7	103	11158	120	0.94	53.84	106	1779	123	17.84	33.13	116	1306	346	0	93.0
BTS 8735	234	337.7	103	9268	100	0.92	53.83	106	1469	102	17.80	27.68	128	1285	326	0	91.6
BTS 8742	207	334.4	102	8983	97	1.02	52.82	104	1412	98	17.73	26.84	129	1447	359	0	90.5
BTS 8749	202	338.0	103	9798	105	1.04	53.91	106	1580	108	17.94	29.02	133	1426	378	0	86.4
BTS 8756	241	343.3	105	9656	104	1.01	55.49	109	1563	108	18.18	28.15	106	1515	347	0	91.5
BTS 8767	235	335.7	102	10123	109	0.89	53.20	104	1601	111	17.67	30.33	125	1376	286	0	86.9
BTS 8770	247	339.0	103	9744	105	0.91	54.21	106	1557	108	17.87	28.80	116	1401	295	0	88.5
BTS 8784	236	348.5	106	9535	103	0.95	57.06	112	1561	108	18.40	27.22	101	1353	345	0	87.9
BTS 8787	219	335.2	102	9528	103	0.99	53.07	104	1500	104	17.73	28.64	120	1450	333	0	83.5
BTS 8798	223	330.4	101	9249	100	1.06	51.64	101	1448	100	17.59	27.90	134	1406	403	0	92.1
Crystal 573RR	225	340.8	104	9962	107	0.92	54.73	107	1599	111	17.97	29.31	112	1344	318	0	84.9
Crystal 578RR	220	335.3	102	9576	103	0.99	53.11	104	1507	105	17.74	28.73	124	1403	346	0	86.3
Crystal 684RR	239	336.2	102	10959	118	1.00	53.35	105	1724	120	17.78	32.88	121	1443	339	0	87.4
Crystal 792RR	218	336.0	102	10093	109	1.05	53.29	105	1603	111	17.84	29.83	120	1475	377	0	87.9
Crystal 793RR	246	350.3	107	9961	107	0.87	57.57	113	1632	113	18.39	28.49	99	1304	292	0	84.4
Crystal 794RR	208	328.5	100	9815	106	1.05	51.05	100	1529	106	17.48	29.84	138	1474	376	0	86.0
Crystal 795RR	215	339.7	103	9458	102	0.99	54.41	107	1506	105	17.96	28.13	120	1399	350	0	88.2
Crystal 796RR	238	330.2	101	10691	115	0.94	51.59	101	1668	116	17.47	32.51	115	1344	335	0	88.8
Crystal 797RR	201	334.1	102	10330	111	1.03	52.74	103	1609	112	17.70	31.47	151	1462	350	0	84.2
Hilleshög HIL9708	222	327.6	100	7828	84	0.85	50.81	100	1213	84	17.24	24.01	124	1256	282	0	86.9
Hilleshög HIL9895	209	324.8	99	7886	85	0.91	49.95	98	1213	84	17.15	24.35	128	1450	276	0	83.7
Hilleshög HIL9920	203	337.8	103	8533	92	0.84	53.86	106	1357	94	17.73	25.20	126	1348	251	0	83.7
Hilleshög HIL9921	204	333.8	102	7833	84	0.93	52.64	103	1229	85	17.63	23.26	131	1350	325	0	75.5
Hilleshög HIL9922	231	320.4	98	7931	85	0.90	48.66	95	1206	84	16.93	24.75	115	1378	292	0	90.9
Hilleshög HIL9923	243	332.9	101	6900	74	1.06	52.40	103	1085	75	17.70	20.84	161	1531	358	0	76.0
Hilleshög HIL9924	237	319.4	97	7058	76	1.11	48.37	95	1063	74	17.07	22.25	146	1481	413	0	75.0
Maribo MA504	229	322.5	98	8073	87	0.88	49.27	97	1234	86	17.01	24.87	133	1344	279	0	87.8
Maribo MA611	245	311.8	95	7337	79	0.98	46.08	90	1081	75	16.56	23.65	139	1481	310	0	83.8
Maribo MA717	232	333.8	102	8168	88	0.88	52.64	103	1284	89	17.57	24.64	111	1354	278	0	84.7
Maribo MA718	221	322.6	98	7464	80	1.07	49.28	97	1128	78	17.17	23.30	185	1488	363	0	72.6
Maribo MA719	213	326.8	100	7028	76	1.08	50.56	99	1093	76	17.43	21.51	136	1593	370	0	82.0
SX RR1861	233	337.1	103	9841	106	0.89	53.65	105	1563	108	17.75	29.26	109	1297	311	0	94.0

Table 14. 2017 Performance of All RR Varieties - ACSC Official Trial

Grand Forks ND																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	352.4	99	11374	97	0.90	58.29	98	1882	97	18.52	32.31	129	1547	239	0	76.6
BTS 8337	116	357.2	100	11258	96	0.87	59.77	100	1881	96	18.73	31.54	130	1523	219	0	77.4
BTS 8363	130	348.0	98	12320	106	0.80	56.98	96	2017	103	18.20	35.39	122	1449	191	0	78.9
BTS 8500	119	351.0	98	12258	105	0.87	57.88	97	2020	104	18.41	34.87	128	1531	218	0	76.6
BTS 8512	109	359.7	101	11280	97	0.86	60.50	102	1900	97	18.85	31.31	122	1529	220	0	78.7
BTS 8524	101	352.2	99	12535	107	0.85	58.23	98	2073	106	18.46	35.62	139	1570	196	0	74.7
BTS 8572	111	363.7	102	11765	101	0.82	61.72	104	1996	102	19.01	32.44	117	1467	205	0	82.3
Crystal 093RR	115	373.2	105	11780	101	0.87	64.61	108	2039	105	19.53	31.56	110	1516	234	0	78.4
Crystal 101RR	107	359.5	101	12368	106	0.86	60.46	101	2079	107	18.84	34.46	139	1613	186	0	78.4
Crystal 246RR	108	355.0	100	12385	106	0.79	59.09	99	2060	106	18.54	34.96	137	1451	175	0	70.3
Crystal 247RR	131	357.4	100	12135	104	0.71	59.82	100	2031	104	18.58	34.04	115	1343	153	0	75.0
Crystal 355RR	104	360.8	101	11601	99	0.85	60.85	102	1955	100	18.90	32.19	121	1490	223	0	75.0
Crystal 467RR	129	346.5	97	12144	104	0.83	56.51	95	1986	102	18.16	35.06	139	1541	186	0	80.2
Crystal 572RR	126	370.3	104	12471	107	0.80	63.73	107	2147	110	19.31	33.57	114	1429	198	0	81.8
Crystal 574RR	110	351.3	98	12300	105	0.85	57.96	97	2029	104	18.42	35.04	125	1560	203	0	80.0
Crystal 966RR	128	355.5	100	12439	107	0.80	59.24	99	2074	106	18.58	34.94	139	1390	205	0	79.2
Hilleshög 4302RR	105	353.9	99	11353	97	0.82	58.77	99	1884	97	18.52	32.03	138	1462	198	0	69.0
Hilleshög 4448RR	112	361.3	101	12812	110	0.82	61.00	102	2157	111	18.89	35.56	118	1487	202	0	70.8
Hilleshög 9528RR	123	346.9	97	11762	101	0.84	56.65	95	1922	99	18.19	33.88	139	1491	207	0	78.4
Hilleshög HIL9707	114	341.6	96	12190	104	0.82	55.03	92	1956	100	17.89	35.79	138	1469	192	32	79.2
Maribo 109	122	371.8	104	10296	88	0.83	64.16	108	1770	91	19.41	27.82	129	1480	200	0	72.9
Maribo 305	106	352.8	99	12554	108	0.81	58.42	98	2074	106	18.44	35.65	128	1417	201	0	64.3
Maribo MA502	121	346.9	97	11785	101	0.91	56.64	95	1924	99	18.26	33.93	173	1608	218	0	77.3
SX Avalanche RR(858)	127	352.7	99	11753	101	0.84	58.40	98	1943	100	18.47	33.34	155	1407	222	0	72.4
SX Canyon RR	118	362.8	102	12704	109	0.75	61.45	103	2157	111	18.89	34.98	106	1396	170	0	73.2
SX Cruze RR	117	331.3	93	12154	104	0.86	51.91	87	1907	98	17.42	36.66	129	1550	207	0	77.9
SX Marathon RR(856)	102	349.7	98	12295	105	0.79	57.48	96	2021	104	18.28	35.14	113	1467	186	0	69.0
SX Winchester RR	125	350.2	98	11362	97	0.83	57.63	97	1868	96	18.34	32.50	127	1565	187	0	66.7
SV RR244TT	120	352.1	99	12859	110	0.78	58.21	98	2128	109	18.38	36.48	115	1453	176	0	70.8
SV RR333	124	354.5	99	12282	105	0.79	58.92	99	2042	105	18.51	34.64	122	1485	174	0	74.5
SV RR351	113	355.6	100	12283	105	0.80	59.28	99	2049	105	18.58	34.44	113	1488	183	0	74.5
RR Filler #01s	132	354.7	99	12122	104	0.85	59.00	99	2013	103	18.59	34.20	144	1616	179	0	77.1
RR Filler #01v	133	351.2	98	12082	103	0.87	57.93	97	1991	102	18.43	34.46	149	1610	191	0	77.9
Experimental Trial (Comm status)																	
BTS 8606	242	365.6	103	12597	108	0.78	62.28	105	2139	110	19.07	34.62	118	1450	176	0	74.6
BTS 8629	224	353.0	99	12078	103	0.77	58.47	98	1993	102	18.42	34.39	144	1333	193	0	80.2
BTS 8735	234	358.3	100	12238	105	0.72	60.08	101	2054	105	18.65	34.19	119	1231	190	0	74.4
BTS 8742	207	353.6	99	11489	98	0.92	58.66	98	1915	98	18.60	32.33	142	1503	261	0	76.9
BTS 8749	202	362.5	102	11449	98	0.84	61.35	103	1937	99	18.97	31.57	120	1493	215	0	80.1
BTS 8756	241	360.9	101	11965	102	0.95	60.84	102	2016	103	18.98	33.09	139	1593	264	0	78.9
BTS 8762	235	346.0	97	11987	103	0.88	56.39	95	1958	100	18.18	34.61	152	1545	212	0	82.8
BTS 8770	247	356.3	100	12474	107	0.80	59.50	100	2084	107	18.62	34.86	125	1473	184	0	71.4
BTS 8784	236	377.9	106	11675	100	0.75	65.94	111	2041	105	19.65	30.83	99	1319	196	0	77.6
BTS 8787	219	359.1	101	11492	98	0.80	60.33	101	1936	99	18.77	31.95	123	1436	199	0	76.5
BTS 8798	223	363.6	102	11725	100	0.78	61.68	103	1977	101	18.95	32.43	108	1320	217	0	81.6
Crystal 573RR	225	364.6	102	12361	106	0.78	61.96	104	2101	108	19.01	33.92	113	1341	213	0	75.4
Crystal 578RR	220	357.5	100	12755	109	0.82	59.85	100	2128	109	18.71	35.81	132	1494	192	0	78.1
Crystal 684RR	239	352.9	99	13397	115	0.87	58.45	98	2211	113	18.51	38.12	146	1528	215	0	76.3
Crystal 792RR	218	357.0	100	11890	102	0.81	59.71	100	1986	102	18.67	33.40	127	1361	228	0	82.5
Crystal 793RR	246	365.5	102	12885	110	0.75	62.25	104	2191	112	19.03	35.26	122	1310	194	0	79.9
Crystal 794RR	208	355.6	100	13006	111	0.79	59.29	99	2167	111	18.59	36.63	135	1389	200	0	72.0
Crystal 795RR	215	362.5	102	11255	96	0.88	61.35	103	1901	97	19.00	31.17	115	1456	252	0	74.3
Crystal 796RR	238	364.4	102	13710	117	0.80	61.91	104	2307	118	19.02	38.04	123	1454	194	0	81.6
Crystal 797RR	201	354.2	99	12478	107	0.82	58.83	99	2075	106	18.54	35.32	135	1567	170	0	68.3
Hilleshög HIL9708	222	355.8	100	11704	100	0.84	59.36	100	1952	100	18.64	32.84	140	1524	200	95	78.1
Hilleshög HIL9895	209	348.6	98	11809	101	0.84	57.19	96	1930	99	18.27	33.99	135	1521	200	0	80.0
Hilleshög HIL9920	203	368.3	103	11285	97	0.80	63.09	106	1926	99	19.23	30.81	155	1417	190	0	68.5
Hilleshög HIL9921	204	367.8	103	10899	93	0.85	62.90	106	1858	95	19.24	29.62	138	1411	240	0	78.2
Hilleshög HIL9922	231	350.3	98	11344	97	0.83	57.68	97	1874	96	18.36	32.16	132	1518	197	0	80.4
Hilleshög HIL9923	243	353.5	99	10108	87	0.93	58.63	98	1674	86	18.60	28.75	174	1483	264	0	60.8
Hilleshög HIL9924	237	358.1	100	10064	86	0.91	60.02	101	1683	86	18.81	28.18	124	1473	266	0	54.4
Maribo MA504	229	345.5	97	13333	114	0.81	56.25	94	2165	111	18.09	38.74	133	1422	209	0	85.5
Maribo MA611	245	347.7	97	11597	99	0.82	56.92	96	1896	97	18.21	33.30	126	1514	194	0	81.0
Maribo MA717	232	366.0	103	12128	104	0.87	62.38	105	2062	106	19.16	33.14	127	1492	224	0	75.9
Maribo MA718	221	361.3	101	10594	91	0.83	60.98	102	1783	91	18.90	29.35	127	1563	186	0	67.4
Maribo MA719	213	353.1	99	11744	101	0.94	58.51	98	1945	100	18.59	33.27	159	1500	269	0	67.4
SX RR1861	233	353.2	99	12204	105	0.79	58.55	98	2010	103	18.45	34.76	138	14			

Table 15. 2017 Performance of All RR Varieties - ACSC Official Trial

Scandia MN																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	344.9	102	9843	110	1.35	56.03	104	1597	112	18.59	28.66	191	1657	535	0	70.2
BTS 8337	116	351.7	104	10028	112	1.31	58.08	108	1652	116	18.90	28.68	231	1626	503	0	58.2
BTS 8363	130	323.8	96	9612	107	1.38	49.65	92	1478	103	17.58	29.62	293	1711	512	0	67.1
BTS 8500	119	332.7	99	10247	114	1.30	52.33	97	1614	113	17.92	30.76	237	1642	478	0	63.3
BTS 8512	109	336.9	100	9737	109	1.30	53.60	100	1553	109	18.14	28.91	240	1597	493	0	74.2
BTS 8524	101	322.3	96	9334	104	1.41	49.20	92	1429	100	17.53	28.85	256	1747	536	32	69.6
BTS 8572	111	349.0	103	9485	106	1.26	57.28	107	1558	109	18.72	27.15	183	1569	494	0	70.4
Crystal 093RR	115	348.2	103	9779	109	1.21	57.03	106	1601	112	18.63	28.13	156	1605	460	0	61.8
Crystal 101RR	107	323.8	96	8243	92	1.48	49.65	92	1262	88	17.66	25.40	330	1818	541	0	59.2
Crystal 246RR	108	326.8	97	9909	110	1.32	50.56	94	1519	106	17.66	30.56	282	1644	481	0	63.8
Crystal 247RR	131	335.4	99	9686	108	1.27	53.16	99	1534	107	18.04	28.97	256	1693	440	0	61.8
Crystal 355RR	104	344.4	102	8990	110	1.36	55.88	104	1600	112	18.58	28.80	259	1684	514	0	69.3
Crystal 467RR	129	333.9	99	10310	115	1.40	52.70	98	1633	114	18.09	30.91	328	1756	493	0	64.3
Crystal 572RR	126	355.1	105	9702	108	1.15	59.12	110	1606	112	18.91	27.41	178	1533	421	0	69.1
Crystal 744RR	110	323.5	96	9744	109	1.31	49.56	92	1496	105	17.48	30.07	250	1683	469	0	68.8
Crystal 966RR	128	345.6	102	9852	110	1.19	56.24	105	1595	112	18.47	28.64	248	1483	438	0	68.6
Hilleshoq 4302RR	105	336.2	100	7920	88	1.25	53.41	99	1262	88	18.07	23.32	277	1659	433	0	46.3
Hilleshoq 4448RR	112	337.4	100	9718	108	1.28	53.77	100	1541	108	18.15	29.00	245	1587	483	0	56.4
Hilleshoq 9528RR	123	343.2	102	9796	109	1.33	55.53	103	1580	110	18.49	28.53	243	1603	517	0	60.0
Hilleshoq HIL9707	114	311.5	92	8228	92	1.58	45.91	85	1216	85	17.16	26.37	336	1809	618	0	36.1
Maribo 109	122	339.8	101	7882	86	1.34	54.48	101	1232	86	18.33	22.48	253	1598	524	0	42.4
Maribo 305	106	337.2	100	9494	106	1.19	53.71	100	1515	106	18.05	28.12	230	1518	431	0	55.3
Maribo MA502	121	334.4	99	9176	102	1.45	52.86	98	1452	102	18.17	27.49	331	1739	537	0	63.7
SX Avalanche RR(858)	127	346.3	103	8593	96	1.23	56.45	105	1405	98	18.54	24.62	273	1626	417	0	62.6
SX Canyon RR	118	346.1	103	9169	102	1.26	56.40	105	1493	104	18.57	26.65	220	1677	455	0	58.6
SX Cruze RR	117	318.3	94	9867	110	1.40	47.97	89	1487	104	17.31	31.13	287	1594	551	0	64.8
SX Marathon RR(856)	102	336.5	100	8900	99	1.27	53.48	100	1414	99	18.09	26.47	231	1613	467	32	54.1
SX Winchester RR	125	324.2	96	8547	95	1.29	49.76	93	1309	92	17.50	26.36	271	1594	476	0	48.9
SV RR244TT	120	324.5	96	8971	100	1.38	49.84	93	1381	97	17.60	27.52	244	1705	528	0	52.1
SV RR333	124	331.6	98	9258	103	1.25	52.01	97	1448	101	17.83	27.97	222	1634	453	0	57.7
SV RR351	113	343.9	102	9324	104	1.18	55.73	104	1508	105	18.37	27.14	206	1604	409	0	57.8
RR Filler #01s	132	330.4	98	9087	101	1.45	51.65	96	1419	99	17.96	27.51	285	1838	526	0	67.2
RR Filler #01v	133	330.4	98	9113	102	1.50	51.65	96	1423	99	18.03	27.67	288	1894	555	0	66.0
Experimental Trial (Comm status)																	
BTS 8606	242	334.9	99	10428	116	1.34	53.06	99	1628	114	18.10	31.68	294	1653	516	0	69.9
BTS 8629	224	325.4	96	10310	115	1.28	50.30	94	1578	110	17.55	31.45	256	1541	500	0	72.3
BTS 8735	234	322.7	96	9987	111	1.21	49.50	92	1536	107	17.35	30.59	228	1442	477	0	62.1
BTS 8742	207	328.7	97	9558	107	1.47	51.23	95	1501	105	17.90	28.66	263	1732	588	0	74.2
BTS 8749	202	346.2	103	9542	106	1.28	56.29	105	1569	110	18.61	27.02	200	1598	506	0	72.3
BTS 8756	241	334.6	99	9635	107	1.47	52.94	99	1515	106	18.18	28.87	265	1754	577	0	74.6
BTS 8767	235	334.4	99	10126	113	1.33	52.88	98	1610	113	18.05	29.86	283	1707	474	0	66.8
BTS 8770	247	340.7	101	8716	97	1.25	54.73	102	1382	97	18.29	25.75	263	1732	426	0	51.6
BTS 8784	236	348.1	103	9355	104	1.19	56.86	106	1530	107	18.61	26.93	218	1499	452	0	62.9
BTS 8787	219	315.6	94	9455	105	1.30	47.44	88	1437	100	17.09	29.28	249	1624	483	0	60.2
BTS 8795	223	339.5	101	8856	99	1.28	54.39	101	1404	98	18.25	26.07	193	1503	529	0	68.0
Crystal 573RR	225	341.5	101	8956	100	1.32	54.95	102	1453	102	18.41	26.15	218	1603	523	0	59.0
Crystal 578RR	220	341.4	101	10035	112	1.23	54.94	102	1607	112	18.32	29.65	254	1656	434	0	71.1
Crystal 684RR	239	317.7	94	9871	110	1.38	48.04	89	1522	106	17.27	30.62	295	1756	492	0	61.7
Crystal 792RR	218	341.8	101	9187	102	1.20	55.03	102	1481	104	18.30	26.76	202	1500	457	0	62.5
Crystal 793RR	246	339.6	101	9330	104	1.25	54.39	101	1517	106	18.24	26.76	248	1549	463	0	66.8
Crystal 794RR	208	320.6	95	9588	107	1.22	48.90	91	1467	103	17.26	29.53	225	1533	464	0	53.9
Crystal 795RR	215	345.3	102	8858	99	1.29	56.06	104	1449	101	18.58	25.49	225	1647	485	0	67.2
Crystal 796RR	238	319.4	95	9667	108	1.28	48.52	90	1480	104	17.26	29.90	243	1650	473	0	67.2
Crystal 797RR	201	330.2	98	9731	108	1.29	51.67	96	1505	105	17.79	29.40	267	1703	454	0	55.5
Hilleshoq HIL9708	222	346.5	103	7915	88	1.21	56.40	105	1285	90	18.55	22.76	235	1559	459	0	61.3
Hilleshoq HIL9895	209	327.3	97	8339	93	1.35	50.83	95	1287	90	17.72	25.59	296	1652	506	95	61.7
Hilleshoq HIL9920	203	343.6	102	9498	106	1.13	55.55	103	1533	107	18.32	27.41	222	1528	394	95	63.3
Hilleshoq HIL9921	204	332.2	98	8711	97	1.28	52.26	97	1361	95	17.90	26.43	293	1504	496	0	69.5
Hilleshoq HIL9922	231	317.9	94	8353	93	1.44	48.13	90	1274	89	17.34	26.06	296	1697	555	0	65.8
Hilleshoq HIL9923	243	333.2	99	7036	78	1.59	52.55	98	1120	78	18.23	20.55	325	1765	635	0	48.4
Hilleshoq HIL9924	237	334.1	99	8175	91	1.47	52.80	98	1293	90	18.17	24.31	248	1651	616	0	55.9
Maribo MA504	229	335.3	99	9661	108	1.24	53.17	99	1529	107	18.01	28.56	247	1610	452	0	65.6
Maribo MA611	245	323.6	96	8964	100	1.35	49.77	93	1389	97	17.54	27.27	257	1620	529	0	64.1
Maribo MA717	232	338.9	100	9300	104	1.23	54.22	101	1477	103	18.17	27.15	206	1529	479	0	62.9
Maribo MA718	221	333.9	99	7570	84	1.33	52.76	98	1192	83	18.03	22.40	290	1799	462	0	53.9
Maribo MA719	213	337.7	100	9076	101	1.50	53.86	100	1430	100	18.37	27.04	280	1718	591	0	57.0
SX RR1861	233	335.6	99	9027	101	1.19	53.26	99	1444	101	17.99	26.58	207	1641	419	0	57.0
SX RR1863	244																

Table 16. 2017 Performance of All RR Varieties - ACSC Official Trial

Stephen MN																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	317.1	100	11965	104	1.28	47.62	99	1799	104	17.14	37.68	270	1677	446	0	74.0
BTS 8337	116	335.0	105	11955	104	1.13	53.02	110	1898	109	17.87	35.56	273	1608	349	0	69.8
BTS 8363	130	308.6	97	12386	107	1.13	45.05	94	1808	104	16.56	39.98	294	1585	350	0	74.5
BTS 8500	119	315.1	99	11956	104	1.27	47.02	98	1793	103	17.00	37.91	292	1690	421	0	66.4
BTS 8512	109	322.2	101	11539	100	1.28	49.17	102	1759	101	17.39	35.88	290	1686	433	0	74.0
BTS 8524	101	316.3	99	11814	102	1.14	47.39	99	1769	102	16.96	37.43	244	1695	346	0	74.0
BTS 8572	111	337.0	106	11989	104	1.16	53.65	112	1903	110	18.03	35.72	190	1483	439	0	71.9
Crystal 093RR	115	342.2	107	11992	104	1.10	55.21	115	1929	111	18.22	35.26	171	1555	381	0	72.1
Crystal 101RR	107	309.1	97	11554	100	1.25	45.19	94	1678	97	16.71	37.63	338	1797	366	0	69.5
Crystal 246RR	108	312.8	98	12029	104	1.22	46.33	96	1784	103	16.84	38.45	338	1638	387	0	67.2
Crystal 247RR	131	319.2	100	12945	112	1.05	48.24	100	1945	112	17.03	40.66	248	1598	302	0	72.9
Crystal 355RR	104	330.5	104	11825	102	1.14	51.66	108	1846	106	17.66	35.92	218	1540	392	0	63.3
Crystal 467RR	129	313.1	98	12786	111	1.22	46.41	97	1887	109	16.88	40.95	370	1653	371	0	70.1
Crystal 572RR	126	340.3	107	12457	108	1.08	54.63	114	2002	115	18.09	36.53	188	1387	401	0	73.4
Crystal 574RR	110	328.6	103	12633	109	1.10	51.09	106	1963	113	17.54	38.40	204	1574	365	0	74.7
Crystal 966RR	128	335.7	105	12149	105	1.03	53.25	111	1926	111	17.82	36.17	221	1401	346	0	73.2
Hillesq 4302RR	105	317.3	100	10814	94	1.10	47.67	99	1624	94	16.97	33.99	311	1623	308	0	64.6
Hillesq 4448RR	112	326.4	102	11548	100	1.20	50.42	105	1780	102	17.51	35.66	282	1544	411	0	63.0
Hillesq 9528RR	123	326.9	103	10890	94	1.17	50.58	105	1682	97	17.51	33.37	267	1559	396	0	70.8
Hillesq HIL9707	114	317.9	100	12537	109	1.03	47.87	100	1879	108	16.93	39.56	235	1509	316	0	71.1
Maribo 109	122	337.0	106	9997	87	1.08	53.65	112	1592	92	17.93	29.58	220	1505	353	0	65.6
Maribo 305	106	323.5	102	10678	93	1.01	49.56	103	1631	94	17.19	33.07	230	1483	304	0	59.1
Maribo MA502	121	311.2	98	10595	92	1.19	45.82	95	1553	89	16.75	34.16	333	1690	353	0	67.2
SX Avalanche RR(858)	127	323.9	102	11350	98	1.07	49.67	103	1737	100	17.27	35.13	280	1554	318	0	65.6
SX Canyon RR	118	339.0	106	12206	106	1.01	54.24	113	1949	112	17.95	36.23	177	1484	324	0	64.8
SX Cruze RR	117	304.0	95	11855	103	1.09	43.64	91	1694	98	16.30	39.09	244	1571	341	0	77.4
SX Marathon RR(856)	102	325.0	102	11969	104	1.12	49.99	104	1834	106	17.37	37.06	218	1565	367	0	67.7
SX Winchester RR	125	321.5	101	10882	94	1.07	48.95	102	1657	95	17.14	33.74	218	1608	320	0	55.7
SV RR244TT	120	316.7	99	11773	102	1.14	47.49	99	1769	102	16.97	37.19	253	1626	356	0	64.6
SV RR333	124	315.9	99	12112	105	1.16	47.24	98	1812	104	16.94	38.37	265	1646	363	0	63.8
SV RR351	113	318.4	100	11851	103	1.17	48.02	100	1788	103	17.08	37.17	264	1630	375	0	70.8
RR Filler #01s	132	305.5	96	11657	101	1.30	44.09	92	1679	97	16.58	38.14	372	1762	400	0	72.4
RR Filler #01v	133	320.2	101	11889	103	1.19	48.56	101	1800	104	17.20	37.19	281	1792	342	0	72.1
Experimental Trial (Comm status)																	
BTS 8606	242	335.7	105	13015	113	1.08	53.13	111	2051	118	17.94	38.86	208	1597	351	0	78.5
BTS 8629	224	321.2	101	13070	113	1.10	48.84	102	1976	114	17.19	41.03	267	1468	371	0	75.0
BTS 8735	234	321.5	101	12463	108	1.15	48.90	102	1883	108	17.25	39.15	282	1475	397	0	81.3
BTS 8742	207	325.6	102	10991	95	1.20	50.14	104	1681	97	17.54	34.02	232	1582	418	0	71.1
BTS 8749	202	328.9	103	11428	99	1.19	51.11	106	1773	102	17.64	34.73	254	1698	381	95	73.0
BTS 8756	241	327.5	103	11933	103	1.19	50.71	106	1836	106	17.61	36.74	225	1705	388	0	79.7
BTS 8767	235	331.0	104	12768	111	1.14	51.74	108	1978	114	17.67	38.90	246	1623	356	0	82.8
BTS 8770	247	326.9	103	11935	103	1.12	50.52	105	1832	106	17.48	36.85	274	1674	333	0	68.0
BTS 8784	236	344.4	108	11467	99	1.04	55.72	116	1845	106	18.29	33.72	185	1415	364	0	78.5
BTS 8787	219	318.9	100	11402	99	1.16	48.17	100	1717	99	17.11	35.88	248	1625	370	0	73.8
BTS 8798	223	327.0	103	10814	94	1.08	50.56	105	1651	95	17.48	33.52	198	1495	367	0	77.0
Crystal 573RR	225	327.6	103	11619	101	1.11	50.73	106	1788	103	17.48	35.64	233	1513	370	0	65.6
Crystal 578RR	220	325.1	102	13422	116	1.14	49.99	104	2052	118	17.37	41.58	256	1634	351	0	77.0
Crystal 684RR	239	326.1	102	12696	110	1.22	50.27	105	1947	112	17.55	39.25	253	1743	395	0	77.4
Crystal 792RR	218	336.1	106	12143	105	1.05	53.23	111	1911	110	17.87	36.46	179	1426	369	0	72.3
Crystal 793RR	246	336.9	106	12357	107	0.98	53.50	111	1959	113	17.83	36.85	185	1491	296	0	69.5
Crystal 794RR	208	334.7	105	12475	108	1.07	52.84	110	1960	113	17.86	37.39	205	1523	348	0	68.4
Crystal 795RR	215	328.3	103	11423	99	1.12	50.95	106	1772	102	17.56	34.89	245	1479	381	0	62.1
Crystal 796RR	238	329.7	104	13349	116	1.09	51.35	107	2068	119	17.62	40.69	204	1618	344	0	77.7
Crystal 797RR	201	314.9	99	12564	109	1.14	46.98	98	1858	107	16.96	40.20	322	1693	325	0	66.4
Hillesq HIL9708	222	323.0	101	10732	93	1.27	49.39	103	1639	94	17.42	33.41	323	1583	440	0	65.6
Hillesq HIL9895	209	307.0	96	10408	90	1.37	44.63	93	1509	87	16.68	33.90	313	1642	500	95	66.0
Hillesq HIL9920	203	334.3	105	12466	108	1.03	52.73	110	1960	113	17.73	37.58	209	1523	319	0	71.1
Hillesq HIL9921	204	340.6	107	9500	82	1.03	54.58	114	1517	87	18.03	28.14	226	1427	331	0	68.4
Hillesq HIL9922	231	311.1	98	11103	96	1.20	45.84	95	1632	94	16.78	35.76	272	1703	378	0	75.0
Hillesq HIL9923	243	328.8	103	9493	82	1.12	51.10	106	1458	84	17.62	29.28	263	1564	364	0	58.2
Hillesq HIL9924	237	313.2	98	9689	84	1.39	46.46	97	1429	82	17.05	31.10	276	1636	531	0	47.7
Maribo MA504	229	329.3	103	12510	108	1.20	51.22	107	1936	111	17.66	38.32	275	1554	408	0	66.8
Maribo MA611	245	317.1	100	11138	97	1.21	47.62	99	1658	95	17.07	35.46	284	1663	390	0	67.2
Maribo MA717	232	332.6	104	11139	97	1.04	52.23	109	1743	100	17.69	33.89	203	1519	319	0	71.5
Maribo MA718	221	309.8	97	10019	87	1.33	45.47	95	1465	84	16.81	32.48	432	1749	417	0	59.8
Maribo MA719	213	324.9	102	10445	91	1.37	49.95	104	1596	92	17.67	32.99	315	1654	495	0	54.3
SX RR1861	233	322.8	101	11780	102	0.95	49.31	103	1793	103	17.08	36.79	1				

Table 17. 2017 Performance of All RR Varieties - ACSC Official Trial

St Thomas ND																	
Variety @	Code	Rec/T lbs.	Rec/T %Bnch	Rec/A lbs.	Rec/A %Bnch	Loss Mol %	Rev/T \$++	Rev/T %Bnch	Rev/A \$++	Rev/A %Bnch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	306.3	100	9516	103	1.16	44.35	99	1378	103	16.47	31.05	245	1656	369	0	77.3
BTS 8337	116	326.6	106	9537	103	0.97	50.50	113	1474	110	17.30	29.23	185	1602	259	0	72.0
BTS 8363	130	296.9	97	9682	105	1.04	41.49	93	1350	101	15.88	32.66	272	1546	294	0	78.8
BTS 8500	119	306.1	100	9832	107	1.03	44.28	99	1424	106	16.34	32.12	238	1654	274	0	74.7
BTS 8512	109	311.0	101	9640	105	0.98	45.76	103	1419	106	16.53	31.00	215	1563	266	0	79.0
BTS 8524	101	305.2	99	9637	105	0.96	44.00	99	1389	104	16.22	31.59	200	1720	215	0	79.6
BTS 8572	111	319.0	104	9937	108	1.00	48.20	108	1498	112	16.95	31.22	191	1475	317	0	73.3
Crystal 093RR	115	326.0	106	10368	112	1.01	50.32	113	1598	119	17.30	31.84	162	1653	281	0	72.3
Crystal 101RR	107	301.9	98	9498	103	1.08	43.01	96	1353	101	16.18	31.46	276	1805	256	0	75.4
Crystal 246RR	108	299.2	97	9393	102	1.01	42.18	95	1324	99	15.96	31.38	265	1610	258	0	72.6
Crystal 247RR	131	298.4	97	9627	104	0.98	41.96	94	1352	101	15.90	32.31	276	1578	239	0	71.7
Crystal 355RR	104	307.9	100	9176	100	1.11	44.83	101	1334	100	16.51	29.84	237	1598	352	0	68.4
Crystal 467RR	129	303.0	99	10560	115	1.02	43.34	97	1509	113	16.17	34.89	350	1599	239	0	77.9
Crystal 572RR	126	336.5	110	9867	107	0.88	53.49	120	1568	117	17.71	29.32	159	1414	252	0	79.9
Crystal 574RR	110	303.2	99	9988	108	0.98	43.41	97	1429	107	16.14	32.96	205	1603	257	0	79.1
Crystal 986RR	128	314.0	102	9841	107	1.00	46.68	105	1464	109	16.70	31.34	270	1490	280	0	72.0
Hilleshög 4302RR	105	312.3	102	8678	94	0.94	46.17	104	1285	96	16.56	27.75	257	1499	239	0	65.8
Hilleshög 4448RR	112	314.0	102	10202	111	0.96	46.67	105	1515	113	16.67	32.54	203	1545	259	0	66.4
Hilleshög 9528RR	123	320.4	104	9585	104	0.96	48.60	109	1449	108	16.97	30.01	207	1484	270	0	71.5
Hilleshög HIL9707	114	306.8	100	10029	109	1.02	44.51	100	1453	109	16.37	32.73	277	1483	295	0	75.8
Maribo 109	122	322.5	105	8074	88	0.92	49.24	110	1235	92	17.04	25.00	190	1478	254	0	69.2
Maribo 305	106	303.4	99	9282	101	0.99	43.45	97	1332	100	16.16	30.56	209	1591	270	0	60.6
Maribo MA502	121	301.6	98	9820	107	1.04	42.93	96	1397	104	16.13	32.58	320	1613	265	0	71.5
SX Avalanche RR(858)	127	319.5	104	9279	101	0.89	48.35	108	1403	105	16.87	29.08	195	1502	217	0	70.7
SX Canyon RR	118	307.4	100	9589	104	0.96	44.67	100	1391	104	16.33	31.24	224	1558	251	0	66.0
SX Cruze RR	117	292.2	95	9975	108	1.04	40.07	90	1364	102	15.64	34.20	264	1564	291	0	81.9
SX Marathon RR(856)	102	308.3	100	10066	109	0.94	44.96	101	1470	110	16.36	32.60	218	1541	242	0	67.9
SX Winchester RR	125	304.4	99	8570	93	0.97	43.77	98	1231	92	16.19	28.17	249	1574	242	0	61.5
SV RR244TT	120	307.9	100	9396	102	0.98	44.83	101	1369	102	16.37	30.50	218	1618	245	0	66.9
SV RR333	124	310.4	101	9882	107	0.97	45.57	102	1452	109	16.48	31.78	204	1575	257	0	67.3
SV RR351	113	307.5	100	9590	104	0.97	44.72	100	1393	104	16.34	31.19	230	1552	256	0	72.6
RR Filler #01s	132	305.3	99	9754	106	1.08	44.05	99	1404	105	16.35	32.00	279	1744	274	0	73.1
RR Filler #01v	133	302.9	99	9836	107	1.13	43.31	97	1404	105	16.28	32.52	294	1811	291	0	73.7
Experimental Trial (Comm status)																	
BTS 8606	242	301.0	98	10022	109	1.11	42.77	96	1425	107	16.16	33.25	281	1678	310	0	80.3
BTS 8629	224	304.4	99	10108	110	1.01	43.80	98	1459	109	16.24	33.08	262	1503	281	0	80.8
BTS 8735	234	307.1	100	9017	98	1.02	44.62	100	1302	97	16.40	29.41	233	1493	319	0	72.7
BTS 8742	207	310.5	101	8851	96	1.15	45.58	102	1297	97	16.66	28.45	268	1665	362	0	62.4
BTS 8749	202	305.3	99	10144	110	1.06	44.05	99	1463	109	16.34	33.22	260	1721	281	0	68.5
BTS 8756	241	313.8	102	8911	97	1.18	46.59	104	1314	98	16.87	28.57	254	1665	377	0	78.1
BTS 8767	235	310.6	101	11021	120	1.03	45.63	102	1623	121	16.58	35.34	229	1646	282	0	84.3
BTS 8770	247	304.2	99	9306	101	1.10	43.73	98	1330	99	16.31	30.68	290	1759	285	0	71.7
BTS 8784	236	329.5	107	9371	102	0.97	51.25	115	1457	109	17.46	28.47	155	1476	303	0	71.4
BTS 8787	219	304.8	99	9810	106	1.06	43.89	98	1413	106	16.31	32.20	260	1666	296	0	66.8
BTS 8798	223	313.4	102	9759	106	0.95	46.48	104	1445	108	16.63	31.16	165	1486	282	0	76.1
Crystal 573RR	225	315.6	103	9714	105	0.97	47.12	106	1446	108	16.76	30.73	194	1514	278	0	66.8
Crystal 578RR	220	302.2	98	10312	112	1.10	43.16	97	1477	110	16.22	34.00	276	1659	312	0	78.9
Crystal 684RR	239	302.8	99	10555	115	1.11	43.33	97	1506	113	16.25	34.97	281	1698	304	0	75.8
Crystal 792RR	218	322.7	105	9730	106	0.98	49.24	110	1481	111	17.14	30.22	168	1513	300	0	76.4
Crystal 793RR	246	329.0	107	10087	109	0.94	51.11	115	1565	117	17.41	30.77	181	1452	265	0	68.4
Crystal 794RR	208	310.0	101	10473	114	1.01	45.45	102	1539	115	16.53	33.65	253	1576	273	0	64.1
Crystal 795RR	215	316.0	103	9524	103	1.04	47.22	106	1422	106	16.85	30.13	220	1632	297	0	68.6
Crystal 796RR	238	311.2	101	10972	119	1.01	45.78	103	1625	121	16.59	34.94	238	1694	252	0	79.5
Crystal 797RR	201	303.9	99	9322	101	1.10	43.64	98	1339	100	16.29	30.72	296	1735	285	0	66.5
Hilleshög HIL9708	222	319.2	104	9195	100	0.97	48.18	108	1384	103	16.94	28.94	226	1461	288	0	62.0
Hilleshög HIL9895	209	302.1	98	9542	104	0.99	43.11	97	1360	102	16.11	31.54	233	1601	256	0	67.4
Hilleshög HIL9920	203	324.1	106	10065	109	1.00	49.64	111	1538	115	17.23	31.10	235	1767	230	0	66.0
Hilleshög HIL9921	204	319.2	104	8292	90	0.99	48.19	108	1244	93	16.97	26.05	222	1493	297	0	71.8
Hilleshög HIL9922	231	299.7	98	9848	107	1.09	42.38	95	1395	104	16.07	32.79	259	1693	295	0	78.4
Hilleshög HIL9923	243	310.1	101	8199	89	1.19	45.47	102	1202	90	16.69	26.49	294	1652	376	0	56.5
Hilleshög HIL9924	237	302.2	98	6541	71	1.22	43.14	97	921	69	16.34	21.72	281	1684	408	0	50.2
Maribo MA504	229	308.4	100	10501	114	0.87	44.99	101	1536	115	16.31	33.89	210	1474	215	0	68.6
Maribo MA611	245	299.7	98	9137	99	1.05	42.39	95	1288	96	16.04	30.68	273	1600	286	0	75.5
Maribo MA717	232	320.2	104	9161	99	0.99	48.48	109	1388	104	17.01	28.66	202	1452	310	0	71.4
Maribo MA718	221	295.7	96	8616	93	1.18	41.20	92	1201	90	15.95	29.16	347	1706	337	0	63.9
Maribo MA719	213	315.4	103	9579	104	1.17	47.04	105	1430	107	16.93	30.39	258	1713	360	0	58.4
SX RR1861	233	306.7	100	9630	104	0.96	44.46	100	1398	105	16.30	31.33	192	1661	240	0	69.

Table 18. 2017 Performance of All RR Varieties - ACSC Official Trial

Bathgate ND																	
Variety @	Code	Rec/T lbs.	Rec/T %Bch	Rec/A lbs.	Rec/A %Bch	Loss Mol %	Rev/T \$++	Rev/T %Bch	Rev/A \$++	Rev/A %Bch	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter per Ac	Emerg %
Commercial Trial																	
BTS 80RR52	103	357.2	100	8690	92	0.96	59.77	100	1454	93	18.82	24.35	142	1725	233	0	95.4
BTS 8337	116	365.4	102	10117	107	0.87	62.22	104	1726	110	19.13	27.72	132	1657	186	0	97.5
BTS 8363	130	346.8	97	10163	108	0.89	56.62	95	1654	105	18.23	29.35	151	1666	198	0	97.7
BTS 8500	119	355.6	100	10477	111	0.90	59.26	99	1752	112	18.68	29.37	142	1664	207	0	97.0
BTS 8512	109	360.2	101	9697	103	0.91	60.65	102	1636	104	18.92	26.79	133	1660	213	0	97.8
BTS 8524	101	348.8	98	10381	110	0.94	57.21	96	1714	109	18.38	29.53	156	1806	196	0	98.8
BTS 8572	111	360.5	101	9419	100	0.86	60.77	102	1589	101	18.88	26.06	136	1542	203	0	94.5
Crystal 093RR	115	363.2	102	10585	112	0.89	61.56	103	1805	115	19.05	28.95	125	1572	226	0	96.2
Crystal 101RR	107	348.9	98	10592	113	0.98	57.23	96	1733	110	18.42	30.50	191	1829	204	0	96.4
Crystal 246RR	108	349.5	98	10093	107	0.86	57.41	96	1656	106	18.33	28.92	157	1574	190	0	95.8
Crystal 247RR	131	346.7	97	9605	102	0.86	56.58	95	1572	100	18.20	27.57	151	1677	171	32	95.2
Crystal 355RR	104	362.4	102	7872	84	0.94	61.34	103	1327	85	19.07	21.85	142	1665	235	0	95.9
Crystal 467RR	129	349.8	98	9703	103	0.91	57.51	96	1587	101	18.40	27.87	208	1719	177	0	95.7
Crystal 572RR	126	374.4	105	9922	105	0.81	64.96	109	1718	110	19.53	26.56	121	1488	190	0	98.3
Crystal 574RR	110	358.4	100	10388	110	0.90	60.12	101	1737	111	18.82	29.11	138	1630	212	0	96.1
Crystal 966RR	128	365.8	103	10692	114	0.84	62.36	105	1832	117	19.13	29.08	153	1621	197	0	98.1
Hillesq 4302RR	105	358.5	100	10492	111	0.86	60.16	101	1760	112	18.78	29.31	137	1657	178	0	93.1
Hillesq 4448RR	112	350.4	98	11531	123	0.86	57.70	97	1898	121	18.38	32.86	148	1601	192	0	94.8
Hillesq 9528RR	123	357.2	100	11312	120	0.86	59.75	100	1895	121	18.72	31.67	148	1584	186	32	96.3
Hillesq HIL9707	114	347.1	97	11076	118	0.91	56.70	95	1809	115	18.27	31.91	156	1710	197	0	96.7
Maribo 109	122	366.0	103	9431	100	0.90	62.41	105	1607	102	19.20	25.81	148	1580	225	0	93.3
Maribo 305	106	355.0	100	11777	125	0.80	59.08	99	1957	125	18.55	33.22	143	1469	181	0	96.1
Maribo MA502	121	353.7	99	9869	105	0.93	58.70	98	1624	104	18.62	28.21	197	1735	193	0	94.9
SX Avalanche RR(858)	127	363.1	102	9740	103	0.83	61.55	103	1651	105	18.98	26.81	135	1564	177	0	93.4
SX Canyon RR	118	355.0	100	10787	115	0.81	59.23	99	1799	115	18.58	30.28	133	1535	169	0	95.5
SX Cruze RR	117	342.3	96	10614	113	0.89	55.25	93	1712	109	18.00	30.98	144	1580	216	32	93.2
SX Marathon RR(856)	102	361.6	101	10903	116	0.82	61.10	102	1844	118	18.90	30.13	142	1555	168	0	93.9
SX Winchester RR	125	357.0	100	9729	103	0.89	59.86	100	1624	104	18.77	27.26	136	1615	211	0	95.1
SV RR244TT	120	352.5	99	10968	117	0.84	58.34	98	1810	115	18.47	31.21	149	1603	170	0	95.7
SV RR333	124	362.1	101	11322	120	0.83	61.24	103	1909	122	18.93	31.37	130	1588	176	0	95.3
SV RR351	113	355.5	100	10591	113	0.86	59.24	99	1762	112	18.63	29.92	165	1562	189	0	97.6
RR Filler #01s	132	355.6	100	10528	112	0.91	59.27	99	1752	112	18.68	29.69	165	1773	175	0	97.7
RR Filler #01v	133	356.8	100	11083	118	0.91	59.62	100	1856	118	18.75	30.96	160	1759	181	0	98.3
Experimental Trial (Comm status)																	
BTS 8606	242	363.0	102	10652	113	0.87	61.49	103	1790	114	19.02	29.65	140	1684	168	0	85.9
BTS 8629	224	346.0	97	9958	106	0.82	56.42	95	1611	103	18.13	29.02	131	1566	171	0	91.2
BTS 8735	234	352.5	99	9481	101	0.85	58.37	98	1569	100	18.49	27.00	147	1560	193	0	94.4
BTS 8742	207	356.6	100	8502	90	0.95	59.56	100	1407	90	18.76	24.07	140	1716	225	0	98.1
BTS 8749	202	350.9	98	8916	95	0.92	57.89	97	1465	93	18.47	25.64	151	1776	183	0	93.7
BTS 8756	241	361.4	101	9315	99	1.00	61.03	102	1567	100	19.07	26.03	161	1800	233	0	93.9
BTS 8767	235	346.7	97	9241	98	0.87	56.63	95	1485	95	18.21	27.33	133	1732	166	0	94.9
BTS 8770	247	353.4	99	9071	96	0.88	58.64	98	1504	96	18.57	25.83	129	1758	165	0	90.6
BTS 8784	236	367.2	103	8910	95	0.86	62.72	105	1507	96	19.22	24.51	141	1565	196	0	90.8
BTS 8787	219	350.5	98	9278	99	0.97	57.76	97	1523	97	18.49	26.61	157	1698	248	0	88.5
BTS 8798	223	354.9	99	8040	85	0.85	59.06	99	1324	84	18.60	23.15	118	1586	191	0	94.2
Crystal 573RR	225	355.4	100	8501	90	0.85	59.22	99	1399	89	18.62	24.15	124	1583	188	0	89.6
Crystal 578RR	220	353.3	99	9700	103	0.84	58.61	98	1583	101	18.52	28.02	142	1652	160	0	90.8
Crystal 684RR	239	345.8	97	9622	102	0.97	56.36	95	1577	101	18.26	27.72	155	1797	215	0	90.1
Crystal 792RR	218	361.1	101	9161	97	0.93	60.90	102	1539	98	18.98	25.58	169	1616	228	0	89.6
Crystal 793RR	246	359.2	101	9689	103	0.86	60.36	101	1623	103	18.83	27.29	137	1638	183	126	94.1
Crystal 794RR	208	355.8	100	10392	110	0.89	59.34	100	1721	110	18.68	29.51	132	1645	202	0	91.6
Crystal 795RR	215	364.0	102	8773	93	0.89	61.78	104	1471	94	19.08	24.53	134	1610	209	0	91.8
Crystal 796RR	238	356.4	100	10312	110	0.86	59.51	100	1725	110	18.69	29.15	137	1661	173	0	93.8
Crystal 797RR	201	353.3	99	10060	107	0.87	58.62	98	1661	106	18.54	28.64	141	1772	147	0	84.2
Hillesq HIL9708	222	364.8	102	9972	106	0.88	62.01	104	1689	108	19.12	27.39	149	1596	200	0	92.4
Hillesq HIL9895	209	333.9	94	8429	90	0.96	52.82	89	1312	84	17.65	25.95	161	1796	205	126	93.6
Hillesq HIL9920	203	368.4	103	10002	106	0.87	63.08	106	1695	108	19.27	27.46	137	1638	184	0	88.7
Hillesq HIL9921	204	368.9	103	8655	92	0.93	63.24	106	1481	94	19.37	23.57	137	1667	223	0	92.2
Hillesq HIL9922	231	349.3	98	8906	95	0.91	57.42	96	1449	92	18.38	25.78	164	1680	198	0	91.4
Hillesq HIL9923	243	359.4	101	9019	96	0.94	60.41	101	1499	96	18.92	25.54	168	1721	214	0	81.7
Hillesq HIL9924	237	363.9	102	8381	89	0.97	61.74	104	1404	90	19.16	23.48	183	1625	253	0	82.4
Maribo MA504	229	355.0	100	10574	112	0.89	59.12	99	1737	111	18.63	30.20	185	1598	192	0	95.9
Maribo MA611	245	349.9	98	8822	94	0.89	57.59	97	1443	92	18.40	25.57	153	1734	175	0	93.6
Maribo MA717	232	372.1	104	10611	113	0.86	64.20	108	1810	115	19.45	28.86	138	1603	181	0	89.1
Maribo MA718	221	347.5	97	8134	86	0.96	56.87	95	1328	85	18.34	23.50	182	1852	187	0	89.2
Maribo MA719	213	346.1	97	8862	94	0.97	56.46	95	1438	92	18.27	25.84	187	1724	231	0	93.9
SX RR1861	233	348.8	98	9182	98	0.92	57.26	96	1474	94	18.35	27.03	211	1601	206	0	96.6
SX RR18																	

Table 19. 2017 Performance of Conventional Varieties - ACSC Official Trials
6 sites - All Characters

*Unadjusted																	
Variety @	Code	Rec/T lbs.	Rec/T %Mean	Rec/A lbs.	Rec/A %Mean	Loss Mol %	Rev/T \$ ++	Rev/T %Mean	Rev/A \$ ++	Rev/A %Mean	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter /Ac	Emerg. %
BETA EXP 687	807	345.1	102	10123	100	1.22	56.11	104	1633	102	18.47	29.59	175	1690	431	0	72.4
BETA EXP 698	808	335.6	99	10304	101	1.14	53.21	99	1615	101	17.92	31.10	213	1659	363	0	75.9
BETA EXP 747	810	333.5	99	10556	104	1.15	52.59	98	1652	103	17.83	31.91	241	1510	405	0	74.8
BETA EXP 758	817	337.8	100	10331	102	1.13	53.88	100	1638	102	18.02	30.79	206	1655	358	0	78.1
Crystal 620	811	338.1	100	10783	106	1.15	53.96	100	1706	106	18.05	32.19	189	1600	401	0	69.4
Crystal 622	801	340.3	101	9650	95	1.25	54.64	102	1532	95	18.26	28.72	201	1681	444	0	66.4
Crystal 735	814	351.8	104	9832	97	1.09	58.13	108	1616	101	18.69	28.12	159	1535	382	0	67.8
Crystal 737	806	336.8	100	9878	97	1.25	53.57	100	1555	97	18.09	29.65	240	1680	436	0	69.4
Crystal R761	819	328.7	97	10896	107	1.28	51.12	95	1691	105	17.72	33.22	247	1813	422	0	74.1
Hilleshög 3035Rz	805	339.3	101	9182	90	1.21	54.33	101	1457	91	18.17	27.34	202	1625	429	18	80.4
Hilleshög 9891Rz	812	341.3	101	9268	91	1.19	54.95	102	1481	92	18.26	27.37	178	1637	425	0	76.6
Maribo MA615Rz	818	330.6	98	10191	100	1.27	51.71	96	1586	99	17.80	30.98	271	1709	430	0	80.0
Maribo MA720Rz	816	342.1	101	9919	98	1.13	55.19	103	1586	99	18.23	29.26	199	1558	390	0	83.6
Seedex 8869	809	338.4	100	10942	108	1.09	54.07	101	1741	108	18.02	32.49	197	1669	333	0	74.7
Seedex Duce	802	337.9	100	11246	111	1.10	53.90	100	1790	111	18.00	33.36	207	1692	329	18	74.8
Strube 12720	813	329.5	98	11314	111	1.11	51.36	96	1753	109	17.58	34.56	226	1687	328	0	75.0
Strube 13722	804	326.3	97	11043	109	1.15	50.40	94	1696	106	17.46	34.04	230	1777	333	0	79.2
SV 48611	815	343.2	102	10325	102	1.13	55.52	103	1669	104	18.30	30.12	191	1598	383	0	68.9
SV 48777	803	349.4	104	10409	103	1.02	57.39	107	1701	106	18.49	29.98	171	1627	296	0	72.0
Crystal 355RR(Check)	820	338.5	100	9880	97	1.24	54.10	101	1563	97	18.17	29.50	197	1680	444	0	75.5
BTS 80RR52(Check)	821	334.1	99	10171	100	1.24	52.77	98	1590	99	17.95	30.77	194	1685	443	0	77.6
Crystal 101RR (Check)	822	330.5	98	10855	107	1.28	51.68	96	1689	105	17.81	33.02	256	1780	426	0	73.1
Hilleshög 4302RR (Check)	823	336.3	100	9167	90	1.12	53.43	99	1446	90	17.94	27.45	233	1636	350	18	60.6
Maribo Ultramono(Filler)	824	330.6	98	7431	73	1.28	51.72	96	1152	72	17.81	22.69	293	1769	411	0	61.2
Benchmark Mean		334.9		10018		1.22	53.00		1572		17.97	30.19	220	1695	416		71.7
Trial Mean		337.3		10154		1.18	53.74		1606		18.04	30.34	213	1665	391		73.4
Coeff. of Var. (%)		2.4		6.5		7.2	4.5		7.3		2.1	6.5	18.5	4.7	13.5		9.9
Mean LSD (0.05)		7.2		718		0.08	2.18		128		0.34	2.06	35	59	50		5.7
Mean LSD (0.01)		9.5		949		0.10	2.89		169		0.45	2.72	46	77	66		7.5
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.																Created	11/01/2017
%Mean = percentage of trial mean.																Trial # =	17ACSCrv
© Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 20. 2017 Performance of Conventional Varieties - ACSC Official Trials

Casselton ND - All Characters

*Unadjusted																	
Variety @	Code	Rec/T lbs.	Rec/T %Mean	Rec/A lbs.	Rec/A %Mean	Loss Mbl %	Rev/T \$ ++	Rev/T %Mean	Rev/A \$ ++	Rev/A %Mean	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter /Ac	Emerg %
BETA EXP 687	807	349.9	102	8737	108	1.54	57.53	103	1438	110	19.02	24.91	157	2105	576	0	73.3
BETA EXP 698	808	351.0	102	7696	95	1.30	57.89	104	1273	97	18.84	21.83	175	2071	399	0	82.8
BETA EXP 747	810	343.3	100	8004	99	1.33	55.53	99	1297	99	18.50	23.28	183	1999	435	0	75.9
BETA EXP 758	817	344.2	100	7397	92	1.49	55.81	100	1198	92	18.71	21.50	199	2109	519	0	74.7
Crystal 620	811	348.5	101	8412	104	1.42	57.13	102	1378	105	18.84	24.16	161	2018	506	0	64.6
Crystal 622	801	355.8	103	8361	104	1.43	59.34	106	1396	107	19.23	23.49	145	2071	512	0	69.4
Crystal 735	814	356.0	103	7362	91	1.32	59.40	106	1226	94	19.12	20.70	135	1910	468	0	67.5
Crystal 737	806	348.9	101	8279	103	1.38	57.25	103	1358	104	18.83	23.76	196	2117	433	0	73.3
Crystal R761	819	327.2	95	8294	103	1.53	50.66	91	1288	98	17.88	25.26	215	2242	514	0	74.5
Hilleshög 3035Rz	805	345.6	100	7217	89	1.45	56.23	101	1177	90	18.72	20.88	166	2080	508	0	75.5
Hilleshög 9891Rz	812	343.8	100	7857	97	1.39	55.71	100	1273	97	18.59	22.85	160	2043	479	0	73.3
Maribo MA615Rz	818	328.9	96	7698	95	1.60	51.19	92	1200	92	18.04	23.35	240	2210	565	0	82.9
Maribo MA720Rz	816	354.0	103	7915	98	1.32	58.79	105	1312	100	19.04	22.43	144	1996	445	0	81.2
Seedex 8869	809	344.7	100	9405	116	1.44	55.96	100	1527	117	18.68	27.30	168	2184	477	0	78.7
Seedex Deuce	802	341.3	99	8644	107	1.40	54.93	98	1392	106	18.47	25.31	159	2205	441	0	77.3
Strube 12720	813	335.6	98	8936	111	1.44	53.20	95	1421	109	18.21	26.53	172	2152	475	0	72.8
Strube 13722	804	340.4	99	10491	130	1.37	54.66	98	1685	129	18.38	30.81	170	2198	416	0	70.6
SV 48611	815	349.5	102	8355	103	1.36	57.43	103	1372	105	18.84	23.89	140	2047	462	0	71.8
SV 48777	803	352.1	102	8521	106	1.24	58.21	104	1407	107	18.83	24.20	154	2092	347	0	68.1
Crystal 355RR(Check)	820	352.9	103	7070	88	1.37	58.46	105	1174	90	19.01	19.98	162	2076	456	0	72.2
BTS 80RR52(Check)	821	343.2	100	7596	94	1.50	55.53	99	1228	94	18.67	22.15	159	2182	527	0	73.0
Crystal 101RR (Check)	822	323.6	94	8275	102	1.62	49.58	89	1266	97	17.82	25.60	230	2270	570	0	72.3
Hilleshög 4302RR (Check)	823	341.0	99	6938	86	1.34	54.85	98	1112	85	18.41	20.48	177	2086	421	0	52.5
Maribo Ultramon(Filler)	824	340.3	99	6319	78	1.57	54.63	98	1013	77	18.58	18.62	260	2152	546	0	54.1
Benchmark Mean		340.2		7470		1.46	54.61		1195		18.48	22.05	182	2154	494		67.5
Trial Mean		344.2		8074		1.42	55.83		1309		18.64	23.47	176	2109	479		72.2
Coeff. of Var. (%)		2.1		6.2		5.7	4.0		6.9		1.8	5.9	12.5	2.3	11.3		7.9
Mean LSD (0.05)		11.2		795		0.12	3.40		144		0.51	2.20	31	75	82		8.3
Mean LSD (0.01)		14.9		1059		0.16	4.52		192		0.68	2.93	41	99	110		11.0
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.														Created 10/31/2017			
%Mean = percentage of trial mean.														Trial # = 178201			
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 21. 2017 Performance of Conventional Varieties - ACSC Official Trials

Hendrum MN - All Characters																	
*Unadjusted	Rec/T	Rec/T	Rec/A	Rec/A	Loss	Rev/T	Rev/T	Rev/A	Rev/A	Sugar	Yield	Na	K	AmN	Bolter	Emerg.	
Variety @	Code	lbs.	%Mean	lbs.	%Mean	Mbl %	\$ ++	%Mean	\$ ++	%Mean	%	T/A	ppm	ppm	ppm	/Ac	%
BETA EXP 687	807	324.4	101	11587	102	1.31	49.82	101	1780	103	17.54	35.74	188	1414	573	0	62.2
BETA EXP 698	808	317.6	99	11707	103	1.29	47.77	97	1757	102	17.17	36.96	274	1325	546	0	62.8
BETA EXP 747	810	317.0	98	11787	104	1.34	47.59	97	1770	102	17.20	37.18	312	1202	602	0	57.9
BETA EXP 758	817	320.9	100	11893	105	1.19	48.76	99	1816	105	17.24	36.89	195	1383	482	0	73.9
Crystal 620	811	320.1	99	11628	102	1.30	48.52	99	1762	102	17.32	36.40	222	1335	577	0	56.6
Crystal 622	801	319.7	99	10612	93	1.37	48.40	99	1599	92	17.34	33.34	209	1380	618	0	59.2
Crystal 735	814	339.2	105	10440	92	1.27	54.31	111	1671	97	18.24	30.81	200	1328	560	0	56.7
Crystal 737	806	312.4	97	10862	96	1.41	46.19	94	1598	92	17.03	34.95	340	1316	617	0	60.1
Crystal R761	819	320.4	100	11495	101	1.36	48.61	99	1746	101	17.38	35.82	253	1496	567	0	64.6
Hilleshög 3035Rz	805	323.1	100	10186	90	1.40	49.43	101	1562	90	17.56	31.49	229	1354	640	95	77.9
Hilleshög 9891Rz	812	327.1	102	10120	89	1.32	50.65	103	1566	91	17.68	31.01	181	1346	599	0	74.6
Maribo MA615Rz	818	316.0	98	11507	101	1.31	47.28	96	1718	99	17.11	36.44	319	1365	533	0	79.6
Maribo MA720Rz	816	322.0	100	10803	95	1.20	49.09	100	1649	95	17.30	33.51	246	1178	527	0	87.8
Seedex 8869	809	333.3	104	12467	110	1.11	52.51	107	1960	113	17.76	37.45	189	1306	440	0	74.4
Seedex Deuce	802	332.9	103	12261	108	1.12	52.39	107	1933	112	17.77	36.78	180	1402	431	0	74.9
Strube 12720	813	313.7	97	12729	112	1.20	46.59	95	1889	109	16.89	40.56	287	1461	440	0	75.1
Strube 13722	804	312.8	97	12459	110	1.25	46.30	94	1842	107	16.88	39.84	261	1521	467	0	80.7
SV 48611	815	333.9	104	10676	94	1.24	52.69	107	1691	98	17.93	31.84	197	1315	537	0	67.8
SV 48777	803	335.6	104	11728	103	1.07	53.22	108	1860	108	17.85	34.92	180	1376	396	0	75.8
Crystal 355RR(Check)	820	313.4	97	12059	106	1.44	46.50	95	1790	104	17.12	38.51	254	1406	651	0	72.6
BTS 80RR52(Check)	821	309.3	96	11950	105	1.49	45.26	92	1747	101	16.96	38.61	258	1364	695	0	74.2
Crystal 101RR (Check)	822	319.7	99	12145	107	1.41	48.40	99	1835	106	17.38	37.98	312	1497	581	0	73.4
Hilleshög 4302RR (Check)	823	320.6	100	10795	95	1.29	48.69	99	1636	95	17.32	33.69	274	1400	530	0	66.9
Maribo Ultramono(Filler)	824	322.1	100	8682	76	1.37	49.13	100	1327	77	17.48	26.95	321	1498	549	0	50.7
Benchmark Mean		315.8		11737		1.41	47.21		1752		17.20	37.20	274	1417	614		71.8
Trial Mean		322.0		11357		1.30	49.09		1729		17.39	35.32	245	1374	548		69.2
Coeff. of Var. (%)		2.8		5.7		8.9	5.5		6.5		2.3	6.3	18.6	6.7	13.9		15.7
Mean LSD (0.05)		13.2		966		0.17	3.99		169		0.60	3.24	65	138	115		16.2
Mean LSD (0.01)		17.5		1284		0.23	5.31		225		0.79	4.30	86	183	152		21.6
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.													Created 11/01/2017				
%Mean = percentage of trial mean.													Trial # = 178204				
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 22. 2017 Performance of Conventional Varieties - ACSC Official Trials

Grand Forks ND - All Characters

*Unadjusted																	
Variety @	Code	Rec/T lbs.	Rec/T %Mean	Rec/A lbs.	Rec/A %Mean	Loss Mol %	Rev/T \$ ++	Rev/T %Mean	Rev/A \$ ++	Rev/A %Mean	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter /Ac	Emerg %
BETA EXP 687	807	375.4	105	12865	101	0.89	65.28	109	2238	105	19.67	34.48	115	1607	218	0	71.0
BETA EXP 698	808	359.9	101	13661	107	0.86	60.59	101	2295	107	18.84	37.78	137	1657	175	0	75.9
BETA EXP 747	810	355.6	99	13165	103	0.76	59.27	99	2210	103	18.54	36.72	136	1369	179	0	78.5
BETA EXP 758	817	358.3	100	13535	106	0.83	60.08	100	2277	107	18.76	37.94	136	1541	185	0	75.7
Crystal 620	811	365.2	102	13266	104	0.78	62.17	104	2255	105	19.04	36.52	121	1467	170	0	73.9
Crystal 622	801	367.2	103	12194	95	0.88	62.77	105	2089	98	19.24	33.08	122	1530	234	0	69.2
Crystal 735	814	372.0	104	12857	100	0.77	64.22	107	2208	103	19.37	34.58	117	1419	182	0	68.8
Crystal 737	806	358.3	100	12181	95	0.92	60.10	100	2045	96	18.83	33.84	148	1641	226	0	70.1
Crystal R761	819	346.4	97	14029	110	0.90	56.50	94	2281	107	18.22	40.60	163	1667	194	0	78.4
Hilleshög 3035Rz	805	359.7	101	11074	87	0.82	60.52	101	1853	87	18.81	31.07	122	1544	180	0	85.9
Hilleshög 9891Rz	812	361.6	101	11123	87	0.84	61.08	102	1872	88	18.92	30.69	132	1544	197	0	80.9
Maribo MA615Rz	818	348.7	98	13330	104	0.92	57.20	96	2196	103	18.36	38.13	189	1615	216	0	78.2
Maribo MA720Rz	816	354.9	99	12595	98	0.86	59.07	99	2101	98	18.60	35.23	142	1569	199	0	87.9
Seedex 8869	809	351.7	98	14304	112	0.80	58.10	97	2342	110	18.39	41.12	124	1511	170	0	74.0
Seedex Deuce	802	362.4	101	14788	116	0.80	61.33	102	2464	115	18.92	41.50	130	1530	163	0	79.5
Strube 12720	813	344.6	96	13718	107	0.86	55.95	93	2227	104	18.09	39.91	167	1631	168	0	72.6
Strube 13722	804	347.6	97	12814	100	0.87	56.86	95	2084	97	18.25	37.09	134	1706	172	0	77.4
SV 48611	815	360.2	101	13690	107	0.77	60.65	101	2312	108	18.79	38.09	114	1414	182	0	69.0
SV 48777	803	363.6	102	13258	104	0.81	61.68	103	2251	105	18.99	36.47	139	1434	196	0	70.8
Crystal 355RR(Check)	820	358.8	100	12341	96	0.90	60.24	101	2072	97	18.83	34.17	126	1599	229	0	77.4
BTS 80RR52(Check)	821	354.3	99	12491	98	0.89	58.88	98	2096	98	18.61	34.90	143	1547	232	0	71.0
Crystal 101RR (Check)	822	342.2	96	12925	101	0.93	55.21	92	2067	97	18.03	38.03	197	1628	214	0	70.6
Hilleshög 4302RR (Check)	823	353.4	99	11559	90	0.83	58.61	98	1924	90	18.50	32.69	163	1522	179	0	67.6
Maribo Ultramo(Filler)	824	359.5	101	9283	73	0.96	60.44	101	1567	73	18.94	25.82	169	1745	223	0	67.1
Benchmark Mean		352.2		12329		0.89	58.24		2040		18.49	34.95	157	1574	214		71.7
Trial Mean		357.6		12794		0.85	59.87		2138		18.73	35.85	141	1560	195		74.6
Coeff. of Var. (%)		2.3		6.3		5.5	4.1		7.0		2.1	5.9	15.5	4.6	11.1		8.9
Mean LSD (0.05)		12.8		1230		0.07	3.87		222		0.60	3.36	33	111	34		9.6
Mean LSD (0.01)		17.0		1636		0.10	5.15		294		0.80	4.48	44	148	45		12.8
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.															Created	11/01/2017	
%Mean = percentage of trial mean.															Trial # =	178207	
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 23. 2017 Performance of Conventional Varieties - ACSC Official Trials

Scandia MN - All Characters

*Unadjusted																	
Variety @	Code	Rec/T lbs.	Rec/T %Mean	Rec/A lbs.	Rec/A %Mean	Loss Mol %	Rev/T \$ ++	Rev/T %Mean	Rev/A \$ ++	Rev/A %Mean	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter /Ac	Emerg. %
BETA EXP 687	807	350.8	104	9906	103	1.34	57.81	107	1634	107	18.88	28.21	197	1727	504	0	64.2
BETA EXP 698	808	321.0	95	9924	103	1.28	48.81	90	1530	100	17.35	30.71	245	1692	447	0	70.3
BETA EXP 747	810	328.1	97	10398	108	1.29	50.94	94	1609	105	17.69	31.56	272	1510	501	0	70.0
BETA EXP 758	817	345.3	102	9869	103	1.23	56.17	104	1599	104	18.50	28.81	224	1677	429	0	74.1
Crystal 620	811	332.0	98	10323	108	1.23	52.14	96	1616	105	17.83	31.09	209	1625	444	0	63.6
Crystal 622	801	345.1	102	9124	95	1.35	56.10	104	1490	97	18.62	26.38	219	1693	514	0	59.9
Crystal 735	814	357.1	105	9498	99	1.20	59.71	110	1596	104	19.06	26.52	177	1579	450	0	61.8
Crystal 737	806	347.4	103	9868	103	1.38	56.78	105	1593	104	18.73	28.66	244	1704	523	0	55.2
Crystal R761	819	326.7	96	9976	104	1.54	50.51	93	1538	100	17.87	30.60	307	1894	571	0	62.1
Hilleshög 3035Rz	805	340.1	100	8351	87	1.52	54.59	101	1339	87	18.53	24.67	294	1710	620	0	70.8
Hilleshög 9891Rz	812	344.7	102	9163	95	1.48	55.96	103	1484	97	18.71	26.62	226	1726	606	0	60.8
Maribo MA615Rz	818	326.5	96	9398	98	1.47	50.47	93	1453	95	17.80	28.74	327	1718	562	0	68.2
Maribo MA720Rz	816	350.4	103	9400	98	1.31	57.71	107	1531	100	18.81	26.99	225	1575	518	0	70.5
Seedex 8869	809	341.6	101	9668	101	1.20	55.02	102	1550	101	18.27	28.34	240	1707	388	0	55.5
Seedex Deuce	802	337.5	100	10238	107	1.26	53.79	99	1617	106	18.12	30.63	276	1703	422	95	51.3
Strube 12720	813	330.5	98	10852	113	1.24	51.67	95	1675	109	17.75	33.16	287	1702	408	0	66.4
Strube 13722	804	319.2	94	10529	110	1.21	48.24	89	1611	105	17.19	32.82	250	1780	376	0	74.9
SV 48611	815	351.2	104	10650	111	1.30	57.93	107	1774	116	18.88	30.19	205	1698	473	0	54.3
SV 48777	803	345.3	102	9278	97	1.12	56.14	104	1509	98	18.39	26.87	209	1628	356	0	61.9
Crystal 355RR(Check)	820	343.6	101	9538	99	1.35	55.64	103	1551	101	18.55	27.77	233	1716	506	0	66.4
BTS 80RR52(Check)	821	341.0	101	9948	104	1.31	54.86	101	1598	104	18.37	29.29	210	1707	490	0	74.7
Crystal 101RR (Check)	822	338.9	100	10585	110	1.52	54.22	100	1683	110	18.46	31.36	325	1895	558	0	65.9
Hilleshög 4302RR (Check)	823	346.5	102	7772	81	1.25	56.51	104	1258	82	18.57	22.50	286	1637	432	95	37.0
Maribo Ultramono(Filler)	824	319.6	94	6112	64	1.33	48.37	89	915	60	17.30	19.24	433	1716	419	0	58.2
Benchmark Mean		342.5		9461		1.36	55.31		1523		18.49	27.73	263	1739	496		61.0
Trial Mean		338.8		9599		1.32	54.17		1532		18.26	28.41	255	1697	480		63.3
Coeff. of Var. (%)		2.7		7.3		7.5	5.0		7.9		2.3	7.6	19.1	4.2	12.3		12.2
Mean LSD (0.05)		13.9		1011		0.15	4.21		171		0.65	3.20	69	108	90		11.3
Mean LSD (0.01)		18.5		1343		0.20	5.60		228		0.86	4.25	91	144	120		15.0
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.															Created 10/31/2017		
%Mean = percentage of trial mean.															Trial # = 178208		
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 24. 2017 Performance of Conventional Varieties - ACSC Official Trials

St Thomas ND - All Characters

*Unadjusted		Rec/T	Rec/T	Rec/A	Rec/A	Loss	Rev/T	Rev/T	Rev/A	Rev/A	Sugar	Yield	Na	K	AmN	Bolter	Emerg.
Variety @	Code	lbs.	%Mean	lbs.	%Mean	Mol %	\$ ++	%Mean	\$ ++	%Mean	%	T/A	ppm	ppm	ppm	/Ac	%
BETA EXP 687	807	291.4	101	9706	98	1.24	39.84	102	1324	99	15.80	33.51	268	1582	456	0	71.2
BETA EXP 698	808	294.5	102	11083	112	1.15	40.78	104	1536	115	15.87	37.67	346	1572	353	0	69.6
BETA EXP 747	810	284.0	98	10363	105	1.26	37.60	96	1382	103	15.48	36.27	454	1439	433	0	72.9
BETA EXP 758	817	293.3	101	10091	102	1.13	40.39	103	1383	103	15.78	34.60	374	1573	320	0	76.8
Crystal 620	811	286.7	99	10876	110	1.25	38.40	98	1451	108	15.58	38.08	327	1541	440	0	64.1
Crystal 622	801	280.7	97	9685	98	1.37	36.58	93	1256	94	15.39	34.69	390	1617	485	0	56.8
Crystal 735	814	307.9	106	10330	104	1.12	44.83	114	1506	112	16.54	33.38	251	1430	400	0	64.2
Crystal 737	806	287.5	99	9781	99	1.37	38.65	98	1310	98	15.73	34.17	387	1557	504	0	64.1
Crystal R761	819	284.2	98	10483	106	1.31	37.64	96	1390	104	15.53	36.80	408	1709	408	0	73.2
Hilleshög 3035Rz	805	295.3	102	10075	102	1.08	41.02	105	1402	105	15.84	34.21	296	1455	346	0	77.1
Hilleshög 9891Rz	812	295.8	102	9414	95	1.18	41.16	105	1307	97	15.97	31.78	299	1489	407	0	74.9
Maribo MA615Rz	818	288.6	100	9614	97	1.29	38.97	99	1287	96	15.72	33.54	403	1545	443	0	75.0
Maribo MA720Rz	816	297.0	103	10242	103	1.09	41.54	106	1433	107	15.92	34.62	316	1437	344	0	76.4
Seedex 8869	809	287.7	99	10208	103	1.15	38.71	99	1378	103	15.54	35.32	363	1575	335	0	72.9
Seedex Deuce	802	282.5	98	10236	103	1.05	37.14	95	1350	101	15.17	36.22	386	1511	275	0	71.9
Strube 12720	813	287.2	99	10825	109	0.99	38.57	98	1458	109	15.36	37.70	343	1449	265	0	68.4
Strube 13722	804	274.4	95	9594	97	1.17	34.67	88	1211	90	14.90	34.93	445	1609	315	0	76.6
SV 48611	815	290.4	100	8803	89	1.18	39.54	101	1187	89	15.72	30.34	357	1431	397	0	64.9
SV 48777	803	314.0	109	10381	105	0.99	46.66	119	1540	115	16.68	33.06	249	1496	273	0	62.6
Crystal 355RR(Check)	820	288.5	100	9487	96	1.28	38.96	99	1284	96	15.70	32.88	294	1585	460	0	69.5
BTS 80RR52(Check)	821	289.5	100	9958	101	1.20	39.26	100	1343	100	15.67	34.53	297	1574	400	0	76.0
Crystal 101RR (Check)	822	289.0	100	10419	105	1.17	39.11	100	1411	105	15.63	35.90	365	1634	335	0	63.9
Hilleshög 4302RR (Check)	823	280.6	97	8270	84	1.11	36.57	93	1077	80	15.14	29.54	408	1518	304	0	57.3
Maribo Ultramon(Filler)	824	275.9	95	7752	78	1.34	35.14	90	989	74	15.15	27.96	450	1618	433	0	59.0
Benchmark Mean		286.9		9534		1.19	38.48		1279		15.54	33.21	341	1578	375		66.7
Trial Mean		289.4		9903		1.19	39.24		1341		15.66	34.24	353	1539	380		69.1
Coeff. of Var. (%)		2.5		5.6		6.6	5.6		6.8		2.1	5.7	16.2	6.3	11.6		8.7
Mean LSD (0.05)		11.5		845		0.12	3.47		142		0.50	2.98	92	146	71		8.8
Mean LSD (0.01)		15.3		1123		0.17	4.62		189		0.66	3.97	122	194	94		11.7
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.																Created	10/31/2017
%Mean = percentage of trial mean.																Trial # =	178211
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 25. 2017 Performance of Conventional Varieties - ACSC Official Trials

Humboldt MN - All Characters

*Unadjusted																	
Variety @	Code	Rec/T lbs.	Rec/T %Mean	Rec/A lbs.	Rec/A %Mean	Loss Mol %	Rev/T \$ ++	Rev/T %Mean	Rev/A \$ ++	Rev/A %Mean	Sugar %	Yield T/A	Na ppm	K ppm	AmN ppm	Bolter /Ac	Emerg. %
BETA EXP 687	807	377.8	102	7746	84	0.95	66.00	103	1367	86	19.84	20.22	100	1730	248	0	92.3
BETA EXP 698	808	370.9	100	7566	82	0.93	63.91	99	1298	82	19.48	20.54	97	1644	251	0	93.6
BETA EXP 747	810	372.2	100	9844	107	0.96	64.29	100	1710	108	19.56	26.22	104	1559	300	0	94.6
BETA EXP 758	817	362.1	97	8780	96	0.87	61.23	95	1483	94	18.98	24.32	112	1646	202	0	92.9
Crystal 620	811	374.6	101	10322	112	0.93	65.02	101	1792	113	19.66	27.54	99	1615	256	0	95.0
Crystal 622	801	374.9	101	7968	87	1.02	65.12	101	1379	87	19.77	21.31	109	1795	272	0	84.3
Crystal 735	814	376.6	101	8681	94	0.91	65.64	102	1515	96	19.74	22.99	101	1537	266	0	87.5
Crystal 737	806	368.1	99	8327	91	1.04	63.07	98	1418	89	19.45	22.79	117	1722	297	0	94.9
Crystal R761	819	368.1	99	11299	123	1.05	63.06	98	1932	122	19.46	30.78	115	1877	276	0	91.9
Hilleshög 3035Rz	805	371.0	100	7994	87	0.94	63.93	99	1378	87	19.48	21.55	102	1618	262	0	94.7
Hilleshög 9891Rz	812	375.1	101	8117	88	0.96	65.18	101	1398	88	19.72	21.89	100	1662	261	0	95.9
Maribo MA615Rz	818	376.6	101	9464	103	1.00	65.63	102	1654	104	19.83	25.09	114	1819	253	0	94.8
Maribo MA720Rz	816	376.2	101	8543	93	0.99	65.51	102	1482	94	19.80	22.80	134	1565	295	0	97.7
Seedex 8869	809	372.6	100	9572	104	0.89	64.42	100	1655	104	19.52	25.71	97	1742	197	0	93.4
Seedex Deuce	802	371.1	100	11330	123	0.99	63.96	99	1949	123	19.55	30.64	110	1803	253	0	94.1
Strube 12720	813	366.1	98	10569	115	0.91	62.44	97	1801	114	19.21	28.92	104	1735	209	0	95.2
Strube 13722	804	363.2	98	10150	110	1.00	61.58	96	1725	109	19.16	27.90	108	1850	247	0	94.5
SV 48611	815	372.9	100	9615	105	0.95	64.52	100	1662	105	19.60	25.88	103	1717	242	0	85.5
SV 48777	803	385.7	104	9470	103	0.93	68.40	106	1675	106	20.22	24.59	103	1733	222	0	94.3
Crystal 355RR(Check)	820	375.4	101	9052	99	1.09	65.26	101	1565	99	19.86	24.24	101	1675	357	0	95.4
BTS 80RR52(Check)	821	367.9	99	8703	95	1.04	62.98	98	1491	94	19.44	23.68	110	1730	308	0	97.1
Crystal 101RR (Check)	822	372.1	100	11134	121	1.03	64.27	100	1917	121	19.63	30.01	107	1701	303	0	93.3
Hilleshög 4302RR (Check)	823	377.0	101	9890	108	0.88	65.75	102	1715	108	19.73	26.37	104	1643	214	0	84.4
Maribo Ultramono(Filler)	824	365.3	98	6395	70	1.09	62.22	97	1082	68	19.36	17.65	117	1900	297	0	77.9
Benchmark Mean		373.1		9695		1.01	64.57		1672		19.67	26.08	106	1687	296		92.5
Trial Mean		372.2		9189		0.97	64.31		1585		19.58	24.74	107	1709	262		92.3
Coeff. of Var. (%)		1.8		6.6		7.0	3.1		7.3		1.6	6.0	12.0	4.7	18.4		3.3
Mean LSD (0.05)		11.3		1143		0.12	3.43		217		0.53	2.84	22	132	79		5.4
Mean LSD (0.01)		15.2		1534		0.16	4.59		291		0.70	3.81	29	176	106		7.2
Sig Lvl		**		**		**	**		**		**	**	**	**	**		**
*Actual data output without adjustment factor.															Created 11/01/2017		
%Mean = percentage of trial mean.															Trial # = 178212		
@ Some varieties not approved for sale. Refer to approval list for approval status.																	
++ Revenue estimates are based on a \$48.49 beet payment at 17.5% sugar & 1.5% loss to molasses and does not consider hauling costs.																	

Table 26. Calculation for Approval of Sugarbeet Varieties for ACSC Market for 2018

Variety	Approval Status	Rec/Ton				Rev/Acre				R/T + S/A	Cercospora Rating +				
		2016	2017	2 Yr	Bench	2016	2017	2 Yr	Bench		Bench	2015	2016	2017	2 Yr Mean
Previously Approved (3 Yr)															
BTS 80RR52	Approved	316.8	334.2	325.5	100.6	1960	1899	1830	103.2	203.8	4.11	4.28	4.37	4.26	<=5.40
BTS 8337	Approved	325.2	349.5	337.4	104.3	1877	1842	1860	104.9	209.2	4.49	4.62	4.36	4.49	
BTS 8363	Approved	309.8	328.7	319.3	98.7	1937	1770	1854	104.6	203.2	3.83	4.33	4.10	4.09	
BTS 8500	Approved	308.7	335.7	322.2	99.6	1966	1862	1914	108.0	207.6	4.45	4.54	4.29	4.43	
BTS 8512	Approved	315.8	339.9	327.9	101.3	1917	1749	1833	103.4	204.8	4.12	4.04	3.69	3.95	
BTS 8524	Approved	305.7	330.0	317.9	98.2	1954	1796	1875	105.8	204.0	4.40	4.74	4.38	4.51	
BTS 8572	Approved	323.3	346.7	335.0	103.5	1913	1817	1865	105.2	208.8	4.60	4.41	4.14	4.38	
Crystal 093RR	Approved	319.1	350.3	334.7	103.5	1942	1866	1904	107.4	210.9	4.76	4.95	4.49	4.73	
Crystal 101RR	Approved	306.3	329.3	317.8	98.2	1849	1718	1784	100.6	198.9	4.65	4.59	4.57	4.60	
Crystal 246RR	Approved	305.3	331.7	318.5	98.4	1845	1775	1810	102.1	200.6	4.49	4.81	4.63	4.64	
Crystal 247RR	Approved	314.5	335.2	324.9	100.4	2014	1832	1923	108.5	208.9	4.19	4.65	4.55	4.47	
Crystal 355RR	Approved	322.3	340.0	331.2	102.4	1947	1711	1829	103.2	205.5	4.43	4.60	4.36	4.46	
Crystal 467RR	Approved	301.0	330.1	315.6	97.5	1845	1804	1825	102.9	200.5	4.34	4.69	4.46	4.49	
Crystal 572RR	Approved	324.7	354.7	339.7	105.0	1982	1891	1937	109.3	214.3	4.65	4.57	4.27	4.50	
Crystal 573RR	Approved	321.4	343.9	332.7	102.8	1970	1785	1878	105.9	208.7	4.15	4.35	4.15	4.22	
Crystal 574RR	Approved	307.8	334.4	321.1	99.3	2070	1875	1973	111.3	210.5	4.30	4.51	4.35	4.39	
Crystal 578RR	Approved	316.6	338.4	327.5	101.2	2017	1899	1958	110.5	211.7	4.93	4.87	4.91	4.91	
Crystal 986RR	Approved	318.8	341.1	330.0	102.0	1895	1776	1836	103.6	205.5	4.97	4.75	4.77	4.83	
Hilleshög HIL9708	Approved	312.4	338.6	325.5	100.6	1957	1840	1749	98.6	199.3	5.04	4.74	4.61	4.80	
Hilleshög 4302RR	Approved	317.4	334.0	325.7	100.7	1801	1597	1699	95.9	196.5	4.13	4.13	3.93	4.06	
Hilleshög 4448RR	Approved	309.1	338.0	323.6	100.0	1873	1829	1851	104.4	204.4	5.29	5.21	5.28	5.26	
Hilleshög 9528RR	Approved	319.1	339.3	329.2	101.8	1982	1785	1884	106.3	208.0	5.16	4.73	4.99	4.96	
Maribo 109	Approved	332.4	347.6	340.0	105.1	1889	1569	1729	97.5	202.6	4.56	4.14	4.14	4.28	
Maribo 305	Approved	307.5	331.7	319.6	98.8	1773	1731	1752	98.8	197.6	4.76	4.72	4.98	4.82	
Maribo MA504	Approved	305.5	333.9	319.7	98.8	1929	1830	1880	106.0	204.9	5.25	5.04	5.50	5.26	
SV RR244TT	Approved	317.6	334.7	326.2	100.8	1877	1796	1837	103.6	204.4	4.17	4.46	4.85	4.49	
SV RR333	Approved	318.3	338.9	328.6	101.6	1950	1823	1887	106.4	208.0	4.54	4.85	4.84	4.74	
SV RR351	Approved	313.2	337.3	325.3	100.5	1971	1783	1877	105.9	206.4	4.62	4.50	4.41	4.51	
SX Avalanche RR(858)	Approved	320.7	342.2	331.5	102.4	1916	1690	1803	101.7	204.2	4.15	4.74	4.64	4.51	
SX Canyon RR	Approved	317.4	342.4	329.9	102.0	1926	1829	1878	105.9	207.9	4.02	4.76	4.92	4.56	
SX Cruze RR	Approved	299.6	318.4	309.0	95.5	1712	1696	1704	96.1	191.6	4.57	4.65	5.37	4.87	
SX Marathon RR(856)	Approved	315.4	340.4	327.9	101.4	2039	1812	1926	108.6	210.0	5.37	4.44	4.54	4.78	
SX Winchester RR	Approved	320.5	331.1	325.8	100.7	1831	1580	1706	96.2	196.9	3.67	3.97	4.07	3.90	
Candidates for Approval (2 Yr)															
BTS 8606	Approved	317.3	340.5	328.9	101.7	2000	1882	1941	109.5	211.2	--	5.12	4.73	4.92	--
BTS 8629	Approved	307.5	332.8	320.2	99.0	1955	1884	1920	108.3	207.3	--	4.99	4.29	4.44	--
Crystal 694RR	Approved	308.1	333.7	320.9	99.2	2111	1899	2005	113.1	212.3	--	4.57	4.34	4.45	--
Hilleshög HIL9707	Not Approved	305.2	324.3	314.8	97.3	1739	1692	1716	96.8	194.1	4.60	4.53	4.96	4.74	4.70
Hilleshög HIL9895	Not Approved	313.7	326.3	320.0	98.9	1873	1547	1710	96.5	195.4	--	4.49	4.84	4.67	--
Maribo MA502	Not Approved	302.7	329.8	316.3	97.8	1825	1642	1734	97.8	195.6	5.04	4.79	5.01	4.90	4.95
Maribo MA611	Not Approved	313.1	325.9	319.5	98.8	1765	1542	1654	93.3	192.0	--	4.47	5.03	4.75	--
SX RR 1861	Approved	316.2	336.3	325.8	100.7	1966	1748	1857	104.8	205.5	--	4.52	4.74	4.63	--
SX RR 1863	Approved	323.4	342.4	332.9	102.9	2006	1773	1890	106.6	209.5	--	4.35	4.08	4.21	--
SV RR265	Approved	315.1	336.8	326.0	100.7	1979	1836	1908	107.6	208.4	--	5.00	5.19	5.09	--
SV RR266	Approved	317.3	337.9	327.6	101.3	1971	1814	1893	106.8	208.0	--	4.74	4.61	4.67	--
SV RR268	Approved	319.0	341.1	330.1	102.0	1954	1802	1878	106.0	208.0	--	5.13	5.06	5.10	--
Benchmark Varieties															
Crystal 875RR	Benchmark	2015	2016	2017		2015	2016	2017							
BTS 81RR17(Check)	Benchmark	308.5				1490									
BTS 80RR52	Benchmark	307.6	310.2			1574	1845								
Hilleshög 4302RR	Benchmark	317.7	316.8	334.2		1701	1960	1699							
Crystal 101RR	Benchmark	319.5	317.4	334.0		1624	1801	1597							
Crystal 355RR	Benchmark	306.3	329.3			1849	1718								
	Benchmark	340.0				1711									
Benchmark mean		313.3	312.7	334.4	323.5	1597	1864	1681	1773						

All Cercospora ratings 2015-2017 were adjusted to 1982 basis.
 Variety approval criteria include: 1) 2 years of official trial data, 2) Cercospora rating must not exceed 5.20 (1982 adjusted data), 3a) R/T >= 100% of Bench or 3b) R/T >= 97% and R/T + S/A >= 202% of Bench. 3 yrs of data may be considered for initial approval.
 Bench for 2017 added: Crystal 355RR and dropped BTS 81RR17(Check).
 To maintain approval, the 3-year Cercospora rating must not exceed 5.40 (1982 adjusted data).

Created 11-04-2017.

Table 27. Projected Calculation for Approval of Sugarbeet Varieties for ACSC Market

Variety	Approval ^	Rec/Ton		Rev/Acre		R/T +	CR Rating ^
		%		%		\$/A	
		2017	Bench	2017	Bench	Bench	2017
Candidates for Retesting (1 Yr)							
BTS 8735	On Track	335.7	100.4	1836	109.2	209.6	4.22
BTS 8742	Not On Track	333.4	99.7	1646	97.9	197.6	4.36
BTS 8749	On Track	337.7	101.0	1719	102.2	203.2	4.05
BTS 8756	On Track	338.4	101.2	1724	102.5	203.7	4.01
BTS 8767	On Track	339.2	101.4	1878	111.7	213.1	4.16
BTS 8770	On Track	337.4	100.9	1801	107.1	208.0	4.30
BTS 8784	On Track	351.4	105.1	1787	106.3	211.4	3.65
BTS 8787	On Track	331.5	99.1	1733	103.1	202.2	4.03
BTS 8798	On Track	338.8	101.3	1695	100.8	202.1	4.30
Crystal 792RR	On Track	344.0	102.9	1799	107.0	209.9	3.94
Crystal 793RR	On Track	347.5	103.9	1896	112.8	216.7	3.93
Crystal 794RR	On Track	333.8	99.8	1835	109.1	209.0	4.92
Crystal 795RR	On Track	340.1	101.7	1708	101.6	203.3	4.39
Crystal 796RR	On Track	337.0	100.8	1950	116.0	216.8	4.85
Crystal 797RR	On Track	330.1	98.7	1809	107.6	206.3	4.17
Hilleshög HIL9920	On Track	347.2	103.8	1785	106.2	210.0	4.89
Hilleshög HIL9921	On Track	345.2	103.2	1585	94.3	197.5	4.47
Hilleshög HIL9922	Not On Track	325.4	97.3	1560	92.8	190.1	4.02
Hilleshög HIL9923	On Track	337.5	100.9	1497	89.0	190.0	4.81
Hilleshög HIL9924	On Track	335.0	100.2	1455	86.5	186.7	4.09
Maribo MA717	On Track	342.0	102.3	1742	103.6	205.9	4.85
Maribo MA718	Not On Track	330.0	98.7	1476	87.8	186.5	4.39
Maribo MA719	On Track	337.1	100.8	1617	96.2	197.0	4.41
SX RR1875	On Track	341.6	102.2	1605	95.5	197.6	4.06
SX RR1876	Not On Track	332.6	99.5	1694	100.8	200.2	4.31
SX RR1877	Not On Track	330.0	98.7	1626	96.7	195.4	4.62
SX RR1878	On Track	335.6	100.4	1756	104.4	204.8	4.71
SX RR1879	On Track	338.5	101.2	1770	105.3	206.5	4.88
SV RR371	On Track	339.0	101.4	1833	109.0	210.4	4.59
SV RR372	On Track	332.7	99.5	1723	102.5	202.0	4.23
SV RR373	Not On Track	331.8	99.2	1613	95.9	195.2	4.31
SV RR374	On Track	337.2	100.8	1776	105.6	206.5	4.71
SV RR375	Not On Track	342.4	102.4	1802	107.2	209.6	5.08
Benchmark Varieties							
BTS 80RR52		334.2	99.9	1699	101.1		
Hilleshög 4302RR		334.0	99.9	1597	95.0		
Crystal 101RR		329.3	98.5	1718	102.2		
Crystal 355RR		340.0	101.7	1711	101.8		
Benchmark Mean		334.4		1681			
^ = not on track for approval. On Track = data is tracking for potential approval.							
^^ All Cercospora ratings 2017 were adjusted to 1982 basis.							
Full market approval criteria include: 1) 2 years of official trial data, 2) Cercospora rating must not exceed 5.00 (1982 adjusted data),							
3a) R/T >= 100% of Bench or 3b) R/T >= 97% and R/T + \$/A equal to 202 of Bench.							
Bench for 2017 added Crystal 355RR and dropped BTS 81RR17(Check).							
							Created 11-04-2017.

Table 28. Calculation for Approval of Sugarbeet Varieties for ACSC Aphanomyces Specialty Market for 2018

Trial Yrs	Variety	Approval Status	Root Aph. Rating					Cercospora Rating +				
			2015	2016	2017	2 Yr	3 Yr	2015	2016	2017	2 Yr	3 Yr
Previously Approved (3 Yrs)			<=4.70					<=5.40				
8	BTS 80RR52	Approved	3.24	4.11	4.36	4.24	3.90	4.11	4.28	4.37	4.33	4.25
5	BTS 8337	Approved	2.55	3.26	3.78	3.52	3.20	4.49	4.62	4.36	4.49	4.49
3	BTS 8500	Approved	3.54	4.22	4.52	4.37	4.09	4.45	4.54	4.29	4.42	4.43
3	BTS 8512	Approved	3.91	4.17	3.78	3.98	3.95	4.12	4.04	3.69	3.87	3.95
3	BTS 8524	Approved	3.33	3.89	4.49	4.19	3.90	4.40	4.74	4.38	4.56	4.51
3	BTS 8572	Approved	4.05	4.46	3.76	4.11	4.09	4.60	4.41	4.14	4.28	4.38
8	Crystal 093RR	Approved	3.86	4.32	4.43	4.38	4.20	4.76	4.95	4.49	4.72	4.73
7	Crystal 101RR	Approved	3.31	3.42	3.92	3.67	3.55	4.65	4.59	4.57	4.58	4.60
5	Crystal 355RR	Approved	3.26	4.46	4.84	4.65	4.19	4.43	4.60	4.36	4.48	4.46
4	Crystal 467RR	Approved	3.55	4.04	3.96	4.00	3.85	4.34	4.69	4.46	4.58	4.50
3	Crystal 573RR	Approved	3.69	4.06	3.84	3.95	3.86	4.15	4.35	4.15	4.25	4.22
3	Crystal 574RR	Approved	2.93	3.69	4.72	4.21	3.78	4.30	4.51	4.35	4.43	4.39
9	Crystal 986RR	Approved	3.87	4.41	4.09	4.25	4.12	4.97	4.75	4.77	4.76	4.83
3	Hilleshög HIL9707	Approved	3.52	3.99	4.70	4.35	4.07	4.60	4.53	4.96	4.75	4.70
7	Hilleshög 4302RR	NO	4.02	4.63	6.66	5.65	5.10	4.13	4.13	3.93	4.03	4.06
5	Hilleshög 9528RR	Approved	2.97	3.77	5.63	4.70	4.12	5.16	4.73	4.99	4.86	4.96
4	Maribo 109	Approved	3.54	4.27	5.06	4.67	4.29	4.56	4.14	4.14	4.14	4.28
3	Maribo MA502	Approved	2.93	3.06	3.53	3.30	3.17	5.04	4.79	5.01	4.90	4.95
5	SV RR333	Approved	3.46	4.71	4.99	4.85	4.39	4.54	4.85	4.84	4.85	4.74
3	SV RR351	Approved	3.53	4.38	4.18	4.28	4.03	4.62	4.50	4.41	4.46	4.51
3	SX Avalanche RR(858)	Approved	3.40	4.44	4.00	4.22	3.95	4.15	4.74	4.64	4.69	4.51
4	SX Canyon RR	Approved	3.59	4.28	4.33	4.31	4.07	4.02	4.76	4.92	4.84	4.57
4	SX Cruze RR	Approved	4.14	3.41	4.79	4.10	4.11	4.57	4.65	5.37	5.01	4.86
5	SX Winchester RR	Approved	3.07	3.85	4.36	4.11	3.76	3.67	3.97	4.07	4.02	3.90
Candidates for Approval			<=4.40					<=5.20				
5	BTS 8363	NO	4.77	4.93	4.60	4.77	4.77	3.83	4.33	4.10	4.22	4.09
2	BTS 8606	NO	--	4.60	4.91	4.76	--	--	5.12	4.73	4.93	--
2	BTS 8629	NO	--	4.14	4.68	4.41	--	--	4.59	4.29	4.44	--
6	Crystal 246RR	NO	4.99	4.85	5.13	4.99	4.99	4.49	4.81	4.63	4.72	4.64
6	Crystal 247RR	NO	4.94	4.77	5.35	5.06	5.02	4.19	4.65	4.55	4.60	4.46
3	Crystal 572RR	NO	4.33	4.74	4.69	4.72	4.59	4.65	4.57	4.27	4.42	4.50
3	Crystal 578RR	NO	4.52	4.44	4.56	4.50	4.51	4.93	4.87	4.91	4.89	4.90
2	Crystal 684RR	Approved	--	3.74	4.31	4.03	--	--	4.57	4.34	4.46	--
3	Hilleshög HIL9708	NO	4.69	4.82	5.94	5.38	5.15	5.04	4.74	4.61	4.68	4.80
2	Hilleshög HIL9895	Approved	--	3.65	4.39	4.02	--	--	4.49	4.84	4.67	--
6	Hilleshög 4448RR	NO	2.80	3.90	6.29	5.10	4.33	5.29	5.21	5.28	5.25	5.26
5	Maribo 305	NO	4.76	4.42	5.67	5.05	4.95	4.76	4.72	4.98	4.85	4.82
3	Maribo MA504	NO	4.60	4.54	6.20	5.37	5.11	5.25	5.04	5.50	5.27	5.26
2	Maribo MA611	Approved	--	3.94	4.00	3.97	--	--	4.47	5.03	4.75	--
4	SV RR244TT	NO	4.23	4.97	4.91	4.94	4.70	4.17	4.46	4.85	4.66	4.49
2	SV RR265	NO	--	4.54	5.35	4.95	--	--	5.00	5.19	5.10	--
2	SV RR266	NO	--	4.62	5.64	5.13	--	--	4.74	4.61	4.68	--
2	SV RR268	Approved	--	4.00	4.71	4.36	--	--	5.13	5.06	5.10	--
3	SX Marathon RR(856)	NO	4.53	4.38	4.52	4.45	4.48	5.37	4.44	4.54	4.49	4.78
2	SX RR1861	NO	--	4.40	5.71	5.06	--	--	4.52	4.74	4.63	--
2	SX RR1863	Approved	--	3.55	4.88	4.22	--	--	4.35	4.08	4.22	--
Approval Criteria new varieties			4.40					5.20				
Criteria to Maintain Approval			4.70					5.40				
+ All Cercospora ratings 2015-2017 were adjusted to 1982 basis. Aphanomyces approval criteria include: 1) Cercospora rating must not exceed 5.20 (1982 adjusted data), 2) Aph root rating <= 4.40 after 2 years. 3 yrs of data may be considered for initial approval. To maintain Aphanomyces approval criteria include: 1) Cercospora 3 year mean must not exceed 5.40, 2) Aph root rating <= 4.70 after 3 years. Previously approved varieties not meeting current approval standards may be sold in 2018.												
Created 11/9/2017												

Table 29. Calculation for Approval of Sugarbeet Varieties for ACSC Rhizoctonia Specialty Market for 2018

Variety	Approval Status	Disease Index +					Cercospora Rating				
		2015	2016	2017	2 Yr Mn	3 Yr Mn	2015	2016	2017	2 Yr Mn	3 Yr Mn
Previously Approved (3 Yr)											
Crystal 355RR	Approved	NE	3.96	4.09	4.03	NE	4.43	4.60	4.36	4.48	4.46
Hilleshög 4302RR	Approved	3.70	3.65	3.60	3.63	3.65	4.13	4.13	3.93	4.03	4.06
Maribo 109	Approved	3.67	3.69	3.63	3.66	3.66	4.56	4.14	4.14	4.14	4.28
Candidates for Approval (2 Yr)											
BTS 80RR52	Not Approved	3.95	4.41	4.14	4.28	4.17	4.11	4.28	4.37	4.33	4.25
BTS 8337	Not Approved	3.87	4.08	4.30	4.19	4.08	4.49	4.62	4.36	4.49	4.49
BTS 8363	Not Approved	4.12	4.34	4.85	4.60	4.44	3.83	4.33	4.10	4.22	4.09
BTS 8500	Not Approved	4.19	4.43	4.57	4.50	4.40	4.45	4.54	4.29	4.42	4.43
BTS 8512	Not Approved	4.28	4.44	4.28	4.36	4.33	4.12	4.04	3.69	3.87	3.95
BTS 8524	Not Approved	4.14	4.20	4.41	4.31	4.25	4.40	4.74	4.38	4.56	4.51
BTS 8572	Not Approved	3.85	4.54	4.32	4.43	4.24	4.60	4.41	4.14	4.28	4.38
BTS 8606	Not Approved	--	4.48	5.00	4.74	--	--	5.12	4.73	4.93	--
BTS 8629	Not Approved	--	3.73	4.21	3.97	--	--	4.59	4.29	4.44	--
Crystal 093RR	Not Approved	3.96	4.37	4.50	4.44	4.28	4.76	4.95	4.49	4.72	4.73
Crystal 101RR	Not Approved	4.64	4.78	4.78	4.78	4.73	4.65	4.59	4.57	4.58	4.60
Crystal 246RR	Not Approved	4.19	4.32	4.23	4.28	4.25	4.49	4.81	4.63	4.72	4.64
Crystal 247RR	Not Approved	4.33	4.32	4.49	4.41	4.38	4.19	4.65	4.55	4.60	4.46
Crystal 467RR	Not Approved	3.97	4.26	4.47	4.37	4.23	4.34	4.69	4.46	4.58	4.50
Crystal 572RR	Not Approved	3.89	4.21	4.47	4.34	4.19	4.65	4.57	4.27	4.42	4.50
Crystal 573RR	Not Approved	4.25	4.55	4.57	4.56	4.46	4.15	4.35	4.15	4.25	4.22
Crystal 574RR	Not Approved	4.16	4.47	4.16	4.32	4.26	4.30	4.51	4.35	4.43	4.39
Crystal 578RR	Not Approved	4.03	4.32	4.40	4.36	4.25	4.93	4.87	4.91	4.89	4.90
Crystal 684RR	Not Approved	--	4.41	4.57	4.49	--	--	4.57	4.34	4.46	--
Crystal 986RR	Not Approved	4.06	4.38	4.39	4.39	4.28	4.97	4.75	4.77	4.76	4.83
Hilleshög 4448RR	Not Approved	3.92	4.51	4.63	4.57	4.35	5.29	5.21	5.28	5.25	5.26
Hilleshög 9528RR	Not Approved	4.10	4.21	4.21	4.21	4.17	5.16	4.73	4.99	4.86	4.96
Hilleshög HIL9707	Not Approved	4.21	4.40	4.43	4.42	4.35	4.60	4.53	4.96	4.75	4.70
Hilleshög HIL9708	Not Approved	4.04	4.28	4.21	4.25	4.18	5.04	4.74	4.61	4.68	4.80
Hilleshög HIL9895	Not Approved	--	4.56	4.34	4.45	--	--	4.49	4.84	4.67	--
Maribo 305	Not Approved	3.83	4.40	4.60	4.50	4.28	4.76	4.72	4.98	4.85	4.82
Maribo MA502	Not Approved	4.14	4.73	4.78	4.76	4.55	5.04	4.79	5.01	4.90	4.95
Maribo MA504	Not Approved	3.98	4.58	4.37	4.48	4.31	5.25	5.04	5.50	5.27	5.26
Maribo MA611	Not Approved	--	4.63	4.44	4.54	--	--	4.47	5.03	4.75	--
SX Avalanche RR(858)	Not Approved	4.21	4.52	4.29	4.41	4.34	4.15	4.74	4.64	4.69	4.51
SX Canyon RR	Not Approved	4.22	4.40	4.51	4.46	4.38	4.02	4.76	4.92	4.84	4.57
SX Cruze RR	Not Approved	4.18	4.69	4.39	4.54	4.42	4.57	4.65	5.37	5.01	4.86
SX Marathon RR(856)	Not Approved	4.16	4.47	4.40	4.44	4.34	5.37	4.44	4.54	4.49	4.78
SX RR1861	Not Approved	--	4.59	4.50	4.55	--	--	4.52	4.74	4.63	--
SX RR1863	Not Approved	--	4.54	4.23	4.39	--	--	4.35	4.08	4.22	--
SX Winchester RR	Not Approved	4.28	4.63	4.47	4.55	4.46	3.67	3.97	4.07	4.02	3.90
SV RR244TT	Not Approved	4.18	4.45	4.50	4.48	4.38	4.17	4.46	4.85	4.66	4.49
SV RR265	Not Approved	--	4.44	4.42	4.43	--	--	5.00	5.19	5.10	--
SV RR266	Not Approved	--	4.20	4.39	4.30	--	--	4.74	4.61	4.68	--
SV RR268	Not Approved	--	4.70	4.57	4.64	--	--	5.13	5.06	5.10	--
SV RR333	Not Approved	4.11	4.44	4.44	4.44	4.33	4.54	4.85	4.84	4.85	4.74
SV RR351	Not Approved	--	4.17	4.25	4.21	--	4.62	4.50	4.41	4.46	4.51
Susceptible Checks											
RH CK#08 CRY539RR		4.65	4.84	4.74							
RH CK#21 CRY576RR		--	--	4.66							
RH CK#24 BETA86RR88		4.82	--	--							
RH CK#25 HILL4043RR		4.35	4.76	4.51							
RH CK#27 HILL4012RR		4.41	--	--							
RH CK#28 CRY565RR		--	4.57	--							
RH CK#29 BETA87RR58		4.77	4.67	4.79							
RH CK#30 SES36711RR		4.91	--	--							
RH CK#31 HILL4000RR		5.03	4.80	4.65							
RH CK#34 BETA86RR66		4.57	--	--							
RH CK#35 SES36812RR		4.37	4.55	4.71							
RH CK#36 BETA85RR02		4.71	--	--							
RH CK#37 SES36918RR		4.34	4.67	--							
RH CK#40 CRY5101RR		4.55	4.65	4.55							
RH CK#45 BTS82RR33		--	--	4.73							
RH CK#47 SES36272RR		--	4.50	4.62							
RH CK#49 CRY5247RR		--	4.38	4.65							
Susceptible Hybrid Mean		4.62	4.64	4.66	4.65	4.64				5.20	5.40
Approval Criteria ++		3.82	3.82	3.82	3.82	3.82					
Disapproval Criteria						4.18					
Rhc and CR ratings were adjusted based upon check performance. Created 11/8/2017											
+ Disease Index is based on a scale of 0 (healthy) to 7 (dead).											
++ Candidates must have better tolerance than susc. check mean * 80%. To maintain approval, tolerance must be better than susc. check mean * 90%.											
Previously approved varieties not meeting current approval standards may be sold in 2018.											

Table 30. 2017 Aphanomyces Ratings for Official Trial Entries
Betaseed Nursery - Shakopee, MN & ACSC - RRV

Chk @	Code	Variety	Adjusted @						Trial Yrs
			Shak 8/30	2017	2 Yr	3 Yr	2016^^	2015 ^^	
				1 loc	3loc	5 loc	2 loc	2 loc	
	529	BTS 80RR52	4.36	4.36	4.23	3.90	4.11	3.24	8
	545	BTS 8337	3.78	3.78	3.52	3.19	3.26	2.55	5
	562	BTS 8363	4.60	4.60	4.76	4.76	4.93	4.77	5
	513	BTS 8500	4.52	4.52	4.37	4.09	4.22	3.54	3
	533	BTS 8512	3.78	3.78	3.97	3.95	4.17	3.91	3
	550	BTS 8524	4.49	4.49	4.19	3.90	3.89	3.33	3
	570	BTS 8572	3.76	3.76	4.11	4.09	4.46	4.05	3
	509	BTS 8606	4.91	4.91	4.75	--	4.60	--	2
	525	BTS 8629	4.68	4.68	4.41	--	4.14	--	2
	577	BTS 8735	4.74	4.74	--	--	--	--	1
	506	BTS 8742	5.02	5.02	--	--	--	--	1
	536	BTS 8749	3.53	3.53	--	--	--	--	1
	540	BTS 8756	5.23	5.23	--	--	--	--	1
	521	BTS 8767	4.80	4.80	--	--	--	--	1
	518	BTS 8770	4.97	4.97	--	--	--	--	1
	567	BTS 8784	4.59	4.59	--	--	--	--	1
	502	BTS 8787	4.71	4.71	--	--	--	--	1
	512	BTS 8798	4.92	4.92	--	--	--	--	1
	549	Crystal 093RR	4.43	4.43	4.38	4.21	4.32	3.86	8
	551	Crystal 101RR	3.92	3.92	3.67	3.55	3.42	3.31	7
	507	Crystal 246RR	5.13	5.13	4.99	4.99	4.85	4.99	6
	560	Crystal 247RR	5.35	5.35	5.06	5.02	4.77	4.94	6
	565	Crystal 355RR	4.84	4.84	4.65	4.19	4.46	3.26	5
	523	Crystal 467RR	3.96	3.96	4.00	3.85	4.04	3.55	4
	503	Crystal 572RR	4.69	4.69	4.71	4.59	4.74	4.33	3
	554	Crystal 573RR	3.84	3.84	3.95	3.86	4.06	3.69	3
	544	Crystal 574RR	4.72	4.72	4.21	3.78	3.69	2.93	3
	571	Crystal 578RR	4.56	4.56	4.50	4.51	4.44	4.52	3
	510	Crystal 684RR	4.31	4.31	4.02	--	3.74	--	2
	547	Crystal 792RR	4.73	4.73	--	--	--	--	1
	557	Crystal 793RR	3.02	3.02	--	--	--	--	1
	534	Crystal 794RR	4.65	4.65	--	--	--	--	1
	522	Crystal 795RR	4.40	4.40	--	--	--	--	1
	553	Crystal 796RR	3.11	3.11	--	--	--	--	1
	528	Crystal 797RR	5.21	5.21	--	--	--	--	1
	532	Crystal 986RR	4.09	4.09	4.25	4.12	4.41	3.87	9
	559	Hilleshög HIL9707	4.70	4.70	4.34	4.07	3.99	3.52	3
	576	Hilleshög HIL9708	5.94	5.94	5.38	5.15	4.82	4.69	3
	561	Hilleshög HIL9895	4.39	4.39	4.02	--	3.65	--	2
	566	Hilleshög HIL9920	4.94	4.94	--	--	--	--	1
	563	Hilleshög HIL9921	5.41	5.41	--	--	--	--	1
	504	Hilleshög HIL9922	5.79	5.79	--	--	--	--	1
	543	Hilleshög HIL9923	5.06	5.06	--	--	--	--	1
	517	Hilleshög HIL9924	5.37	5.37	--	--	--	--	1
	505	Hilleshög 4302RR	6.66	6.66	5.65	5.10	4.63	4.02	7
	542	Hilleshög 4448RR	6.29	6.29	5.09	4.33	3.90	2.80	6
	531	Hilleshög 9528RR	5.63	5.63	4.70	4.12	3.77	2.97	5
	556	Maribo 109	5.06	5.06	4.66	4.29	4.27	3.54	4
	539	Maribo 305	5.67	5.67	5.05	4.95	4.42	4.76	5

	526	Maribo MA502	3.53	3.53	3.29	3.17	3.06	2.93	3
	514	Maribo MA504	6.20	6.20	5.37	5.11	4.54	4.60	3
	568	Maribo MA611	4.00	4.00	3.97	--	3.94	--	2
	574	Maribo MA717	5.31	5.31	--	--	--	--	1
	530	Maribo MA718	4.46	4.46	--	--	--	--	1
	538	Maribo MA719	4.75	4.75	--	--	--	--	1
	564	SV RR244TT	4.91	4.91	4.94	4.70	4.97	4.23	4
	511	SV RR265	5.35	5.35	4.95	--	4.54	--	2
	555	SV RR266	5.64	5.64	5.13	--	4.62	--	2
	572	SV RR268	4.71	4.71	4.36	--	4.00	--	2
	541	SV RR333	4.99	4.99	4.85	4.39	4.71	3.46	5
	573	SV RR351	4.18	4.18	4.28	4.03	4.38	3.53	3
	515	SV RR371	4.55	4.55	--	--	--	--	1
	501	SV RR372	4.42	4.42	--	--	--	--	1
	508	SV RR373	4.93	4.93	--	--	--	--	1
	578	SV RR374	5.20	5.20	--	--	--	--	1
	546	SV RR375	4.54	4.54	--	--	--	--	1
	537	SX Avalanche RR(858)	4.00	4.00	4.22	3.95	4.44	3.40	3
	548	SX Canyon RR	4.33	4.33	4.31	4.07	4.28	3.59	4
	535	SX Cruze RR	4.79	4.79	4.10	4.11	3.41	4.14	4
	519	SX Marathon RR(856)	4.52	4.52	4.45	4.48	4.38	4.53	3
	558	SX RR1861	5.71	5.71	5.05	--	4.40	--	2
	527	SX RR1863	4.88	4.88	4.21	--	3.55	--	2
	516	SX RR1875	4.13	4.13	--	--	--	--	1
	520	SX RR1876	4.73	4.73	--	--	--	--	1
	569	SX RR1877	3.84	3.84	--	--	--	--	1
	552	SX RR1878	5.54	5.54	--	--	--	--	1
	524	SX RR1879	4.18	4.18	--	--	--	--	1
	575	SX Winchester RR	4.36	4.36	4.11	3.76	3.85	3.07	5
1	1001	AP Ck-32 CRY5981RR	3.19	3.19	3.45	3.38	3.71	3.25	9
1	1002	AP CK-33 CRY5768RR	4.74	4.74	4.73	4.77	4.71	4.86	11
1	1003	AP CK-34 HILL4000RR	6.76	6.76	6.13	6.00	5.49	5.73	11
1	1004	AP CK-35 BETA87RR58	4.86	4.86	5.03	5.29	5.20	5.79	11
1	1005	AP CK-41 CRY5765RR	6.01	6.01	5.91	6.19	5.81	6.73	7
1	1006	AP CK-43 BTS80RR32	4.64	4.64	4.65	4.86	4.66	5.26	8
1	1007	AP CK-44 SX VISION RR	5.17	5.17	5.07	5.16	4.97	5.33	9
1	1008	AP CK-45 CRY5986RR	4.22	4.22	4.41	4.32	4.60	4.14	9
1	1009	AP CK-47 CRY5101RR	3.83	3.83	3.62	3.46	3.41	3.14	7
1	1010	AP CK-49 BTS82RR33	6.29	6.29	5.96	6.00	5.63	6.09	6
1	1011	AP CK-51 CRY5246RR	4.65	4.65	4.77	4.84	4.89	4.99	6
1	1012	AP CK-52 HILL4094RR	4.58	4.58	4.74	4.69	4.90	4.60	10
1	1013	AP CK-53 CRY5093RR	4.19	4.19	4.37	4.20	4.55	3.86	8
1	1014	AP CK-54 SES36273RR	5.05	5.05	4.76	4.63	4.46	4.38	6
1	1015	AP CK-55 CRY5247RR	4.00	4.00	4.59	4.71	5.19	4.94	6
	1016	AP CHK SUS HYB#3	4.99	4.99	5.34	5.90	5.70	7.03	11
	1017	AP CHK MOD RES RR	4.65	4.65	4.71	4.54	4.76	4.22	11
	1018	AP CHK RES RR	4.49	4.49	4.21	4.00	3.93	3.59	12
	1019	AP CHK SUS HYB#3	5.40	5.40	5.55	6.04	5.70	7.03	11
	1020	AP CHK SUS HYB#4	5.99	5.99	5.92	6.46	5.85	7.56	11

1021	AP CHK MOD RES RR#2	4.78	4.78	4.76	4.68	4.74	4.51	11
1022	AP CHK MOD RES RR#4	4.74	4.74	4.76	4.82	4.77	4.94	6
1023	AC CHK RES RR#3	3.23	3.23	3.13	2.88	3.02	2.38	10
1024	AP CHK SUS HYB#4	6.20	6.20	6.02	6.53	5.85	7.56	11
	Conventional							
919	BETA EXP 687	4.30	4.30	4.59	--	4.88	--	2
918	BETA EXP 698	3.62	3.62	3.65	--	3.69	--	2
905	BETA EXP 747	3.60	3.60	--	--	--	--	1
909	BETA EXP 758	3.29	3.29	--	--	--	--	1
901	Crystal 620	4.09	4.09	4.18	--	4.28	--	2
906	Crystal 622	4.05	4.05	4.20	--	4.36	--	2
913	Crystal 735	3.93	3.93	--	--	--	--	1
910	Crystal 737	2.25	2.25	--	--	--	--	1
902	Crystal R761	4.01	4.01	3.79	--	3.57	--	11
914	Hilleshög 3035Rz	5.18	5.18	4.79	--	4.40	--	13
917	Hilleshög 9891Rz	4.89	4.89	4.67	--	4.45	--	2
904	Maribo MA615Rz	5.30	5.30	5.05	--	4.80	--	2
916	Maribo MA720Rz	5.15	5.15	--	--	--	--	1
911	Seedex 8869	4.99	4.99	4.85	--	4.70	--	2
907	Seedex Deuce	6.04	6.04	5.87	--	5.70	--	10
912	Strube 12720	8.11	8.11	--	--	--	--	1
908	Strube 13722	7.54	7.54	--	--	--	--	1
903	SV 48611	4.25	4.25	4.36	--	4.47	--	2
915	SV 48777	4.20	4.20	--	--	--	--	1
1001	AP Ck-32 CRY5981RR	2.93	2.93	3.32	3.30	3.71	3.25	9
1003	AP CK-34 HILL4000RR	6.36	6.36	5.92	5.86	5.49	5.73	11
1006	AP CK-43 BTS80RR32	5.57	5.57	5.11	5.16	4.66	5.26	8
1009	AP CK-47 CRY5101RR	3.01	3.01	3.21	3.19	3.41	3.14	7
1011	AP CK-51 CRY5246RR	5.20	5.20	5.05	5.03	4.89	4.99	6
	Check Mean	4.81	4.81					
15	Trial Mean	4.75	4.75					
	Coeff. of Var. (%)	24.9						
	Mean LSD (0.05)	1.44						
	Mean LSD (0.01)	1.90						
	Sig Lvl	**						
	Adjustment Factor	1.11						
	@ 2017 Root Rating was taken in early fall (1=healthy, 9=severe damage).						Created 11/3/2017	
	Ratings adjusted to 2003 basis. (2000-2002 Aph nurseries). Adjustment based on variety checks.							

Table 31. 2017 Cercospora Ratings for ACSC Official Trial Entries
Betaseed (Randolph MN), BSDF (Frankenmuth MI) & NDSU (Foxhome MN)

Chk @	Code	Variety	Adjusted to 1982 Basis @								Trial Yrs
			Beta	BSDF	Foxhome						
			Avg	Avg	Avg	2017	2 Yr	3 Yr	2016	2015	
			5 Dates+	5 Dates+	8 Dates+	3 loc	6 loc	9 loc	3 loc	3 loc	
	529	BTS 80RR52	3.59	5.40	4.13	4.37	4.33	4.26	4.28	4.11	8
	545	BTS 8337	4.25	4.37	4.46	4.36	4.49	4.49	4.62	4.49	5
	562	BTS 8363	3.96	4.48	3.87	4.10	4.21	4.09	4.33	3.83	5
	513	BTS 8500	4.44	4.52	3.90	4.29	4.41	4.43	4.54	4.45	3
	533	BTS 8512	2.99	4.29	3.79	3.69	3.86	3.95	4.04	4.12	3
	550	BTS 8524	4.61	4.55	3.98	4.38	4.56	4.51	4.74	4.40	3
	570	BTS 8572	3.51	4.46	4.45	4.14	4.27	4.38	4.41	4.60	3
	509	BTS 8606	4.76	4.81	4.62	4.73	4.92	--	5.12	--	2
	525	BTS 8629	4.18	4.46	4.22	4.29	4.44	--	4.59	--	2
	577	BTS 8735	3.92	4.77	3.97	4.22	--	--	--	--	1
	506	BTS 8742	3.73	4.65	4.71	4.36	--	--	--	--	1
	536	BTS 8749	3.42	4.65	4.08	4.05	--	--	--	--	1
	540	BTS 8756	3.27	4.65	4.12	4.01	--	--	--	--	1
	521	BTS 8767	4.07	4.30	4.10	4.16	--	--	--	--	1
	518	BTS 8770	4.18	4.96	3.77	4.30	--	--	--	--	1
	567	BTS 8784	2.93	4.23	3.81	3.65	--	--	--	--	1
	502	BTS 8787	3.66	4.60	3.84	4.03	--	--	--	--	1
	512	BTS 8798	3.92	4.58	4.42	4.30	--	--	--	--	1
	549	Crystal 093RR	4.05	4.60	4.81	4.49	4.72	4.73	4.95	4.76	8
	551	Crystal 101RR	4.84	4.41	4.47	4.57	4.58	4.60	4.59	4.65	7
	507	Crystal 246RR	4.90	4.69	4.30	4.63	4.72	4.64	4.81	4.49	6
	560	Crystal 247RR	4.95	4.41	4.30	4.55	4.60	4.47	4.65	4.19	6
	565	Crystal 355RR	4.06	4.65	4.38	4.36	4.48	4.46	4.60	4.43	5
	523	Crystal 467RR	4.49	4.61	4.27	4.46	4.57	4.49	4.69	4.34	4
	503	Crystal 572RR	4.01	4.30	4.51	4.27	4.42	4.50	4.57	4.65	3
	554	Crystal 573RR	3.84	4.18	4.42	4.15	4.25	4.22	4.35	4.15	3
	544	Crystal 574RR	4.56	4.54	3.96	4.35	4.43	4.39	4.51	4.30	3
	571	Crystal 578RR	5.46	4.80	4.47	4.91	4.89	4.91	4.87	4.93	3
	510	Crystal 684RR	4.26	4.65	4.10	4.34	4.45	--	4.57	--	2
	547	Crystal 792RR	3.04	4.55	4.22	3.94	--	--	--	--	1
	557	Crystal 793RR	3.28	4.31	4.20	3.93	--	--	--	--	1
	534	Crystal 794RR	5.30	5.04	4.42	4.92	--	--	--	--	1
	522	Crystal 795RR	3.92	4.88	4.38	4.39	--	--	--	--	1
	553	Crystal 796RR	4.84	4.94	4.78	4.85	--	--	--	--	1
	528	Crystal 797RR	3.73	4.49	4.29	4.17	--	--	--	--	1
	532	Crystal 986RR	4.25	4.89	5.16	4.77	4.76	4.83	4.75	4.97	9
	559	Hilleshög HIL9707	4.76	5.19	4.92	4.96	4.74	4.70	4.53	4.60	3
	576	Hilleshög HIL9708	4.71	4.59	4.55	4.61	4.68	4.80	4.74	5.04	3
	561	Hilleshög HIL9895	4.77	4.88	4.87	4.84	4.67	--	4.49	--	2
	566	Hilleshög HIL9920	4.31	5.04	5.33	4.89	--	--	--	--	1
	563	Hilleshög HIL9921	4.31	4.75	4.34	4.47	--	--	--	--	1
	504	Hilleshög HIL9922	3.76	4.54	3.77	4.02	--	--	--	--	1
	543	Hilleshög HIL9923	5.09	5.26	4.08	4.81	--	--	--	--	1
	517	Hilleshög HIL9924	3.78	4.81	3.68	4.09	--	--	--	--	1
	505	Hilleshög 4302RR	3.63	4.11	4.04	3.93	4.03	4.06	4.13	4.13	7
	542	Hilleshög 4448RR	5.46	4.83	5.54	5.28	5.24	5.26	5.21	5.29	6
	531	Hilleshög 9528RR	5.13	5.01	4.83	4.99	4.86	4.96	4.73	5.16	5
	556	Maribo 109	3.96	4.49	3.96	4.14	4.14	4.28	4.14	4.56	4
	539	Maribo 305	4.77	5.39	4.77	4.98	4.85	4.82	4.72	4.76	5

	526	Maribo MA502	4.98	5.30	4.76		5.01	4.90	4.95	4.79	5.04	3
	514	Maribo MA504	5.07	5.76	5.66		5.50	5.27	5.26	5.04	5.25	3
	568	Maribo MA611	4.95	5.45	4.69		5.03	4.75	--	4.47	--	2
	574	Maribo MA717	4.65	4.84	5.05		4.85	--	--	--	--	1
	530	Maribo MA718	4.32	4.53	4.30		4.39	--	--	--	--	1
	538	Maribo MA719	4.13	5.14	3.96		4.41	--	--	--	--	1
	564	SVRR244TT	5.10	4.48	4.95		4.85	4.65	4.49	4.46	4.17	4
	511	SVRR265	5.26	5.07	5.23		5.19	5.09	--	5.00	--	2
	555	SVRR266	4.35	5.01	4.48		4.61	4.67	--	4.74	--	2
	572	SVRR268	5.27	4.80	5.13		5.06	5.10	--	5.13	--	2
	541	SVRR333	4.65	5.07	4.79		4.84	4.84	4.74	4.85	4.54	5
	573	SVRR351	4.16	4.63	4.44		4.41	4.46	4.51	4.50	4.62	3
	515	SVRR371	3.73	5.25	4.79		4.59	--	--	--	--	1
	501	SVRR372	3.94	4.41	4.32		4.23	--	--	--	--	1
	508	SVRR373	3.83	4.65	4.45		4.31	--	--	--	--	1
	544	SVRR374	4.59	4.89	4.65		4.71	--	--	--	--	1
	546	SVRR375	5.10	5.21	4.91		5.08	--	--	--	--	1
	537	SX Avalanche RR(858)	4.58	4.54	4.79		4.64	4.69	4.51	4.74	4.15	3
	548	SX Canyon RR	4.22	5.84	4.69		4.92	4.84	4.56	4.76	4.02	4
	535	SX Cruze RR	5.82	5.41	4.87		5.37	5.01	4.87	4.65	4.57	4
	519	SX Marathon RR(856)	4.05	4.90	4.67		4.54	4.49	4.78	4.44	5.37	3
	558	SX RR1861	4.40	4.88	4.95		4.74	4.63	--	4.52	--	2
	527	SX RR1863	3.52	4.25	4.47		4.08	4.21	--	4.35	--	2
	516	SX RR1875	3.19	4.96	4.04		4.06	--	--	--	--	1
	520	SX RR1876	4.36	4.41	4.15		4.31	--	--	--	--	1
	569	SX RR1877	4.56	5.21	4.08		4.62	--	--	--	--	1
	552	SX RR1878	4.54	4.97	4.61		4.71	--	--	--	--	1
	524	SX RR1879	4.53	5.23	4.87		4.88	--	--	--	--	1
	575	SX Winchester RR	3.42	4.75	4.03		4.07	4.02	3.90	3.97	3.67	5
1	1101	CR CK-19 CRY5539RR	5.98	4.89	5.59		5.49	5.39	5.37	5.30	5.31	13
1	1102	CR CK-24 HILL4012RR	5.06	4.87	5.47		5.13	5.22	5.23	5.31	5.24	12
1	1103	CR CK-28 HILL4010RR	5.24	6.15	4.94		5.44	5.44	5.36	5.43	5.20	12
1	1104	CR CK-33 HILL4043RR	5.33	5.18	5.13		5.21	4.97	5.01	4.73	5.09	11
1	1105	CR CK-34 HILL4000RR	4.92	5.12	4.95		5.00	4.88	4.80	4.77	4.64	11
1	1106	CR CK-41 CRY5981RR	5.30	4.56	4.84		4.90	4.89	4.97	4.89	5.12	9
1	1107	CR CK-42 CRY5985RR	3.24	4.44	4.06		3.91	4.07	4.20	4.23	4.45	9
1	1108	CR CK-43 CRY5246RR	4.70	4.95	4.67		4.77	4.77	4.68	4.77	4.49	6
1	1109	CR CK-44 BET A80RR32	5.51	4.42	4.88		4.94	4.99	4.97	5.04	4.92	8
1	1110	CR CK-45 HILL4448RR	5.03	5.34	5.34		5.24	5.12	5.18	5.00	5.29	6
1	1111	CR CK-46 HILL4062RR	3.90	4.25	4.18		4.11	4.24	4.29	4.37	4.39	10
1	1112	CR CK-47 HILL4094RR	4.25	4.29	4.40		4.31	4.30	4.30	4.28	4.30	10
	1113	CR CK MOD SUS HYB#3	5.64	4.80	5.53		5.32	5.33	5.24	5.33	5.05	13
	1114	CR CK MOD SUS HYB#3	5.58	5.26	5.61		5.49	5.41	5.29	5.33	5.05	13
	1115	CR CK MOD RES HYB#4	3.23	4.95	4.71		4.30	4.27	4.35	4.24	4.52	10
	1116	CR CK MOD RES HYB#4	3.35	4.54	4.51		4.13	4.19	4.30	4.24	4.52	10
	1117	CR CK MOD SUS HYB#5	4.76	5.13	5.45		5.11	5.04	5.10	4.97	5.21	11
		Conventional										
	919	BETA EXP 687	3.58	4.05	4.35		3.99	4.07	--	4.14	--	2
	918	BETA EXP 698	4.52	3.87	4.14		4.18	4.23	--	4.27	--	2

905	BETA EXP 747	3.96	4.36	4.88	4.40	--	--	--	--	1
909	BETA EXP 758	4.39	4.44	4.74	4.52	--	--	--	--	1
901	Crystal 620	4.20	3.87	4.34	4.14	4.17	--	4.19	--	2
906	Crystal 622	2.88	3.93	4.34	3.72	3.84	--	3.96	--	2
913	Crystal 735	4.52	4.04	4.76	4.44	--	--	--	--	1
910	Crystal 737	3.46	3.77	4.55	3.92	--	--	--	--	1
902	Crystal R761	5.23	4.40	5.16	4.93	4.96	--	4.99	--	11
914	Hilleshog 3035Rz	4.12	4.23	4.93	4.42	4.47	--	4.53	--	13
917	Hilleshog 9891Rz	3.76	3.90	4.73	4.13	4.27	--	4.42	--	2
904	Maribo MA615Rz	4.92	4.71	4.80	4.81	4.92	--	5.04	--	2
916	Maribo MA720Rz	5.03	3.80	4.80	4.54	--	--	--	--	1
911	Seedex 8869	5.63	4.33	5.67	5.21	4.99	--	4.76	--	2
907	Seedex Deuce	4.60	4.48	5.20	4.76	4.72	--	4.68	--	10
912	Srube 12720	5.58	5.11	6.26	5.65	--	--	--	--	1
908	Srube 13722	3.79	3.57	4.80	4.06	--	--	--	--	1
903	SV 48611	5.95	4.30	5.59	5.28	5.06	--	4.85	--	2
915	SV 48777	4.32	4.43	5.54	4.76	--	--	--	--	1
1101	CR CK-19 CRY5539RR	5.98	5.32	5.53	5.61	5.45	5.41	5.30	5.31	13
1106	CR CK-41 CRY5981RR	5.30	4.72	5.39	5.14	5.01	5.05	4.89	5.12	9
1107	CR CK-42 CRY5985RR	3.24	3.98	5.11	4.11	4.17	4.27	4.23	4.45	9
1109	CR CK-44 BET A80RR32	5.51	4.26	4.92	4.90	4.97	4.95	5.04	4.92	8
1110	CR CK-45 HILL4448RR	5.03	5.38	5.39	5.27	5.13	5.19	5.00	5.29	6
	Check Mean	4.77	4.89	4.96	4.87					
12	Trial Mean	4.38	4.80	4.54	4.57					
	Coeff. of Var. (%)	10.15	9.25	5.97						
	Mean LSD (0.05)	0.55	0.59	0.32						
	Mean LSD (0.01)	0.73	0.79	0.43						
	Sig Mrk	**	**	**						
	Adj Factor	1.20	1.16	1.01						
	Lower numbers indicate better Cercospora resistance (1-Ex, 9=Poor).									
	@ Ratings adjusted to 1982 basis (5.5 equivalent in 1978-81 CR nurseries). Adjustment based on check varieties.									
	Chk = varieties used to adjust CR readings to 1982 basis. Ratings * (Adj. factor) = Adj Rating.									
	+ Average rating based upon multiple rating dates.									
										Created 11/3/2017

Table 32. 2017 Rhizoctonia Ratings for ACSC Official Trial Entries
Rhizoctonia Nursery - BSDF, NWROC & Two ACSC Sites

Sus Chk Chk ^ @	Code	Variety	Adjusted @							Trial Yrs
			BSDF		2 Yr	3 Yr	2016	2015		
			8/24	2017						
				1 loc	5 loc	9 loc	4 loc	4 loc		
	529	BTS 80RR52	4.14	4.14	4.27	4.17	4.41	3.95	8	
	545	BTS 8337	4.30	4.30	4.19	4.08	4.08	3.87	5	
	562	BTS 8363	4.85	4.85	4.59	4.44	4.34	4.12	5	
	513	BTS 8500	4.57	4.57	4.50	4.40	4.43	4.19	3	
	533	BTS 8512	4.28	4.28	4.36	4.33	4.44	4.28	3	
	550	BTS 8524	4.41	4.41	4.31	4.25	4.20	4.14	3	
	570	BTS 8572	4.32	4.32	4.43	4.24	4.54	3.85	3	
	509	BTS 8606	5.00	5.00	4.74	--	4.48	--	2	
	525	BTS 8629	4.21	4.21	3.97	--	3.73	--	2	
	577	BTS 8735	4.38	4.38	--	--	--	--	1	
	506	BTS 8742	4.23	4.23	--	--	--	--	1	
	536	BTS 8749	3.95	3.95	--	--	--	--	1	
	540	BTS 8756	4.34	4.34	--	--	--	--	1	
	521	BTS 8767	4.75	4.75	--	--	--	--	1	
	518	BTS 8770	4.57	4.57	--	--	--	--	1	
	567	BTS 8784	4.64	4.64	--	--	--	--	1	
	502	BTS 8787	4.31	4.31	--	--	--	--	1	
	512	BTS 8798	4.52	4.52	--	--	--	--	1	
	549	Crystal 093RR	4.50	4.50	4.44	4.28	4.37	3.96	8	
	551	Crystal 101RR	4.78	4.78	4.78	4.73	4.78	4.64	7	
	507	Crystal 246RR	4.23	4.23	4.28	4.25	4.32	4.19	6	
	560	Crystal 247RR	4.49	4.49	4.40	4.38	4.32	4.33	6	
	565	Crystal 355RR	4.09	4.09	4.02	--	3.96	--	5	
	523	Crystal 467RR	4.47	4.47	4.37	4.23	4.26	3.97	4	
	503	Crystal 572RR	4.47	4.47	4.34	4.19	4.21	3.89	3	
	554	Crystal 573RR	4.57	4.57	4.56	4.45	4.55	4.25	3	
	544	Crystal 574RR	4.16	4.16	4.31	4.26	4.47	4.16	3	
	571	Crystal 578RR	4.40	4.40	4.36	4.25	4.32	4.03	3	
	510	Crystal 684RR	4.57	4.57	4.49	--	4.41	--	2	
	547	Crystal 792RR	3.88	3.88	--	--	--	--	1	
	557	Crystal 793RR	4.26	4.26	--	--	--	--	1	
	534	Crystal 794RR	4.15	4.15	--	--	--	--	1	
	522	Crystal 795RR	3.94	3.94	--	--	--	--	1	
	553	Crystal 796RR	4.23	4.23	--	--	--	--	1	
	528	Crystal 797RR	4.26	4.26	--	--	--	--	1	
	532	Crystal 986RR	4.39	4.39	4.38	4.28	4.38	4.06	9	
	559	Hilleshög HIL9707	4.43	4.43	4.41	4.35	4.40	4.21	3	
	576	Hilleshög HIL9708	4.21	4.21	4.25	4.18	4.28	4.04	3	
	561	Hilleshög HIL9895	4.34	4.34	4.45	--	4.56	--	2	
	566	Hilleshög HIL9920	4.48	4.48	--	--	--	--	1	
	563	Hilleshög HIL9921	3.85	3.85	--	--	--	--	1	
	504	Hilleshög HIL9922	4.39	4.39	--	--	--	--	1	
	543	Hilleshög HIL9923	4.58	4.58	--	--	--	--	1	
	517	Hilleshög HIL9924	4.62	4.62	--	--	--	--	1	
	505	Hilleshög 4302RR	3.60	3.60	3.63	3.65	3.65	3.70	7	
	542	Hilleshög 4448RR	4.63	4.63	4.57	4.35	4.51	3.92	6	
	531	Hilleshög 9528RR	4.21	4.21	4.21	4.18	4.21	4.10	5	
	556	Maribo 109	3.63	3.63	3.66	3.66	3.69	3.67	4	
	539	Maribo 305	4.60	4.60	4.50	4.28	4.40	3.83	5	
	526	Maribo MA502	4.78	4.78	4.76	4.55	4.73	4.14	3	

		514	Maribo MA504	4.37	4.37	4.47	4.31	4.58	3.98	3
		568	Maribo MA611	4.44	4.44	4.53	--	4.63	--	2
		574	Maribo MA717	4.28	4.28	--	--	--	--	1
		530	Maribo MA718	4.13	4.13	--	--	--	--	1
		538	Maribo MA719	4.28	4.28	--	--	--	--	1
		564	SV RR244TT	4.50	4.50	4.48	4.38	4.45	4.18	4
		511	SV RR265	4.42	4.42	4.43	--	4.44	--	2
		555	SV RR266	4.39	4.39	4.30	--	4.20	--	2
		572	SV RR268	4.57	4.57	4.63	--	4.70	--	2
		541	SV RR333	4.44	4.44	4.44	4.33	4.44	4.11	5
		573	SV RR351	4.25	4.25	4.21	--	4.17	--	3
		515	SV RR371	4.31	4.31	--	--	--	--	1
		501	SV RR372	4.47	4.47	--	--	--	--	1
		508	SV RR373	4.38	4.38	--	--	--	--	1
		578	SV RR374	4.30	4.30	--	--	--	--	1
		546	SV RR375	4.25	4.25	--	--	--	--	1
		537	SX Avalanche RR(858)	4.29	4.29	4.40	4.34	4.52	4.21	3
		548	SX Canyon RR	4.51	4.51	4.45	4.38	4.40	4.22	4
		535	SX Cruze RR	4.39	4.39	4.54	4.42	4.69	4.18	4
		519	SX Marathon RR(856)	4.40	4.40	4.43	4.34	4.47	4.16	3
		558	SX RR1861	4.50	4.50	4.55	--	4.59	--	2
		527	SX RR1863	4.23	4.23	4.39	--	4.54	--	2
		516	SX RR1875	4.34	4.34	--	--	--	--	1
		520	SX RR1876	4.42	4.42	--	--	--	--	1
		569	SX RR1877	4.42	4.42	--	--	--	--	1
		552	SX RR1878	4.31	4.31	--	--	--	--	1
		524	SX RR1879	4.36	4.36	--	--	--	--	1
		575	SX Winchester RR	4.47	4.47	4.55	4.46	4.63	4.28	5
1	1	1301	RH CK#08 CRY539RR	4.74	4.74	4.79	4.74	4.84	4.65	9
	1	1302	RH CK#20 CRY5765RR	4.31	4.31	4.33	4.29	4.35	4.22	9
	1	1303	RH CK#21 CRY5768RR	4.66	4.66	4.49	4.41	4.32	4.25	9
1	1	1304	RH CK#25 HILL4043RR	4.51	4.51	4.63	4.54	4.76	4.35	9
1	1	1305	RH CK#28 CRY5658RR	4.36	4.36	4.46	4.34	4.57	4.09	12
1	1	1306	RH CK#29 BETA87RR58	4.79	4.79	4.73	4.75	4.67	4.77	11
1	1	1307	RH CK#31 HILL4000RR	4.65	4.65	4.72	4.83	4.80	5.03	11
1	1	1308	RH CK#35 SES36812RR	4.71	4.71	4.63	4.54	4.55	4.37	10
	1	1309	RH CK#36 BETA85RR02	4.10	4.10	4.28	4.42	4.45	4.71	13
1	1	1310	RH CK#37 SES36918RR	4.43	4.43	4.55	4.48	4.67	4.34	9
1	1	1311	RH CK#40 CRY5101RR	4.55	4.55	4.60	4.58	4.65	4.55	7
	1	1312	RH CK#45 BTS82RR33	4.73	4.73	4.46	4.37	4.19	4.18	6
1	1	1313	RH CK#47 SES36272RR	4.62	4.62	4.56	4.50	4.50	4.39	6
	1	1314	RH CK#48 HILL4094RR	3.80	3.80	3.85	3.71	3.90	3.44	10
1	1	1315	RH CK#49 CRY5247RR	4.65	4.65	4.51	4.45	4.38	4.33	6
		1316	RES RHC #1	3.62	3.62	3.73	3.64	3.83	3.47	12
		1317	MOD RHC #6	4.68	4.68	4.50	4.36	4.32	4.09	12
		1318	SUS RHC #3	4.32	4.32	4.51	4.57	4.70	4.69	13
		1319	SUS RHC #9	4.43	4.43	4.54	4.47	4.65	4.34	9
		1320	MOD RHC #5	4.34	4.34	4.53	4.44	4.71	4.27	12
		1321	RES RHC #2	3.65	3.65	3.83	3.78	4.01	3.68	10
		1322	SUS RHC #3	4.95	4.95	4.85	4.79	4.74	4.69	13

		1323	SUS RHC #9	4.51	4.51	4.54	4.47	4.57	4.34	9
		1324	SUS RHC #10	4.28	4.28	4.51	4.60	4.75	4.77	9
			Conventional							
		919	BETA EXP 687	4.20	4.20	4.18	--	4.16	--	2
		918	BETA EXP 698	4.45	4.45	4.40	--	4.35	--	2
		905	BETA EXP 747	3.93	3.93	--	--	--	--	1
		909	BETA EXP 758	4.31	4.31	--	--	--	--	1
		901	Crystal 620	4.37	4.37	4.45	--	4.54	--	2
		906	Crystal 622	4.49	4.49	4.31	--	4.14	--	2
		913	Crystal 735	4.61	4.61	--	--	--	--	1
		910	Crystal 737	4.25	4.25	--	--	--	--	1
		902	Crystal R761	4.54	4.54	4.55	--	4.57	--	11
		914	Hilleshög 3035Rz	4.07	4.07	4.00	--	3.93	--	13
		917	Hilleshög 9891Rz	4.46	4.46	4.34	--	4.22	--	2
		904	Maribo MA615Rz	4.73	4.73	4.63	--	4.54	--	2
		916	Maribo MA720Rz	4.55	4.55	--	--	--	--	1
		911	Seedex 8869	4.40	4.40	4.53	--	4.67	--	2
		907	Seedex Deuce	4.39	4.39	4.52	--	4.66	--	10
		912	Strube 12720	4.59	4.59	--	--	--	--	1
		908	Strube 13722	4.73	4.73	--	--	--	--	1
		903	SV 48611	4.35	4.35	4.50	--	4.66	--	2
		915	SV 48777	4.59	4.59	--	--	--	--	1
		1301	RH CK#08 CRY539RR	4.74	4.74	4.79	4.74	4.84	4.65	9
		1303	RH CK#21 CRY5768RR	4.66	4.66	4.49	4.41	4.32	4.25	9
		1311	RH CK#40 CRY5101RR	4.55	4.55	4.60	4.58	4.65	4.55	7
		1314	RH CK#48 HILL4094RR	3.80	3.80	3.85	3.71	3.90	3.44	10
		1315	RH CK#49 CRY5247RR	4.65	4.65	4.51	4.45	4.38	4.33	6
	15		Mean of Check Varieties	4.51	4.51	4.51	4.46	4.51	4.38	
	10		Mean of Susc Checks	4.60	4.60	4.62	4.58	4.64	4.49	
			Trial Mean	4.38						
			Coeff. of Var. (%)	7.0						
			Mean LSD (0.05)	0.43						
			Mean LSD (0.01)	0.56						
			Sig Lvl	**						
			Adjustment Factor	0.72						
			Approval Limit (80% of susc checks)	5.08	3.68	3.70	3.66	3.71	3.59	
			@ Adjustment is based upon check varieties.							
			Lower numbers indicate better tolerance (0=Ex, 7=Poor).							
			^ Approval criteria is based upon mean of 10 susc varieties (approval option 1) or 3.82 (approval option 2).							
										Created 11/3/2017

Table 33. 2017 Fusarium Ratings for ACSC Official Trial Entries
Two Moorhead, MN Sites

Chk @	Code	Variety	N Mhd		S Mhd		Adjusted @					Trial Yrs
			4 Dates+	4 Dates+	2017	2 Yr	3 Yr	2016	2015			
					2 loc	4 loc	6 loc	2 loc	2 loc			
	529	BTS 80RR52	2.61	2.77	2.69	2.75	2.77	2.81	2.83	8		
	545	BTS 8337	3.76	3.90	3.83	3.92	3.85	4.01	3.72	5		
	562	BTS 8363	3.45	3.54	3.49	3.30	3.15	3.11	2.85	5		
	513	BTS 8500	1.79	2.48	2.14	2.02	2.15	1.90	2.41	3		
	533	BTS 8512	2.89	3.02	2.96	2.83	2.79	2.71	2.70	3		
	550	BTS 8524	3.21	3.28	3.24	3.31	3.17	3.38	2.88	3		
	570	BTS 8572	2.07	3.02	2.54	2.39	2.44	2.23	2.54	3		
	509	BTS 8606	2.49	3.14	2.81	2.75	--	2.69	--	2		
	525	BTS 8629	4.15	4.26	4.20	4.12	--	4.04	--	2		
	577	BTS 8735	4.00	3.86	3.93	--	--	--	--	1		
	506	BTS 8742	2.21	2.98	2.59	--	--	--	--	1		
	536	BTS 8749	2.95	3.61	3.28	--	--	--	--	1		
	540	BTS 8756	2.36	2.99	2.67	--	--	--	--	1		
	521	BTS 8767	2.65	2.78	2.71	--	--	--	--	1		
	518	BTS 8770	2.39	3.24	2.82	--	--	--	--	1		
	567	BTS 8784	2.21	3.05	2.63	--	--	--	--	1		
	502	BTS 8787	1.98	3.02	2.50	--	--	--	--	1		
	512	BTS 8798	3.17	3.56	3.37	--	--	--	--	1		
	549	Crystal 093RR	3.22	3.74	3.48	3.42	3.35	3.35	3.22	8		
	551	Crystal 101RR	2.14	3.31	2.72	2.56	2.59	2.40	2.64	7		
	507	Crystal 246RR	3.10	3.38	3.24	3.17	3.11	3.10	3.00	6		
	560	Crystal 247RR	2.97	3.02	3.00	2.90	2.77	2.80	2.51	6		
	565	Crystal 355RR	2.58	2.94	2.76	2.71	NE	2.65	NE	5		
	523	Crystal 467RR	1.75	2.21	1.98	1.91	2.09	1.84	2.46	4		
	503	Crystal 572RR	2.33	2.95	2.64	2.23	2.27	1.82	2.36	3		
	554	Crystal 573RR	3.05	3.16	3.10	3.29	3.20	3.49	3.02	3		
	544	Crystal 574RR	1.87	2.59	2.23	2.02	2.02	1.82	2.00	3		
	571	Crystal 578RR	2.15	2.66	2.41	2.20	2.27	1.99	2.42	3		
	510	Crystal 684RR	1.73	2.30	2.01	1.89	--	1.76	--	2		
	547	Crystal 792RR	2.70	2.93	2.81	--	--	--	--	1		
	557	Crystal 793RR	2.72	3.18	2.95	--	--	--	--	1		
	534	Crystal 794RR	2.09	2.80	2.45	--	--	--	--	1		
	522	Crystal 795RR	2.39	2.93	2.66	--	--	--	--	1		
	553	Crystal 796RR	2.06	2.62	2.34	--	--	--	--	1		
	528	Crystal 797RR	3.12	3.24	3.18	--	--	--	--	1		
	532	Crystal 986RR	4.73	4.73	4.73	4.79	4.49	4.86	3.89	9		
	559	Hilleshög HIL9707	4.13	4.06	4.09	4.49	4.22	4.88	3.68	3		
	576	Hilleshög HIL9708	4.82	4.40	4.61	4.45	4.20	4.29	3.69	3		
	561	Hilleshög HIL9895	3.93	4.36	4.15	3.27	--	2.40	--	2		
	566	Hilleshög HIL9920	6.01	5.84	5.92	--	--	--	--	1		
	563	Hilleshög HIL9921	4.72	4.60	4.66	--	--	--	--	1		
	504	Hilleshög HIL9922	4.58	4.40	4.49	--	--	--	--	1		
	543	Hilleshög HIL9923	4.91	5.67	5.29	--	--	--	--	1		
	517	Hilleshög HIL9924	4.54	4.62	4.58	--	--	--	--	1		
	505	Hilleshög 4302RR	4.99	5.19	5.09	5.09	4.74	5.09	4.05	7		
	542	Hilleshög 4448RR	5.75	4.94	5.35	5.30	NE	5.26	NE	6		
	531	Hilleshög 9528RR	4.52	3.97	4.25	4.39	4.26	4.52	4.00	5		
	556	Maribo 109	4.45	4.02	4.23	4.37	4.11	4.50	3.58	4		
	539	Maribo 305	5.91	5.86	5.89	5.89	5.60	5.89	5.02	5		
	526	Maribo MA502	2.70	3.34	3.02	2.47	2.42	1.92	2.33	3		
	514	Maribo MA504	4.62	4.43	4.52	4.56	4.41	4.60	4.11	3		
	568	Maribo MA611	3.58	3.97	3.78	2.87	--	1.96	--	2		
	574	Maribo MA717	5.10	4.80	4.95	--	--	--	--	1		

	530	Maribo MA718	4.26	4.96	4.61	--	--	--	--	1
	538	Maribo MA719	6.36	5.16	5.76	--	--	--	--	1
	564	SV RR244TT	3.85	3.62	3.74	3.94	3.91	4.14	3.86	4
	511	SV RR265	5.19	5.46	5.32	5.29	--	5.26	--	2
	555	SV RR266	6.14	5.14	5.64	5.41	--	5.18	--	2
	572	SV RR268	5.37	4.65	5.01	5.11	--	5.20	--	2
	541	SV RR333	5.66	5.03	5.35	5.09	NE	4.84	NE	5
	573	SV RR351	5.07	4.86	4.96	4.86	NE	4.75	NE	3
	515	SV RR371	4.92	4.90	4.91	--	--	--	--	1
	501	SV RR372	4.33	4.06	4.19	--	--	--	--	1
	508	SV RR373	5.64	4.70	5.17	--	--	--	--	1
	578	SV RR374	4.74	4.13	4.44	--	--	--	--	1
	546	SV RR375	5.64	5.25	5.44	--	--	--	--	1
	537	SX Avalanche RR(858)	5.67	5.84	5.75	5.57	5.42	5.38	5.12	3
	548	SX Canyon RR	5.21	5.04	5.12	5.19	4.74	5.26	3.85	4
	535	SX Cruze RR	3.98	3.97	3.98	3.39	NE	2.80	NE	4
	519	SX Marathon RR(856)	5.25	4.43	4.84	4.87	4.87	4.90	4.87	3
	558	SX RR1861	5.07	5.02	5.05	4.90	--	4.75	--	2
	527	SX RR1863	6.45	5.64	6.04	5.92	--	5.80	--	2
	516	SX RR1875	3.38	3.75	3.57	--	--	--	--	1
	520	SX RR1876	3.74	3.96	3.85	--	--	--	--	1
	569	SX RR1877	3.93	4.49	4.21	--	--	--	--	1
	552	SX RR1878	5.21	4.86	5.03	--	--	--	--	1
	524	SX RR1879	4.76	4.52	4.64	--	--	--	--	1
	575	SX Winchester RR	4.62	4.67	4.64	4.38	4.23	4.11	3.95	5
1	1201	FS CK #07 CRY5658RR	2.45	3.26	2.85	2.76	2.73	2.66	2.67	12
1	1202	FS CK #08 HILL4000RR	6.50	6.68	6.59	6.37	6.30	6.15	6.16	11
1	1203	FS CK #09 HILL4010RR	6.63	6.20	6.41	6.42	6.40	6.42	6.35	12
1	1204	FS CK #12 HILL4012RR	6.28	5.49	5.89	6.02	6.00	6.15	5.96	12
1	1205	FS CK #13 HILL4043RR	6.22	6.39	6.31	6.18	6.12	6.05	6.01	11
1	1206	FS CK #17 CRY5765RR	3.90	4.13	4.02	4.06	4.13	4.10	4.26	9
1	1207	FS CK #18 CRY5768RR	4.38	4.36	4.37	4.38	4.29	4.40	4.09	9
1	1208	FS CK #26 BETA87RR6	4.64	5.45	5.05	4.78	4.70	4.51	4.53	8
1	1209	FS CK #28 SES36918RF	5.61	4.48	5.04	5.09	5.14	5.13	5.25	9
1	1210	FS CK #29 CRY5875RR	4.68	4.86	4.77	4.73	4.60	4.68	4.35	10
	1211	FS CHK RES RR #1	2.73	2.73	2.73	2.55	2.62	2.37	2.77	7
	1212	FS CHK SUS RR #2	6.39	6.35	6.37	6.25	6.34	6.12	6.53	7
	1213	FS CHK MOD RR RES #	4.55	4.15	4.35	4.26	4.22	4.17	4.14	11
	1214	FS CHK MOD RR SUS #	4.64	4.59	4.61	4.92	4.88	5.23	4.81	11
	1215	FS CHK RES RR #2	1.97	2.82	2.40	2.22	2.20	2.04	2.15	6
	1216	FS CHK SUS RR #10	5.34	5.06	5.20	5.29	5.23	5.38	5.11	4
	1217	FS CHK SUS RR #10	5.66	5.19	5.43	5.37	5.28	5.32	5.11	4
	1218	FS CHK SUS RR #11	5.74	5.48	5.61	5.75	5.51	5.89	5.02	5
		Conventional								
	919	BETA EXP 687	3.65	3.38	3.51	3.46	--	3.41	--	2
	918	BETA EXP 698	2.99	3.13	3.06	2.90	--	2.74	--	2
	905	BETA EXP 747	4.64	4.53	4.58	--	--	--	--	1
	909	BETA EXP 758	3.79	4.03	3.91	--	--	--	--	1
	901	Crystal 620	2.55	3.03	2.79	2.76	--	2.73	--	2
	906	Crystal 622	3.45	3.62	3.53	3.55	--	3.57	--	2
	913	Crystal 735	3.69	3.55	3.62	--	--	--	--	1
	910	Crystal 737	3.79	3.25	3.52	--	--	--	--	1
	902	Crystal R761	3.18	3.28	3.23	3.24	--	3.25	--	11

	914	Hilleshög 3035Rz	3.76	3.63		3.70	3.67	--	3.65	--	13
	917	Hilleshög 9891Rz	3.71	3.60		3.66	3.71	--	3.76	--	2
	904	Maribo MA615Rz	4.93	4.52		4.72	4.92	--	5.11	--	2
	916	Maribo MA720Rz	3.44	3.17		3.31	--	--	--	--	1
	911	Seedex 8869	3.51	3.55		3.53	3.23	--	2.92	--	2
	907	Seedex Deuce	4.53	4.56		4.54	4.61	--	4.68	--	10
	912	Strube 12720	5.49	5.71		5.60	--	--	--	--	1
	908	Strube 13722	6.23	7.02		6.63	--	--	--	--	1
	903	SV 48611	5.84	5.64		5.74	5.49	--	5.24	--	2
	915	SV 48777	3.90	4.03		3.96	--	--	--	--	1
	1201	FS CK #07 CRY5658RR	3.01	2.99		3.00	2.83	2.78	2.66	2.67	12
	1205	FS CK #13 HILL4043RR	6.04	6.17		6.10	6.08	6.05	6.05	6.01	11
	1207	FS CK #18 CRY5768RR	4.23	4.20		4.21	4.30	4.23	4.40	4.09	9
	1209	FS CK #28 SES36918Rf	5.61	5.46		5.54	5.33	5.30	5.13	5.25	9
	1210	FS CK #29 CRY5875RR	4.46	4.53		4.50	4.59	4.51	4.68	4.35	10
10		Check Mean	4.86	4.75		4.81					
		Trial Mean	4.06	4.14		4.10					
		Coeff. of Var. (%)	12.67	13.16							
		Mean LSD (0.05)	0.70	0.70							
		Mean LSD (0.01)	0.93	0.93							
		Sig Mrk	**	**							
		Adj Factor	0.9346	0.9505							
		@ Adjustment is based upon check varieties.									
		+ Average rating based upon multiple rating dates. Lower numbers indicate better tolerance (1=Ex, 9=Poor).									
		NE indicates variety was not evaluated in disease nursery.									
									Created 11/3/2017		

Table 34. Herbicides and Fungicides Applied to ACSC Official Trials						
Location	Herbicide/Insecticide			Fungicide		
	Herbicide & Rate	Spray Dates	Method	Fungicide Used	Spray Dates	Method
Casselton	Conventional	5/15,5/24,6/5	Ground	Quadris	5/31,6/20	Ground
				CR.1/CR.2/CR.3/CR.4	7/10,7/20,8/7,8/18	Ground
Felton	RU1	6/5	Ground	Quadris	5/19,6/8	Ground
	RU2	6/22	Ground	CR.1/CR.2/CR.3/CR.4	7/10,7/20,8/1,8/21	Ground
Georgetown	RU1	6/5	Ground	Quadris	6/1,6/23	Ground
	RU2	6/22	Ground	CR.2/CR.3/CR.4	7/14,7/25,8/15,8/21	Ground
Hendrum	RU1	5/15, 5/26	Ground	Quadris	5/19,6/8	Ground
	RU2	6/26	Ground	CR.1/CR.2/CR.3/CR.4	7/10,7/20,8/1,8/18	Ground
	Conventional	5/15,5/26,6/5	Ground			
Hillsboro	RU1	6/1	Ground	Quadris	5/19,6/8	Ground
	RU2	6/20	Ground	CR.1/CR.2/CR.3/CR.4	7/10,7/20,8/7,8/18	Ground
Climax	RU1	6/5	Ground	Quadris	6/1,6/20	Ground
	RU2	6/22	Ground	CR.1/CR.2/CR.3/CR.4	7/14,7/25,8/15,8/21	Ground
Grand Forks + #	RU1	5/24	Ground	Quadris	5/31,6/22	Ground
	RU2	6/12,7/7	Ground	CR.1/CR.2/CR.3	7/14,7/25,8/15	Ground
	Conventional	5/15,5/24,6/5	Ground			
Scandia	RU1	6/5	Ground	Quadris	5/24,6/12	Ground
	RU2	6/22	Ground	CR.1/CR.2/CR.3	7/14,7/25,8/15	Ground
	Conventional	5/15,5/24,6/5	Ground			
Stephen	RU1	5/26	Ground	Quadris	5/24,6/12	Ground
	RU2	6/12	Ground	CR.1/CR.2/CR.3	7/14,7/26,8/17	Ground
St. Thomas+#	RU1	6/1	Ground	Quadris	6/5,6/19	Ground
	RU2	6/20	Ground	CR.1/CR.2/CR.3	7/19,7/26,8/17	Ground
	Conventional	5/23,5/31,6/8	Ground			
Humboldt	Conventional	5/23,5/31,6/8	Ground	Quadris	6/6,6/16	Ground
Bathgate#	RU1	6/1	Ground	CR.1/CR.2/CR.3	7/18,7/26,8/17	Ground
	RU2	6/20	Ground	Quadris	6/5,6/16	Ground
				CR.1/CR.2/CR.3	7/18,7/26,8/17	Ground
Ground applications complete by Technical Service personnel from ACSC.				Quadris=first application on 2 leaf beet, second on 4-8 leaf beet.		
RU1 = Roundup Powermax (32 oz./A), Event (1 gal./100 gal water).				CR.1=Insire XT + Penncozeb		
RU1, *- Early application of 22oz to control cover crop.				CR.2=Agritin + Incognito		
RU2 = Roundup Powermax (22 oz./A), Event (1 gal./100 gal water).				CR.3=Penncozeb		
				CR.4=Headline + Agritin		
+ Counter 20G applied at 9.0 lbs./A at Grand Forks & St Thomas. Thimet applied at Grand Forks & St Thomas near peak fly in early June.						
# Warhawk 4E applied near peak root maggot fly in early June.						